### DEVELOPMENT AND VERIFICATION OF A KINETIC MODEL OF POLLUTANT DESORPTION FROM DREDGE SEDIMENT AND SEWAGE SLUDGE

### FINAL REPORT

by

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### **OVERVIEW**

This final report encompasses a two phase research effort. The initial direction was toward the development and verification of a kinetic model of heavy metal desorption from dredge sediment and sewage sludge. The second phase used the insights and results gained from that effort to begin an investigation of the toxicity of cadmium in sediments. This change of direction was agreed upon by both the Manhattan College and EPA Narragansett Laboratory researchers and was reflected in the renewal application for the second year of funding. The report is divided into distinct parts, reflecting each topic.

The first part reports on the role of solid phase sulfide in determining the toxicity of cadmium in sediments. For marine sediments the importance of sulfide and the possibility of the formation of insoluble metal sulfides has often been pointed out. We have shown that acid volatile sulfide - solid phase amorphous FeS(s) and MnS(s) which are soluble in cold acid - is the dominant sediment property that controls the cadmium binding capacity of marine sediments and provides the proper normalization for establishing the toxicity of cadmium in a variety of marine sediments. This is a major finding of our research project. The results are presented as a paper which is being submitted for publication. An additional Appendix II is included which presents all the experimental data in tabular form.

The second part of this report presents the results of the development of the methodology that was needed for preforming the experiments and measurements to assess the toxicity of cadmium in sediments. The experimental calibration of the cadmium electrode and the design and testing of a diffusional sampler is presented. Also a validation experiment for the Acid Volatile Sulfide extraction method is presented.

The third part of this report presents the results of the sludge and sediment desorption experiments. These data confirm that the original model proposed for this reaction was correct. A three phase model is appropriate: A reversibly sorbed component; a metal sulfide component that is released via oxidation; and a refractory component that is not released within the time scale of the experiment.

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### TOXICITY OF CADMIUM IN SEDIMENTS: THE ROLE OF ACID VOLATILE SULFIDE

by

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### **INTRODUCTION**

The toxicity of chemicals in sediments is strongly influenced by the extent to which the chemical binds to the sediment. This modifies the chemical potential to which the organisms are subjected. As a consequence different sediment types will exhibit different degrees of toxicity for the same total quantity of chemical. These differences have been reconciled by relating organism response to the chemical concentration in the interstitial water of the sediments [see the review in EPA, 1989]. As a consequence the relevant sediment properties are those which influence the distribution of chemical between the solid and aqueous phases.

The varying toxicity of non-ionic organic chemicals in different sediments has been found to be primarily determined by the organic carbon content of the sediments [EPA, 1989]. The purpose of this paper is to establish the importance of another sediment phase: the acid volatile sulfide phase - the sediment sulfides that are soluble in cold acid - in determining the toxicity of cadmium in sediments. By implication, this phase is likely to be important for all metals which form insoluble sulfides. Most freshwater and marine sediments - completely aerobic sediments are the exception - contain sufficient acid volatile sulfide for this phase to be the predominant determinant of toxicity.

### METHODS AND MATERIALS

### A. Exposure System Design

Sediment dwelling amphipods were exposed for 10 days to control and cadmium-spiked sediments in a 900 mL flow-through chamber with 200 mL of sediment (3.5 cm depth) and 600 mL of overlying seawater. Lighting was continuous to inhibit the amphipods' swimming behavior. Filtered air and seawater flow (10 volume replacements/day) ensured acceptable dissolved oxygen concentrations and cadmium free overlying water.

A diffusion sampler ("peeper") [Hesslein, 1976; Carignan et al., 1984; 1985], designed to fit within the exposure chamber and sample the interstitial and overlying water concentrations, was constructed of Plexiglas G grade unshrunk cast acrylic sheet:  $6 \times 3 \times 2$  in. deep with 6 rows of 3 3/4 in. diameter 1 1/2 in. deep holes, each of which has a volume of about 5 mL. The open side of the

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peeper is covered by a sheet of 1 micron polycarbonate membrane (Nucleopore), followed by a 30 mil low density polyethylene gasket and a 1/2 inch Plexiglas cover plate, both of which have the same hole pattern as the body and secured with PVC-1 cap screws and nuts. Equilibration time was measured to be less than one day.

### **B.** Organism Collection and Holding

Ampelisca abdita were collected from tidal flats in the Pettaquamscutt (Narrow) River, a small estuary flowing into Narragansett Bay, RI, transferred to the laboratory within one half hour, and sieved through a 0.5mm mesh screen. Ampelisca were collected with a dip net after flotation on the air/water interface. Rhepoxynius hudsoni were collected in shallow water at Ninigret Pond, RI. Adult animals were sieved from the sediment through a 1 mm mesh screen in the field, transported to the laboratory within an hour, sieved again and transferred to holding containers. The amphipods were maintained in presieved uncontaminated collection site sediment and flowing filtered seawater, and acclimated to the assay temperature at the rate of 1 to 3°C per day. During acclimation, the Ampelisca were fed, ad libitum, the laboratory cultured diatom Phaeodactylum tricornutum. Rhepoxynius were not fed.

### C. Sediment Acid Volatile Sulfide

The principal property of concern of the sediments used in these experiments was the acid volatile sulfide (AVS) concentration. It is the solid phase sulfide in the sediment that is soluble in cold acid. The measurement technique is to convert the sulfides to  $H_2S(aq)$ , purge it with a gas, and trap it [see Morse et al., 1987 for a review]. A 500 mL Erlenmeyer flask reaction vessel fitted with a three-hole stopper is followed by three sequentially connected 250 mL Erlenmeyer flask trapping vessels. The first is a chloride trap with 200 mL of pH 4 buffer (0.05M potassium hydrogen phthlate) to prevent chloride carry over. The second and third traps contain 200 mL of a 0.1M silver nitrate solution for trapping H<sub>2</sub>S. The four flasks are connected with airtight appropriately shaped glass and Tygon tubing.

A nitrogen gas flow allows continuous purging of the system. In order to prevent oxidation the gas flows through an oxygen-scrubbing system consisting of a vanadous chloride solution in the first scrubbing tower and the matrix of the analyte (seawater) in the second tower. Vanadous chloride is prepared using four grams of ammonium metavanadate boiled with 50 mL of concentrated hydrochloric acid and diluted to 500 mL. Amalgamated zinc, prepared by taking about 15 grams of zinc, covering it with deionized water and adding 3 drops of concentrated hydrochloric acid before adding a small amount of mercury to complete the amalgamation, is then added to the vanadous chloride solution.

The sediment sample (10-15 grams of wet sediment) or standard to be analyzed is placed in the reaction vessel after the entire system has been purged with nitrogen for about an hour. The system is again purged for 5-10 minutes, and deaerated 6M hydrochloric acid is added from a thistle tube to achieve a final concentration in the vessel of 0.5M. The system is run at room temperature for one hour which has been found to be sufficient to complete the extraction. The nitrogen gas flows at a bubble rate of about four per second. The sample vessel is swirled every five or ten minutes. At completion all hydrogen sulfide produced has been converted to silver sulfide in the first silver nitrate trap and no precipitate is found the second trap. The suspension in the first silver nitrate trap is passed through a 1.2 micron GF fiber filter, dried at 102°C, and weighed.

Standards prepared from appropriate quantities of iron(II) sulfate and sodium sulfide (the latter being added from a solution standardized against lead perchlorate), typically gave yields of 95-103%. Silver sulfide precipitates were usually in the range 20-30 mg. When a blank was run (sample without acid), about 0.9 mg silver sulfide was obtained. When the acid was run without a sample, about 0.6 mg silver chloride was obtained. This corresponds to a detection limit of ~ 0.5  $\mu$ mol/g.

### **D. Sediment Characterization and Spiking Procedure**

Sediments of three different acid-volatile sulfide concentrations were used in the toxicity tests. The LI Sound sediment, with a high AVS concentration, was collected from an uncontaminated site in central Long Island Sound (40°7.95'N and 72°52.7'W) with a Smith-MacIntyre grab sampler, returned to the laboratory, press sieved wet through a 2 mm mesh stainless steel screen, homogenized, and stored at 4°C. *A. abdita* has been tested many times in this sediment and both its survival and reproduction have been good (Scott and Redmond, in press). The Ninigret Pond

sediment was a low AVS sand collected from the *Rhepoxynius* collection site. The upper few inches of sediment were collected with a shovel, returned to the laboratory, sieved wet through a 2 mm stainless steel screen, rinsed several times to remove high-organic fine particles, homogenized, and stored at 4°C. The third sediment was a 50/50 (volume) mixture of LI Sound and Ninigret Pond sediments.

Sediments were spiked by adding 1000 mL of wet sediment to 1500 mL of 20°C filtered seawater into which a weighted amount of cadmium chloride had been dissolved. An additional 500 mL of filtered seawater was used to rinse the sediment container. The mixture was stirred with a nylon spatula, capped and placed on a paint shaker for 5 minutes to ensure complete mixing, and held at ambient temperature (~15°C) water bath for 7 days to ensure equilibrium of the cadmium and sediment. A thin layer of cadmium sulfide precipitate that had formed on the surface of the sediment was removed, the test sediments were then homogenized, and 200 mL were transferred to each of three replicate exposure containers. For the experiments with peepers, they were inserted at this time. Exposure containers were placed in the water bath with air and seawater delivery.

### **E.** Toxicity Experiment

The amphipods were sieved from holding containers through a 0.5 mm stainless steel screen and distributed sequentially into 100 mL plastic beakers. After sorting and eliminating dead or outsized animals, the beakers were randomized, air delivery in the exposure system was halted, and one beaker of amphipods was added to the two replicate exposure containers in each treatment. *Rhepoxynius* were added to the Ninigret Pond treatments, and *Ampelisca* to the LI Sound and mixture treatments. The third replicate exposure container in each treatment received no amphipods and was used as a chemical control. Salinity and temperature of the overlying seawater remained relatively constant at  $20.7 \pm 0.3^{\circ}$ C (n=11) and  $30.3 \pm 0.5\%$  (n=11) during the 10 day exposure period.

After termination the contents of each exposure container were sieved through a 0.5 mm screen. For *Ampelisca*, material retained on the sieve was preserved in 5% buffered formalin with

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Rose Bengal stain for later sorting. For *Rhepoxynius*, material retained on the sieve was examined immediately after sieving. In both cases, recovered animals were counted, and any missing individuals were counted as mortalities. The peeper interstitial and overlying water cadmium concentrations were determined as was the AVS and solid phase cadmium in the chemical control vessels.

### F. Cadmium Determinations and Titrations

The cadmium ion concentration in both the peeper samples and the titrations described below was measured as  $Cd^{2+}$  activity using an Orion 94-48 cadmium ion selective electrode and a double junction reference electrode (Orion 90-02). The electrode was standardized with a serial dilution of a 1 g/L cadmium solution that was also used as the titrant. Sediment cadmium was determined using a cold concentrated nitric acid (16M, 5mL) digestion of 10mL wet sediment followed by a peroxide oxidation (10mL 30%) and evaporation to dryness. The residue is reconstitute to 20mL using 0.1M nitric acid and the cadmium measured using an AA.

Cadmium titrations of FeS suspensions (prepared in the same manner as the AVS standards) and sediments were performed using sample sizes of 5 to 10 gm dry wt. added to 50 mL seawater which was constantly stirred. Cadmium chloride was added and dissolved cadmium was monitored using the electrode. Anaerobic conditions were maintained using a nitrogen atmosphere provided by a glove box or by constantly bubbling nitrogen through the covered titration vessel. In the sediment titrations where electrode response was slow, a uniform differential response-time procedure was employed to obtain consistent voltage readings.

### EXPERIMENTAL RESULTS AND INTERSTITIAL WATER CORRELATIONS

### A. Dry weight normalization

The toxicity of cadmium to *Rhepoxynius hudsoni* in Ninigret Pond sediment; and to *Ampe-lisca* in Long Island Sound sediment and an equal parts mixture of the two sediments, is shown in Fig.1. The curves are log-logistic concentration response functions with the same slope parameter (Table 1). The LC50s range from 318 µmol/gm to 3200 µmol/gm on a sediment dry weight basis. As shown below these two organisms have virtually the same LC50s in water only exposures: 0.012

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mg Cd<sup>2+</sup>/L (0.23 mg Cd/L) and 0.017 mg Cd<sup>2+</sup>/L (0.34 mg Cd/L) for *Rhepoxynius* and *Ampelisca* respectively. The concentrations are for free cadmium, Cd<sup>2+</sup>, and total dissolved cadmium, Cd, respectively. Hence the differences in the cadmium toxicity are likely to be attributable to varying sediment properties. In addition Swartz et al., (1985) reported the *Rhepoxynius abronius* cadmium LC50 for a Yaquina Bay sediment to be 25  $\mu$ mol/gm. Thus a factor of ten separates each of the LC50s for these three marine sediments: Long Island Sound (3200  $\mu$ mol/gm), Ninigret Pond (318  $\mu$ mol/gm) and Yaquina Bay (25  $\mu$ mol/gm). An explanation for the over two order of magnitude variation in LC50s would surely be useful.

### **B.** Correlation to Interstitial Water concentration

The correlation between organism toxicity and interstitial water concentration for sediments with different dry weight sediment toxicity has been reported (Adams et al., 1985; Swartz et al., 1985; Kemp and Swartz, 1986). In addition the evidence suggests [Borgmann, 1983] that biological response correlates to chemical activity, in particular to the divalent metal activity,  $\{Me^{2+}\}$  [Sunda and Guillard, 1976; Sunda et al., 1978; Zamuda and Sunda, 1982]. The claim is not that the only bioavailable form of the metal is  $Me^{2+}$  - for example MeOH<sup>+</sup> may also bioavailable - but that the DOC or other ligand complexed fractions are not bioavailable.

These two hypotheses are examined in Fig. 2a, a comparison of the observed mortality to the observed interstitial water cadmium activity, measured with the specific ion electrode, for the three sediments in Fig. 1. The concentration response curves for *Ampelisca* and *Rhepoxynius* in water only exposures are nearly identical. The interstitial water concentration data from the sediment exposures are somewhat scattered. However the grouped data, presented in Fig. 2b as medians (50<sup>th</sup> percentile) and interquartile ranges (25<sup>th</sup> to 75<sup>th</sup> percentiles) parallel the water only exposure curve. These results conform to previous observations that the concentration response curves for sediment exposures, which are quite different on a sediment cadmium dry weight basis (Fig. 1), are quite comparable on an interstitial water basis. Table 1 presents the results.

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### C. Sediment Cadmium vs. Interstitial Water

The prediction of the toxicity of cadmium in sediments requires that the relationship between sediment cadmium concentration and interstitial water concentration be established. A plot of solid phase versus aqueous phase cadmium concentrations - which is regularly used for the analysis of sorption data - is shown in Fig. 3.

The data can be envisioned as a titration in which cadmium is added incrementally to the sediment and the resulting aqueous and solid phase cadmium distribution is measured. Initially the solid phase concentration increases but the aqueous phase concentration remains below the detection limit of the cadmium electrode. Then a critical sediment concentration is reached at which point the aqueous concentration increases sharply - in the region marked "transition" in Fig. 3. Note that the increase is over two orders of magnitude in aqueous concentration while the sediment concentration remains nearly constant. As more cadmium is added, the data then appear to follow a linear trend which is characteristic of a sorption reaction.

It is apparent that the critical part of the relationship between solid and aqueous phase cadmium is the onset of the transition region. There is a sudden increase in interstitial water cadmium activity (mg Cd<sup>2+</sup>/L) and total dissolved concentration (mg Cd/L) from nonlethal levels below 0.001 mg Cd<sup>2+</sup>/L (0.02 mg Cd/L), passing the water only LC50: 0.015 mg Cd<sup>2+</sup>/L (0.30 mg Cd/L) for *Rhepoxynius* and *Ampelisca*, to concentrations in excess of 0.1 mg Cd<sup>2+</sup>/L (2.0 mg Cd/L). This marks the transition between nontoxic and toxic sediments. The solid phase aqueous phase relationship at the lower sediment cadmium concentrations is unclear since the aqueous concentrations are below detection. However the data do not appear to conform to a straight line sorption isotherm that would be inferred by extrapolation from the high concentration data since detectable dissolved concentrations would have been present. The more likely possibility is that a precipitation reaction is maintaining the aqueous phase concentration at below detectable values in the region of low sediment concentrations. Since these are marine sediments the possibility of the formation of a cadmium sulfide precipitate is suggested.

### METAL SULFIDES AND CADMIUM TITRATIONS

The importance of sulfide in the control of interstitial water metal concentrations of marine sediments is well documented (Boulegue, 1983; Emerson et al., 1983; Davies-Cooley et al., 1985; Morse et al., 1987). Metal sulfides are very insoluble and the equilibrium interstitial water metal concentrations in their presence are small. It is possible that the interstitial water sulfide concentration in the sediment samples used for these toxicity tests was initially high enough that so that as cadmium was added to the sediment, cadmium sulfide was precipitating following the reaction:

$$Cd^{2^{+}} + S^{2^{-}} \to CdS(s) \tag{1}$$

However direct measurements of the interstitial water sulfide activity, {S<sup>2-</sup>}, with a sulfide electrode failed to detect any free sulfide in the unspiked sediments. This was a most puzzling result since it was visually clear that a bright yellow cadmium sulfide precipitate was forming as cadmium was added to the sediment.

The lack of significant quantity of dissolved sulfide in the interstitial water and the evident formation of solid phase cadmium sulfide suggested the following possibility. Most of the sulfide in sediments is in the form of solid phase iron sulfides. Perhaps the source of the sulfide is the *solid phase* sulfide initially present. Then as cadmium is added to the sediment it causes the solid phase iron sulfide to dissolve releasing sulfide which is available for the formation of cadmium sulfide. This possibility is examined below.

### A. Solubility Relationships and Displacement Reactions

The majority of sulfide in sediments is in the form of iron monosulfides (mackinawite and greigite) and iron bisulfide (pyrite) of which the former are most reactive [see the review by Morse et al., 1987]. Iron monosulfide, FeS(s), is in equilibrium with aqueous phase sulfide via the reaction:

$$FeS(s) \leftrightarrow Fe^{2+} + S^{2-} \tag{2}$$

If cadmium is added to the aqueous phase, then the result is:

$$Cd^{2^{+}} + FeS(s) \leftrightarrow Cd^{2^{+}} + Fe^{2^{+}} + S^{2^{-}}$$
 (3)

As the cadmium concentration increases,  $[Cd^{2^+}][S^{2^-}]$  will exceed the solubility product of cadmium sulfide and CdS(s) will start to form. Since cadmium sulfide is more insoluble than iron monosulfide, FeS(s) should start to dissolve in response to the lowered sulfide concentration in the interstitial water. The overall reaction is:

$$Cd^{2^{+}} + FeS(s) \rightarrow CdS(s) + Fe^{2^{+}}$$

$$\tag{4}$$

where the iron in FeS(s) is displaced by cadmium to form soluble iron and solid cadmium sulfide, CdS(s). A theoretical analysis of the Cd(II)-Fe(II)-S(II) system, presented in Appendix I, supports this conclusion. The relevant parameter, which can be termed the metal sulfide solubility parameter for any metal, Me, is  $\alpha_{Me^2}$ .  $K_{MeS}$ . It is the product of  $\alpha_{Me^2} = [\Sigma Me(aq) / [Me^{2+}]$ , the ratio of total dissolved Me to the divalent species concentration; and  $K_{MeS} = [Me^{2+}][S^{2-}]$ , the metal sulfide solubility product. These are given in Table 2. The sulfide solubility parameters, shown in Fig. 4, determines the behavior of [FeS(s)] and and [MeS(s)] as the metal is added to the sediment. For example since the cadmium sulfide solubility parameter is less than the iron sulfide solubility parameter, cadmium will form a sulfide at the expense of the iron sulfide which will dissolve. Note that all the metals examined in Fig. 4 are predicted to dissolve FeS and MnS.

### **B. Experimental Results - FeS**

The calculations presented above reflect the chemical composition expected at thermodynamic equilibrium. However many solid phase reactions are not at equilibrium with respect to either the aqueous phase of other solid phases because of the slow kinetics involved in the necessary transformations. Therefore a direct test of the extent to which this reaction takes place has been performed.

A quantity of freshly precipitated iron sulfide is titrated by adding dissolved cadmium. The resulting aqueous cadmium activity, measured with the cadmium electrode versus the ratio of cadmium added, [Cd]<sub>A</sub>, to the amount of FeS initially present, [FeS(s)]<sub>i</sub>, is shown in Fig. 5. The electrode potentials (left) correspond to a very low cadmium concentration during the initial portion of

the titration. Then a sharp upward inflection occurs near  $[Cd]_A \approx [FeS]_i$  indicating that all the iron sulfide has dissolved to form CdS and any additional cadmium added appears as free cadmium. The plot of dissolved cadmium versus cadmium added (right) illustrates the rapid increase in dissolved cadmium that occurs near  $[Cd]_A / [FeS]_i = 1$ . A similar experiment has been performed for amorphous MnS with comparable results. It is interesting to note that such a replacement reaction was postulated by Pankow (1979) to explain an experimental result using copper and FeS.

These experiments plainly demonstrate that solid phase amorphous iron and manganese sulfide can readily be dissolved by adding cadmium. As a consequence it is a source of available sulfide which must be taken into account in evaluating the relationship between solid phase and aqueous phase cadmium in sediments.

### C. Titration results - Sediments

A similar titration procedure has been used to evaluate the behavior of sediment samples taken from four quite different marine environments: Black Rock Harbor; the Long Island Sound and Ninigret Pond sediments used in the toxicity tests; and the Hudson River. The results are shown in Fig. 6. The binding capacity for cadmium is estimated by extrapolating a straight line fit to the dissolved cadmium data. The equation is:

$$[\Sigma Cd(aq)] = \max\{0, m([Cd]_A - [Cd]_B)\}$$
(5)

where  $[\Sigma Cd(aq)]$  is the total dissolved cadmium,  $[Cd]_A$  is the cadmium added,  $[Cd]_B$  is the bound cadmium, and m is the slope of the straight line. The sediments exhibit quite different binding capacities for cadmium, listed in Table 3, ranging from approximately 1 µmol/gm to more than 100 µmol/gm. The question is whether this binding capacity is explained by the solid phase acid volatile sulfide present in the samples.

### **D.** Correlation to Sediment AVS

Sulfides in sediments can be partitioned into three broad classes which reflect the techniques used for quantification [Berner, 1971; Goldhaber and Kaplan, 1974; Morse et al., 1987]. The most labile fraction, acid volatile sulfide (AVS), is associated with the more soluble iron and manganese

monosulfides. The more resistant sulfide mineral phase, iron pyrite, is not soluble in the cold acid extraction used to measure AVS. Neither is the third compartment, organic sulfide associated with the organic matter in sediments [Landers et al., 1983].

The possibility that acid volatile sulfide is a direct measure of the solid phase sulfide that reacts with cadmium is examined in Fig. 7 (left): a plot of the sediment binding capacity for cadmium versus the measured initial AVS for each sediment. The line of perfect agreement is shown. The sediment cadmium binding capacity appears to be somewhat less than the initial AVS for the sediments tested. However a comparison between the initial AVS of the sediments and that remaining after the cadmium titration is completed, Fig. 7 (right), suggests that some AVS is lost during the titration procedure. It is possible that a portion is oxidized even though deoxygenated N<sub>2</sub> gas is passed through the reactor. Or it is possible that some of the AVS is lost as H<sub>2</sub>S(g) via stripping into the gas stream. In any case the correlation of sediment binding capacity and final AVS is apparent in Fig. 7. This strongly suggests that AVS is the proper quantification of the solid phase sulfides that can be dissolved by cadmium.

### SEDIMENT TOXICITY AND AVS NORMALIZATION

The toxicity experiment illustrated in Fig. 1 was designed to test the utility of AVS as a predictor of the cadmium binding capacity of sediments and therefore a predictor of the concentration of cadmium that would cause sediment toxicity.

### **A. Experimental Results**

Fig. 8 presents the AVS and cadmium concentration data at the start and end of the experiment. The initial concentrations of AVS are averages of multiple measurements from the stock supply. The initial concentrations of cadmium are calculated from the weighted amounts added to the sediments. The final cadmium and AVS concentrations are measured in parallel chemical control vessels. The AVS results are interesting. A constant amount of AVS, ~2  $\mu$ mol/gm, is lost from each vessel. Since each sediment is exposed to aerobic overlying water for the same amount of time it is probable that the loss is via oxidation. The final cadmium concentrations are within 70% of the initial values reflecting the loss during initial preparation and via the flowing overlying water.

The toxicity experimental results are shown in Fig. 9. The sediment cadmium is normalized by the AVS for that sediment. The averages of the initial and final values are used for AVS. The relationship in Fig. 8 is used to estimate the final cadmium concentration from the initial concentration if it was not measured. Note that the increase in mortality occurs at the point where the sediment cadmium begins to exceed the sediment AVS on a molar basis. Total mortality occurs at [Cd]/[AVS] > 3. The LC50 that results from a combined fit of the data is 1.97 µmol Cd /µmol AVS.

The critical point is that the sediment AVS can be used to normalize the sediment cadmium concentration in the same way that sediment organic carbon is used to normalize non-ionic organic chemicals. The reason that both methods work is that they properly account for the chemical activity of the chemical in both the aqueous and sediment phases. Below 1  $\mu$ mol Cd / $\mu$ mol AVS the cadmium is all precipitated as CdS(s) and the activity of Cd is very low. Above 1  $\mu$ mol Cd / $\mu$ mol Cd / $\mu$ mol AVS there exists free cadmium in the interstitial water, sorbed cadmium in the sediment phase, as well as CdS(s). The activity of cadmium in the system is now high enough to cause mortality. This is true for sediments with an appreciable amount of AVS, >1  $\mu$ mol/gm. The reason is that the additional cadmium added in excess of 1  $\mu$ mol Cd/gm/  $\mu$ mol AVS/gm is large enough to exceed the activity of cadmium in the system that causes mortality even in the presence of some sorption phases - see Fig.5.

### IMPLICATIONS FOR METAL TOXICITY IN SEDIMENTS

The first order importance of AVS in determining the toxicity of cadmium in sediments has important implications. These are discussed below.

### A. Application to Other Metals and Mixtures

The other potentially toxic metals all form metal sulfide precipitates that are more insoluble than iron sulfide. Fig. 4 presents the sulfide solubility parameters for divalent metals. The iron and manganese sulfides have  $log(\alpha K_{sp}) > -25$  whereas the remaining sulfides have  $log(\alpha K_{sp}) <$ -25. The implication is that the results found for cadmium are applicable to these other metals as well since, at equilibrium, they can displace iron and manganese sulfide to form a more insoluble sulfide precipitate.

In particular it is likely that the LC50 for any metal is at least  $1 \mu mol/\mu mol AVS$ . Given the high concentrations of AVS in most sediments, the LC50s of these metals are likely to be large concentrations. For a molecular weight range of Ni ~ 50 to Pb ~ 200 gm/mol, the LC50s for an AVS of 1 (10)  $\mu$ mol AVS/gm would range from 50 (500) to 200 (2000)  $\mu$ g/gm.

A additional conjecture is that the molar AVS normalized toxicity of metals is additive. Since all the divalent metals in Fig. 4 have lower sulfide solubility parameters than FeS, they would all exist as metal sulfides if their molar sum is less than the AVS. For this case no metal toxicity would be expected and:

$$\frac{\sum_{i} [Me_{T}]_{i}}{[AVS]} < 1 \tag{6}$$

where [MeT]<sub>i</sub> is the total cold acid extractable metal concentration in the sediment. On the other hand if their molar sum is greater than the AVS concentration then a portion of the metals with the lowest sulfide solubility parameters would exist as free metal and presumably exert a toxicity. For this case the following would be true:

$$\frac{\sum_{i} [Me_{T}]_{i}}{[AVS]} > 1 \tag{7}$$

But these two equations are precisely the formulas that one would employ to determine the extent of metal toxicity in sediments assuming additive behavior and neglecting the effect of partitioning. Whether the normalized sum is less than or greater than one discriminates between non toxic and toxic sediments. The additivity does not come from the nature of the mechanism that causes toxicity. Rather it results from their equal ability of the metals to form metal sulfides with the same stoichiometric ratio of Me and S.

This discussion is predicated on the assumption that all the metal sulfides behave similarly to cadmium sulfide. Further it has been assumed that only acid soluble metals are reactive enough to affect the free metal activity. At present no experimental data to support either of these conjectures exists so that this discussion purely speculative.

### **B. AVS and Sediment Quality Criteria**

Since AVS can bind cadmium and presumably metals and thereby reduce their toxicity AVS will obviously play a role in the determination of sediment quality criteria for metals. For sediments with very little or no AVS - fully oxidized sediments for example - an AVS normalization would not be appropriate. Rather the partitioning would be controlled by other sediment phases such as iron and manganese oxides and organic carbon (Jenne et al., 1986). An estimate of when partitioning to other phases can be important can be made using the proposed sediment quality criteria formula [EPA, 1989]:

$$r_{SQC} = K_P c_{WQC} \tag{8}$$

where  $r_{SQC}$  is the sediment quality criteria,  $K_p$  is the partition coefficient, and  $c_{WQC}$  is the chronic water quality criteria. For the case where there is only one metal competing for the AVS, the molar equivalent of the AVS would not bioavailable. Therefore it should be added to the allowable concentration so that:

$$[r_{soc}] = [AVS] + K_P[c_{Woc}]$$
<sup>(9)</sup>

where  $[r_{SQC}]$  is the molar sediment quality criteria (µmol/gm),  $K_p$  is the partition coefficient (L/gm), and  $[c_{WQC}]$  is the molar chronic water quality criteria (µmol/L). The range for freshwater acute (chronic) criteria for the metals in Fig. 4 (hardness = 100 mg/L) is 0.01 to 31. (.0001 to 1.6) µmol/L. The marine criteria are 0.01 to 3.8 (.0001 to 0.88) µmol/L [EPA, 1986]. The

importance of partitioning can be judged by comparing the product  $K_P[c_{WQC}]$  to the AVS concentration. Consider an AVS concentration of 1 µmol/gm. If the partition coefficient is  $K_p = 1 L/gm$  then a metal with a criteria concentration of 1 µmol/L would have its sediment quality criteria doubled due to the partitioning. For  $K_p = 10 L/gm$  the criteria concentration at which partitioning doubles the sediment quality criteria drops to 0.1 µmol/L. Hence the effect of partitioning only becomes significant for relatively low AVS concentrations (~ 1 µmol AVS/gm) and for the metals with larger partition coefficients and criteria concentrations. For all cases with only a single metal involved, the minimum molar sediment quality criterion is [AVS].

### **C. AVS in Freshwater Sediments**

Acid volatile sulfide is commonly found in marine sediments. It is produced by the diagenesis of particulate organic carbon, represented as CH<sub>2</sub>O, with sulfate as the electron acceptor [Goldhaber and Kaplan, 1974]:

$$2CH_2 O + SO_4^{2^-} \to 2CO_2 + S^{2^-} + 2H_2 O \tag{10}$$

and the precipitation of iron sulfide [Berner, 1971]:

$$Fe^{2^+} + S^{2^-} \to FeS(s) \tag{11}$$

It might be expected that AVS is significant only in marine sediments since the concentration of sulfate in seawater is  $28 \text{ mM} = 2700 \text{ mg SO}_4/\text{L}$ . By contrast average river water sulfate concentration is  $0.12 \text{ mM} = 11.5 \text{ mg SO}_4/\text{L}$  [Stumm and Morgan, 1981]. However sedimentary organic matter is present in either locale and the sulfate in freshwater may be sufficient to produce a significant quantity of AVS. This is confirmed by the observations reported in Table 4. Surprisingly large values are found for sediments from the Great Lakes, rivers and other freshwater lakes. The magnitudes are nearly 1 µmol/gm to more than 100 µmol/gm. This strongly suggests that the AVS concentration in freshwater sediments must be considered when addressing cadmium and other metal toxicity.

### **D. Vertical and Temporal AVS Profiles**

The normal method for sediment preparation in sediment bioassays is to produce a uniform distribution of chemical and sediment by careful mixing. For these systems the AVS is uniformly distributed and the concentration to be used for normalization is unambiguous.

However the distribution of AVS in intact sediment cores exhibits both vertical and temporal variation over the annual cycle. Table 4 presents a summary of some observations [Aller 1980; Reaves, 1984]. There is a seasonal variation in the surface concentration of AVS at the Long Island Sound NWC station and all stations exhibit a strong vertcal gradient between the surface 1 cm. and the average of the top 10 cm.

This variation in AVS concentration makes it more difficult to decide what AVS concentration should be used in evaluating the potential toxicity of metals in natural sediments. This is in contrast to the distribution of sediment organic carbon which is more spatially uniform and temporally stable. Hence it appears that intact cores should be used for sediment toxicity testing if metal toxicity is suspected. Indigenous predators such as *Nephtys incisa* should be elimated, however, pehaps by asphyxiation [Scott and Redmond, in press].

### E. Sediment sampling and interstitial water generation

Ferrous sulfide oxidizes very rapidly in aerobic environments. For suspensions, oxidation is virtually complete within a few hours [Nelson, 1978]. We also have noted a decline in AVS for sediments that are held for a long period or are exposed to air. It is clear, therefore, that care should be taken to keep sediments anaerobic before AVS measurements or toxicity testing.

The use of elutrates as a surrogate for interstitial water is also suspect since oxidation of metal sulfides and release of soluble metals can occur. Procedures for producing large volumes of "pore" water by equilibrating suspensions of sediments must be checked for the extent of AVS oxidation that occurs.

### CONCLUSIONS

It has been shown that AVS is the proper normalization parameter for cadmium toxicity in sediments. The observed amphipod mortality versus normalized cadmium concentration, [Cd]/[AVS], is the same for sediments with over an order of magnitude difference in dry weight normalized cadmium LC50s. The correlation between mortality and interstitial water metal activity has also been confirmed. Although the fact that metals can form insoluble sulfides is well known, it apparently has not been recognized that FeS and MnS, quantified as AVS, is a reactive pool of *solid phase* sulfide that is available to bind with metals which have sulfide solubility parameters smaller than FeS.

Titrations of FeS and MnS with cadmium demonstrate that the displacement reaction, Eq. 4. does occur. Further, titrations of sediments with cadmium indicates that an abrupt increase of dissolved cadmium occurs when the added cadmium exceeds the measured AVS. However, these data are not as certain since AVS appears to be lost during the titration and the relationship is only approximate (Fig. 7). Nevertheless, the AVS normalized toxicity data (Fig. 9) does demonstrate that the normalization is quantitative.

Surprisingly, the AVS of freshwater sediments is in the same range as marine sediments. Therefore, AVS should also be the proper normalization for these sediments. The other sorption phases are expected to be important for low AVS sediments and for metals with large partition coefficients and water effect concentrations.

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### **APPENDIX I**

### Solubility Relationships for Metal Sulfides

The behavior of iron sulfide during a titration with cadmium can be analyzed using a simplified equilibrium model of the Cd(II)-Fe(II)-S(II) system. The mass action laws for the sulfide solubilities are:

$$\gamma_{cd^{2*}}[Cd^{2*}]\gamma_{S^{2-}}[S^{2-}] = K_{cdS}$$
(12)

$$\gamma_{Fe^{2*}}[Fe^{2*}]\gamma_{S^{2-}}[S^{2-}] = K_{FeS}$$
(13)

where  $[Cd^{2+}]$ ,  $[Fe^{2+}]$ , and  $[S^{2-}]$  are the molar concentrations;  $\gamma_{cd^{2+}}, \gamma_{Fe^{2+}}$ , and  $\gamma_{S^{2-}}$  are the activity coefficients; and  $K_{FeS}$  and  $K_{cdS}$  are the sulfide solubility products. The mass balance equations for total cadmium, iron(II), and sulfide are:

$$\alpha_{Cd^{2^{*}}}[Cd^{2^{*}}] + [CdS(s)] = [Cd]_{A}$$
(14)

$$\alpha_{Fe^{2^{*}}}[Fe^{2^{*}}] + [FeS(s)] = [FeS(s)]_{i}$$
(15)

$$\alpha_{s^{2}}[S^{2}] + [CdS(s)] + [FeS(s)] = [FeS(s)]_{i}$$
(16)

where  $\alpha_{Cd^{2^{+}}} = [\Sigma Cd(aq)]/[Cd^{2^{+}}], \alpha_{Fe^{2^{+}}} = [\Sigma Fe(aq)]/[Fe^{2^{+}}], \text{ and } \alpha_{S^{2^{-}}} = [\Sigma S(aq)]/[S^{2^{-}}]$ 

are the ratios of the total dissolved Cd, Fe(II), and S(II) to the divalent species concentrations, respectively. [CdS(s)] and [FeS(s)] are the concentration of solid phase cadmium and iron sulfide;  $[FeS]_i$  is the initial iron sulfide in the sediment, and  $[Cd]_A$  is the added cadmium.

The solution of these equations begins with substituting Eqs.(14) and (15) into Eq.(16). Noting that  $\alpha_{s^2}[S^2] = [\Sigma S(\alpha q)] \ll [Cd]_A$ , which states that the total dissolved sulfide in the interstitial water is much less than the cadmium added, it follows that:

$$\gamma_{S^{2^{-}}}[S^{2^{-}}] \approx \frac{\alpha_{Fe^{2^{+}}} K_{FeS} / \gamma_{Fe^{2^{+}}} + \alpha_{Cd^{2^{+}}} K_{CdS} / \gamma_{Cd^{2^{+}}}}{[Cd]_{A}}$$
(17)

Then substituting Eqs. (12), (13) and (17) into Eqs.(14) and (15) yields the concentrations of solid phase sulfides:

$$[CdS(s)] \approx [Cd]_{A} \left( 1 - \frac{\alpha_{Cd^{2*}} K_{CdS}}{\alpha_{Cd^{2*}} K_{CdS} + \alpha_{Fe^{2*}} K_{FeS}} \right)$$
(18)

$$[FeS(s)] \approx [FeS]_i - \frac{\alpha_{Fe^{2*}} K_{FeS}}{\alpha_{Cd^{2*}} K_{CdS} + \alpha_{Fe^{2*}} K_{FeS}} [Cd]_A$$
(19)

where it has been assumed that the activity coefficients for  $Cd^{2+}$  and  $Fe^{2+}$  are equal,  $\gamma_{cd^{2+}} \approx \gamma_{Fe^{2+}}$ , since they are both divalent cations. The relative magnitudes of  $\alpha_{Fe^{2+}}K_{FeS}$  and  $\alpha_{cd^{2+}}K_{cdS}$  determines the behavior of [FeS(s)] and [CdS(s)] as cadmium is added to the sediment. For this reason they are termed sulfide solubility parameters. Table 2 presents reported values. Since the cadmium solubility parameter is much less than the iron sulfide solubility parameter, i.e.,  $\alpha_{cd^{2+}}K_{cdS} \ll \alpha_{Fe^{2+}}K_{FeS}$  Eqs. (18) and (19) become:

$$[CdS(s)] \approx Cd_A \tag{20}$$

and:

$$[FeS(s)] \approx [FeS]_i - Cd_A \tag{21}$$

Hence as cadmium is added to this system cadmium sulfide forms at the expense of iron sulfide. The overall reaction is:

$$Cd^{2+} + FeS(s) \rightarrow CdS(s) + Fe^{2+}$$
(22)

Note that if  $\alpha_{Cd^{2*}}K_{CdS} \gg \alpha_{Fe^{2*}}K_{FeS}$  then  $[FeS(s)] \approx [FeS]_i$ ;  $[CdS(s)] \approx 0$  and no cadmium sulfide would form.

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Experiment	R <sub>0</sub> (%)	β	LC50	Units	Fig. No.
LI Sound(b)	3.32	4.50	3200.	μg Cd/gm	(1)
Mixture(b)	11	*	1130.	98	**
Ninigret Pd(b)	11	**	318.	11	H
Yaquina Bay	0.0	3.34	25.5		
Water Only(c)					
A. abdita	0.0	2.34	0.34	mg Cd/L(d)	(2)
R. hudsoni	0.0	2.33	0.24	U , II	H H
Joint(f)	0.0	2.12	0.29		11
All Sediments	5.88	4.48	1.97	µmol Cd/	(9)
AVS Normalized				µmol AVS	

## TABLE 1Log-Logistic Parameters(a)

(a) Concentration - response formula:

$$R = R_{o} + \frac{100 - R_{o}}{1 + (LC50/c)^{\beta}}$$

R = mortality (%) at concentration c

 $R_0 = \text{control mortality}(\%)$ 

LC50 = concentration for 50% mortality

 $\beta$  = population sensitivity parameter

(b) The three sediments are fit assuming one value of  $R_0$  and  $\beta$ .

(c) Water only exposures - no sediment present in the exposure vessels. Exposure for 96 hrs.

(d) Divide by 20 to obtain mg  $Cd^{2+}/L$ .

(e) Data from Swartz et al., (1985) is fit to the log-logistic function.

(f) Joint fit of the Ampelisca abdita and Rhepoxynius hudsoni water only exposure data.

### TABLE 2

### Metal Sulfide Solubility and

Motol Sulfide	log V		log	~	$\log(\alpha K)$
Metal Sullice	$\log K_{sp,2}$	IUG A sp	iug	u.	log(uK <sub>sp</sub> )
			pH=7.6	pH=8.2	Average
<u> </u>	<u> </u>			<b></b>	
HgS	-38.50	-57.25	15.10	15.10	-42.15
CuS	-22.19	-40.94	0.50	0.92	-40.23
PbS	-14.67	-33.42	1.12	1.32	-32.20
CdS	-14.10	-32.85	1.50	1.50	-31.35
ZnS	-9.64	-28.39	0.12	0.14	-28.26
NiS	-9.23	-27.98	0.11	0.17	-27.84
FeS	-3.64	-22.39	0.10	0.12	-22.28
FeS(am)	-3.05	-21.80	0.10	0.12	-21.69
MnS	-0.40	-19.15	0.13	0.13	-19.02

### **Ratio of Total Dissolved to Free Cation Metal Concentration**

Solubility products,  $K_{sp,2}$ , for the reaction:  $Me^{2+}$  HS<sup>-</sup>  $\leftrightarrow$  MeS(s) + H<sup>+</sup> for CdS (Greenockite), FeS(amorphous) and Mackinawite, MnS (Alabandite), and NiS (Millerite), from Emerson et al., (1983). Solubility products for CuS (Covellite), HgS (Metacinnabar), PbS (Galena), and ZnS (Wurtzite), and pK<sub>2</sub> = 18.57 for the reaction HS<sup>-</sup>  $\leftrightarrow$  H<sup>+</sup> + S<sup>2-</sup>, from Schoonen and Barnes, (1988). K<sub>sp</sub> is for the reaction: Me<sup>2+</sup> S<sup>2-</sup>  $\leftrightarrow$  MeS(s) is computed from log K<sub>sp,2</sub> and pK<sub>2</sub>. Ratios of total to free metal concentrations:  $\alpha = [\Sigma Me(\alpha q)]/[Me^{2+}]$ , from Byrne et al., (1988) at T = 5°C. log( $\alpha K_{sp}$ ) = log  $\alpha$  + log  $K_{sp}$ . All logs are log<sub>10</sub>.

### TABLE 3

### Initial AVS Cd Binding Sediment **Final AVS** $(\mu mol/g)(a)$ $(\mu mol/g)(b)$ Capacity (µmol/g) Black Rock Harbor 175. 114. Hudson River 12.6 8.58 LI Sound 15.9 4.57 13.9 Mixture 5.45 3.23 ... Ninigret Pond 2.34 0.28 1.12

### **Cadmium Binding Capacity and AVS of Sediments**

(a) Average AVS of repeated measurements of the stock

(b) AVS after the sediment toxicity experiment

### TABLE 4

Location	T°C(a)	AVS	s (μmol/gm)	Reference			
	Depth Interval	(0 - 1 cm)	(0 - 10 cm)				
				·			
	Fres	h Water Sedimen	<u>ts</u>				
Everglades peat basin	-	-	0.31 - 1.3	Altschuler et al., 1983			
Lake Mendota	-	-	8.7 - 112.	Nriagu, 1968			
Lake Ontario	-	11.6	27.1	Nriagu et al., 1976			
Lake Erie	(W)	15.0	7.5	Matisoff et al., 1981			
Marine Sediments							
Long Island Sound				Aller, 1980			
NWC	3.0	0.0	8.35				
NWC	13.2	0.60	10.5				
NWC	19.0	0.097	10.3				
DEEP-1	18.5	0.62	17.4				
FOAM-1	20.0	7.50	13.3				
Sapelo Island				Reaves, 1984			
Mud Flat	(W)	1.88	14.6				
Mud Flat	(S)	3.44	43.2				
Tidal Ck.	(W)	9.69	28.4				
Tidal Ck.	(S)	5.94	31.9				

AVS in Freshwater and Marine Sediments

(a)(W) = Winter; (S) = Summer

### **FIGURE CAPTIONS**

Figure 1. Toxicity test results for sediments from Ninigret Pond (*Ampelisca*) Long Island Sound and the mixture (*Rhepoxynius*). Cadmium concentrations on a sediment dry weight basis.

Figure 2. Mortality versus interstitial water cadmium activity. Water only exposure data for *Ampelisca* and *Rhepoxynius*. The line is a joint fit to both data sets (Table 1). Toxicity test results for the sediments in Fig. 1. Top panel - individual data. Bottom panel - statistical summary of all the sediment interstitial water data

Figure 3. Sediment cadmium versus interstitial water cadmium activity for LI Sound sediment.

Figure 4. Metal sulfide solubility parameters for seawater:  $\alpha_{Me^{2}} K_{MeS}$  for the metals as indicated. pH = 7.2 to 8.2; T = 20°C (Table 2).

Figure 5. Cadmium titrations of amorphous FeS. Abscissa is cadmium added normalized by FeS initially present. Ordinate is cadmium electrode response (left panel) and total dissolved cadmium (right panel).

Figure 6. Cadmium titration of sediments: Black Rock Harbor, Long Island Sound, Hudson River, Ninigret Pond. Cadmium added per unit dry weight of sediment versus total dissolved cadmium.

Figure 7. Sediment binding capacity - from the x axis intercepts of the data in Fig. 6. versus sediment AVS concentration at the start of the titration (left). Initial AVS versus final AVS at the end of the cadmium titration.

Figure 8. AVS and cadmium concentrations for the sediments used in the toxicity test (Fig. 1). Initial and final AVS (left); initial and final cadmium (right).

Figure 9. Mortality versus AVS normalized sediment cadmium for Lond Island Sound, Ninigret Pond, and a 50/50 volume mixture.

# **MORTALITY vs SEDIMENT CADMIUM**



## **DRY WEIGHT NORMALIZATION**

Figure 1. Toxicity test results for sediments from Ninigret Pond (Ampelisca) Long Island Sound and the mixture (Rhepoxynius). Cadmium concentrations on a sediment dry weight basis.

SEDIMENT CADMIUM (ug/gm)


### MORTALITY vs INTERSTITIAL WATER CADMIUM

Figure 2. Mortality versus interstitial water cadmium activity. Water only exposure data for *Ampelisca* and *Rhepoxynius*. The line is a joint fit to both data sets (Table 1). Toxicity test results for the sediments in Fig. 1. Top panel - individual data. Bottom panel - statistical summary of all the sediment interstitial water data

# SEDIMENT vs INTERSTITIAL WATER CADMIUM













 $pH = 7.2 \text{ to } 8.2; T = 20^{\circ}C (Table 2).$ 

CADMIUM TITRATION OF IRON SULFIDE



initially present. Ordinate is cadmium electrode response (left panel) and total dissolved cadmium Figure 5. Cadmium titrations of amorphous FeS. Abscissa is cadmium added normalized by FeS (right panel).

# CADMIUM TITRATION OF SEDIMENTS

# **DRY WEIGHT NORMALIZATION**



Figure 6. Cadmium titration of sediments: Black Rock Harbor, Long Island Sound, Hudson River, Ninigret Pond. Cadmium added per unit dry weight of sediment versus total dissolved cadmium.





ment AVS concentration at the start of the titration (left). Initial AVS versus final AVS at the end Figure 7. Sediment binding capacity - from the x axis intercepts of the data in Fig. 6. versus sedi-

of the cadmium titration.



Figure 8. AVS and cadmium concentrations for the sediments used in the toxicity test (Fig. 1). Initial and final AVS (left); initial and final cadmium (right).

Figure 9. Mortality versus AVS normalized sediment cadmium for Lond Island Sound, Ninigret Pond, and a 50/50 volume mixture.



# SEDIMENT CADMIUM (umol Cd / umol AVS)

(%) ҮТІЛАТЯОМ

# MORTALITY vs SEDIMENT CADMIUM

ACID VOLATILE SULFIDE NORMALIZATION

**APPENDIX II** 

## DATA TABLES

-33- N<sup>N</sup>

### Table 1

# **Final Data for Toxicity Experiments**

Expt.#	Sediment	r (ug/g)	[Cd]/[AVS] (mol/mol)	D (	issolved mg Cd <sup>2+</sup> /L	Cd(a)	Mortalit (%) (b)
				I Chi	4 СШ	Celicit	
1 000	IT Sound	0.0	0.0	0 001	0 001	1	3 000
1 000	HI Sound	106 748	0.064	0.001	0 001	•	3,000
1 000	11	150 995	0.000	0.001	0.001	•	3.000
1 000	Ħ	229,153	0.137	0.001	0.001		3.000
1.000	н	288.237	0.172	0.001	0.001		3,000
1.000	n	789.775	0.472	0.001	0.001		3.000
2.000	11	0.0	0.0	0.001	0.001	•	3.000
2.000	"	789.775	0.472	0.012	0.001		13.000
2.000	tt	2396.442	1.431	0.222	0.074		75.000
2.000	tr	8423.713	5.030	7.210	5.030		100.000
2.000	11	25560.363	15.262	22.890	9.580	•	100.000
2.000	н	89846,995	53.648	268.260	101.380		100.000
3,000		0.0	0.0	0.001	0.001		6.000
3,000	81	726.606	0.434	0.001	0.001		6.000
3.000	91	980.801	0.586	0.001	0.001		9.000
3.000	91	1315.596	0.786	0.001	0.001		9.000
3.000	11	2396.442	1.431	0.006	0.076	•	24.000
3.000	11	3961.468	2.365	1.880	2.280	•	73.000
4.000	11	0.0	0.0	0.001		0.001	1.65
4.000	99	175.533	0.105	0.001		0.001	8.350
4.000	11	544.771	0.325	0.001	•	0.001	16.700
4.000	**	1872.224	1.118	0.001	•	0.001	10.000
4.000	**	5810.512	3.469	4.330	•	1.420	100.000
4.000	**	19969.073	11.924	24.200	•	3.940	88.400
4.000	Mixture	0.000	0.000	0.001	•	0.001	16.700
4.000	79	31.078	0.064	0.001	•	0.001	11.700
4.000	11	196.491	0.403	0.001	•	0.001	23.400
4.000	**	1082.707	2.219	0.001	•	0.024	46.700
4.000	19	2325.815	4.768	1.740	•	1.350	100.000
4.000	**	5443.609	11.159	5.900	•	3.400	85.000
4.000	Ninigret	0.000	0.000	0.000	•	0.000	5.000
4.000	**	17.195	0.117	0.001	•	0.001	12.500
4.000	**	71.788	0.488	0.001	•	0.001	12.500
4.000	11	287.150	1.950	0.001	•	0.001	40.000
4.000	11	638.191	4.334	0.475	•	0.050	95.000
4.000	11	2727.925	18.527	0.146	•	0.090	100.000

(a) Detection limit = 0.001 mg  $Cd^{+2}/L$ (b) Centrifugate from the sediment of the chemical control sediment

Organism	Dissolved Cd (mg Cd <sup>2+</sup> /L)	Mortality (%)	
Ampelisca	0.000	2.000	
<b>1</b> 11	0.007	10.000	
12	0.011	36.000	
"	0.018	50.000	
**	0.031	80.000	
91	0.051	98.000	
Rhepoxynius	0.000	0.000	
1 J 11	0.004	2.500	
11	0.006	17.900	
11	0.010	41.500	
	0.018	74.400	
11	0.028	85,000	
**	0.048	92.500	
"	0.080	92.500	
11	0.128	97.500	

Table 2Toxicity Tests - Water Only Exposure

# Chemistry and Toxicity Data for Sediment Toxicity Tests

Variable Identification a	nd Units for Data Table
EXPT	Experiment number
SED\$	Sediment ID
DEPTH	Depth of peeper sampling. Depth = $0$ corresponds to water only exposures.
CDCL2	Concentration of CdCl <sub>2</sub> in the sediment mixture (g CdCl <sub>2</sub> /L) based on the initial weighed sample.
CDCL2H2O	Concentration of CdCl2 • 2.5H <sub>2</sub> O in the sediment mixture (g CdCl <sub>2</sub> • 2.5H <sub>2</sub> O/L) based on the initial weighed sample.
СТ	Concentration of Cd in the sediment mixture (g Cd/L) based on the initial weighed sample.
CT_OBS	Observed concentration of Cd in the sediment mixture (g Cd/L)
CT_FNL	Final concentration of Cd in the sediment mixture (g Cd/L). CT_FNL = CT_OBS if it was measured. If not then $CT_FNL = 10^{(-0.1568 + 1.028 \log 10(CT))}$ which is the regression of log CT_OBS versus log CT.
Μ	Concentration of sediment solids in the sediment mixture (g/L).
R	Cd concentration on the solids based on CT ( $\mu$ g/gm). R = CT/M
R_OBS	Cd concentration on the solids based on CT_OBS ( $\mu$ g/gm). R = CT_OBS/M
R_FNL	Cd concentration on the solids based on CT_FNL ( $\mu$ g/gm). R = CT_FNL/M
RM	Molar Cd concentration on the solids based on CT_FNL ( $\mu$ mol/gm). R = CT_FNL/M/112.4
CD	Dissolved cadmium activity (mg $Cd^{2+}/L$ ) for the water only experiments (EXPT = 0).
CD_PEEP	Dissolved cadmium activity (mg $Cd^{2+}/L$ ) for the peeper cavities within the sediment. All cavities in the overlying water were below detection.

CD_CENTR	Dissolved cadmium activity (mg $Cd^{2+}/L$ ) for the centrifugate separated from the sediment in the chemical controls at the end of the experiment.
LCD_PEEP	Log <sub>10</sub> (CD_PEEP)
LCD_CENT	Log <sub>10</sub> (CD_CEMTR)
AVS1	Final AVS at the end of the experiment (µmol/gm)
AVS2	Initial AVS at the start of the experiment (µmol/gm)
CD_AVS1	Sediment cadmium concentration normalized by AVS1. CD_AVS1 = R_FNL/AVS1
LCD_AVS1	Log <sub>10</sub> (CD_AVS1)
CD_AVS2	Sediment cadmium concentration normalized by AVS2. CD_AVS2 = R_FNL/AVS2
LCD_AVS2	Log <sub>10</sub> (CD_AVS2)
CD_AVS	Sediment cadmium concentration normalized by AVS. CD_AVS = R_FNL/AVS
LCD_AVS	Log <sub>10</sub> (CD_AVS)
MORT	Organism mortality (%)
SURVIVOR	Organism survival (%). SURVIVOR = 100 - MORT

CT ENL						58,1778	58.1778	82.2922 82.2922	124.8884	124.8884	157.0880	430.4275	430.4275	•	430.4275	430.4275	1306.0612	1500.0012	4590.9235	13930.3976	48966.6120	48966.6120	396.0000	534.5366	534.5366	717.0000	1306-0612	2159,0000	2159.0000	2159.0000	105 87.43	328.4972	1128-9513	5505-7508 12041-3508	00000	31-0000	1080.0000	2320.0000 5430.0000	0,0000	100.0000	400.0000	3800.0000
CT OBS						•		•		•	•			•			•			•		•	396.0000	596-UUUU		717 0000	•	2159,0000	2159.0000	2159-0000	•	• •		•	0.0000	31.0000	1080.0000	2320.0000 5430.0000	0.000	100.0000	400.0000	3800.0000
CT	0000 0	0.1300	0.3600	0.6000	00000	00000	73.9979	103.6857	155.5777	155.5777	194-4722	518.4284	518.4284	00000	518.4284	518.4284	1526-2374	1526-2574 518/ 2878	5184.2838	15262.3741	51842.8384	51842.8384	393.8677	595.8677 640.0350	640.0350	984.0693	1526.2374	4763-0261	2461.6732	2461.6732	00000	398.5816	1324.5172	3985-8156 12245-1718	0.000	95.6596	956.5957	2863.6552 9565.9574	00000	93.8200	312.1200	3121.2002
12420				•	0.0000	0.0000	0.1503	0.2106	0.3160	0.3160	0.3950	1.0530	1.0530	0,000	1,0530	1.0530	3.1000	3.1000	10.5300	31.0000	105.3000	105.3000	0.8000	0.8000	1.3000	2,0000	3.1000	5.1000	5.0000	5.0000	•	•		•								
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	AVS2					-1.2238	-1.2238	-1.0732	-0.8920	-0.7924	-0.3547	-0.3547		-0.3547	0.1274	0.6733	1.1554	1.7013	1.7013	-0.3909	-0.2606	-0.1330	-0.1550 0.1274	0.1274 0.3457	0.3457	0.3457	-1.0078	0.0202	1.0482	-1.2947	0.2473	0.5794 0.9487	-1.1846	-0.5639 0.0381 0.3850 1.0159
	O_AVS2 LCI					0.0597	0.0597	0.0845	0.1282	0.1613	0.4419	0.4419		0.4419	1.3409	4.7135	4. (132	14.3022 50.2736	50.2736	0.4066	0.5488	0.7361	0.7361	1.3409 2.2166	2.2166	2.2166	0.0982	1.0476	11.1736	0.0507	1.7675	3.7968 8.8864	0.0000	0.2729 1.0918 2.4264 10.3717
	CAVS1 CI		•			-1,1654	-1.1654	-1.0148	-0.8337	0.7340	-0.2963	-0.2963		-0.2963	0.1858 0.1858	0.7317	1.2138	1.2138	1.7597	-0.3325	-0.2022	2707.0-	-0.0747 0.1858	0.1858 0.4041	0.4041	0.4041	-0.9494	0.0786	1.1066	-1.0675	0.4745	0.8066	-0.2625	0.3581 0.9602 1.3070 1.9379
	AVS1 LCC		•			0.0683	0.0683	0.0966	0.1467	0.1845	0.5055	0.5055		0.5055	1.5339	5.3917	16.3601	16.3601 57.5072	57.5072	0.4651	0.6278	0.8421	0.8421	1.5339 2.5356	2.5356	2.5356	0.1124	1.1983	12.7814	0.0856	2.9822	6.4063 14.9940	0.5464	2.2810 9.1240 20.2781 86.6779
CDFNI	AVS2 CI		•		15.9000	15.9000 15.9000	15.9000	15.9000	15.9000 15.0000	15.9000	15.9000	15.9000 15.9000	15.9000	15.9000	15.9000	15.9000	15.9000	15.9000	15.9000 15.9000	15.9000	15.9000	15.9000	15.900	15.9000	15.9000	15.9000	15.9000	15.9000	15.9000	5.4500	5.4500	5.4500	2.3400	2.3400 2.3400 2.3400
	AVS1		-		13.9000	13.9000 13.9000	13.9000	13.9000	13.9000 13.0000	13.9000	13.9000	13.9000 13.9000	13.9000	13.9000	13.9000 13.9000	13.9000	13.9000	13.9000 13.9000	13.9000	13.9000	13.9000	13.9000	13.9000 13.9000	13.9000	13.9000	13.9000	13.9000	13.9000	13.9000	2.2300	3.2300	3.2300	0.2800	0.2800 0.2800 0.2800 0.2800
	LCD_CENT		•								•••	•			•				•					•			0000	-3.0000	0.5955	0000.5-	-1.6198	0.1303	-3.0000	-3.0000 -3.0000 -1.3010 -1.0458
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CD_AVS		-1.1956 -1.1956 -1.0450 -0.8638 -0.8638	-0.7642 -0.7642 -0.3264 -0.3264 -0.3264	0.1556 0.1556 0.7016 1.1836 1.1836 1.7296		0.3739 0.3739 0.3739 -0.9796 -0.4877 0.0484 0.5403 1.0764	-1.1958 -0.3949 0.6783 1.0476 -0.9326 -0.3369 0.6369 1.2678
CD_AVS L		0.0637 0.0637 0.0902 0.1368 0.1368 0.1368	0.4716	-1.4309 -1.4300 -1.4300 -1.430	0.4339 0.5856 0.5856 0.5855 0.5855 0.7855 1.4309 1.4309 2.3654 2.3654 2.3654	2.3654 2.3654 0.1048 0.3253 1.1179 3.2695 1.179	0.0637 0.4028 11.1591 11.1591 0.4168 0.4178 1.9505 1.9505 18.5266
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# Chemistry Data for Cadmium Titrations of FeS and Sediments

Variable Identification ar	nd Units for Data Table
NUM	Expt number
NAME\$	Sediment name
CAL_CD	Calibration - cadmium concentration (mg/L)
LCAL_CD	" log10 " "
CAL_POT	Calibration - electrode potential mv
ADD_CD	Volume of cadmium stock solution added (mL)
UMCD_UMS	μmol Cd added / μmol AVS
РОТ	Dissolved total cadmium - electrode potential (mv)
LOGCD	Log10 Dissolved Cd - from electrode potential and calibration
PH	pH during the titration
CD	Dissolved total Cadmium - from electrode (mg/L)
Μ	Mass of sediment in the titration (gm)
V	Initial volume of seawater (mL)
CD_MOL	Cd concentration of the titrant (M)
UMCD_G	µmol Cd added / g sediment
UMCDDS_G	µmol dissolved Cd / sediment
CD_TOT	Total Cd added (mg/L)

LOGCD	
POT	-161.0000 -113.2000 -113.2000 -113.2000 -113.2000 -113.2000 -113.2000 -110.2000 -110.2000 -125.2000 -125.2000 -125.2000 -125.2000 -125.2000 -125.2000 -125.2000 -125.2000 -125.2000 -125.2000 -125.2000 -125.2000 -125.2000 -125.2000 -125.2000 -125.2000 -125.2000 -125.2000 -125.2000 -125.2000 -125.2000 -125.2000 -125.2000 -125.2000 -125.2000 -125.2000 -125.2000 -255.2000 -255.2000 -255.2000 -125.2000 -125.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2000 -255.2
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NUM    AMAES    CAL_OD    LOAL_OD    AD_DD      XXX    XXX    XXX    XXX    XXX    XXX    XXX      XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXX    XXXX    XXXXX    XXXXX    XXXXX    XXXXX    XXXXX    XXXXX    XXXXXX    XXXXXX    XXXXXXXXX    XXXXXXX    XXX	UMCD_UMS	0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1530 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.1520 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200 0.15200000000000000000000000000000000000
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NUN    MAGS      CASE    2243      CASE    2244      CASE    2245      CASE    2245      CASE    2244      CASE    2255      CASE    2555      CASE    2556      CASE <th>CAL_CD</th> <th>00000000000000000000000000000000000000</th>	CAL_CD	00000000000000000000000000000000000000
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UMCDDS_G	00000000000000000000000000000000000000
UMCD_G	23333333333333333333333333333333333333
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UMCDDS_G	•	•		•	•	•	•	•					•	•	•		•		•	•		•	•			•	•	•			•		• •			•	0,0000	0.000	0,000	0.0001	0.0001	2000-0	7000 D	0.0004	0.0006	2100.0	0,0102	0.0111	0.0173	0.00	
UMCD_G	•	•	•	•	•	•	•	•				•		•	•			•		•		•	•	•		•		•		•				•			ບູບບບ	2.3739	4.7478	8.3086	8.9021	9.4955	10.4825	11.2760	11.8694	12.4629	12.0004	14.2433	14.8368		
CD_MOL	•	•	•	•	•	•	•	•	• •		• •	•	•	•	•				•	•	•	•	•	•			•	•		• •		•		•	• •		ບຸບບາບ	0,0040	0.0040	0,000.0	0,0040	0.0040	0.0040	0,0040	0.0040	0,0040		0.0040	0,00,0	0	
>	•	•	•	•	•	•	•	•	•	• •		•	•	•	•	• 1		•			•		•	•	• •			•	• •			•		•	•		35,000	35.0000	35.0000	35.0000	35.0000	35.0000	55.0000	35.0000	35.0000	35.0000	20,000	35,0000	35.0000		
× .		•	•	•	•	•	•	•	•	• 1		•			•	•		•		•	•	•	•	•	• •		•	•	• •						•		2,2700	3.3700	3.3700	3.3700	3.3700	3.3700	5.5/00	3.3700	3.3700	5.5/00	2.5/00	3.3700	3.3700	0010-0	
8	0.0646	0.2754	0.5248	0.7762	1.0715	0.000	0.000		7000	0.0013	0.0035	0.0074	0.0195	0.0513		2702-0	0.3311	0.4677	0.7762	1.0965	0,000	0.0043	0.0049	2400.0	2,0893	3.8905	6.1660	16-9824 20 5121	42.6580	57.5440	0000		0,000	0.0001	1000-0	0.0056	0.0120	0.0001	0.0004	0.0011	0.0012	0.0014	0-0020	0.0036	0,0049	0.0102	21 CU . U	0.0891	0.1380	0. 1070	
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	61	62	63	64	65	9 9	, 9,	00	22	25	22	R	74	62	25	20	22	80	81	82	83	<b>7</b> 8	82 202	91 80	28	38	83	50	26	55	ድሪ የ	000	98	8	100	102	103	105	106	108	109	110		11	114	115	116	118	119	2	
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CD_TOT	125 126 128 128 128 128 128 128 128 128 128 128
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UMCD_G	66 66 66 66 66 66 66 66 66 66
CD_MOL	
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8	0.00289 0.00264 0.00264 0.00265 0.00275 0.00275 0.0007 0.000756 00
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**X** (1)

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UMCD_G	2-1-1-000222202020222222222222222222222
CD_MOL	
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HA	88888888888888888888888888888888888888
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CD_TOT	65.795 79.3412 79.3412 79.3412 79.3412 79.2412 79.2505 79.2000 79.2505 79.2505 70.0000 88.3835 70.2505 70.2000 774.0195 88.3835 70.2000 774.0195 88.3835 70.2000 711.2695 88.3835 711.2695 712.2695 712.2
UMCDDS_G	0.0000 0.0113 0.00000 0.00000 0.00000 0.00000 0.00000 0.00000 0.00000 0.00000 0.00000 0.00000 0.00000 0.00000 0.00000 0.00000 0.00000 0.00000 0.00000 0.000000
	22222222222222222222222222222222222222
CD_MOL	0,000000000000000000000000000000000000
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ទ	1.2802 1.2802 1.2802 1.2802 1.2808 1.
Hd	8 5 5 5 5 5 5 5 5 5 5 5 5 5
	00000000000000000000000000000000000000
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### **AVS Determinations**

Variable Identification and Units for Data Table Sediment identifier NAME\$ Date of the experiment DATE\$ Dry weight of sediment extracted DRYWT Weight of Ag<sub>2</sub>S in sulfide trap AG2S UMS\_G µmol/gm AVS  $\mu$ mol/gm AVS - These were extractions after a cadmium titration was UMS\_G1 completed. Ratio of UMS\_G1 to UMS\_G RATIO

		NAME\$	DATE\$	DRYWT	AG2S	UMS_G	UMS_G1	RATIO
CASE	1	BRH	8/5/88	3.9554	0.1998	203.8500	•	-
CASE	ż	BRH	9/22/88	2.1800	0.0789	146.0600	92.4600	0.6330
CASE	3	HR	7/11/88	11.8200	0.0312	10.6500	•	•
CASE	4	HR	7/22/88	5.8500	0.0211	14.5600	•	-
CASE	5	HR1	10/11/88A	10.5300	0.0350	27.9100	•	•
CASE	6	HR1	10/13/88A	7.1300	0.0426	24.1100	•	-
CASE	7	HR1	9/27/88	13.0700	0.0340	23.9900	•	•
CASE	8	HR1	9/27/88B	11.4800	0.0387	24.8800	•	•
CASE	9	LIB	7/11/88	8.9100	0.0339	15.3500	•	•
CASE	10	LIB	7/11/88	8.9200	0.0336	15.2100	•	•
CASE	11	LIB	7/13/88	6.7100	0.0237	14.2500	•	•
CASE	12	LIB	7/13/88	15.9300	0.0503	12.7500	•	•
CASE	13	LIB	7/13/88	15.8000	0.0467	11.9300	•	•
CASE	14	LIB	7/14/88	17.9200	0.0545	12.2700	•	•
CASE	15	LIB	7/15/88	7.9200	0.02/9	14.2200	-	•
CASE	16	LIB	//15/88	6.0500	0.0243	10.2100	•	
CASE	17	LIB	//18/88	13.0800	0.0451	12.0000	•	•
CASE	18	LIB	7/19/88	15.8000	0.0408	10.4200	•	•
CASE	19	LIB	////88	11.5200	0.0400	15.9700	•	•
CASE	20	LIB	//8/88	11.0100	0.0419	12.3500	<b>1</b> 5000	0 3125
CASE	21	LIB	8/1/88	41, 2500	0,0504	14.4000	4.3000	0.5125
CASE	22	LIBI	(/24/88	7 1170	0.0001	12 0200	A 2100	0,4802
CASE	23	LIBI	0/2/00	12 4400	0.0220	7 5000	0.2100	0.4005
CASE	<u> </u>		9/20/00	12.0000	0.0230	11 / 100	-	•
CASE	22	LIBI	7/10/00	10.7000	0.0303	7 1500	•	-
CASE	29	55	7/10/00	40.0500	0.0313	2 0600	-	•
CASE	21	33	7/9/00	41.1000	0.0302	2 1800	-	-
LASE	20	33	9/7/99	11 5200	0.0101	3 1500	2 3700	0 7524
CASE	27	22	8// 199	15 1000	0.0070	2 1100	2 4400	1 1564
LASE	20	55	0/4/00	25 2100	0.0079	3 1000	6.7700	11104
LASE	20	55 601	7/23/00	15 0/00	0 0103	2 6100	•	•
LASE	52	221	7/2//00	13.9400	0.0105	2.0100	•	•

# Comparison of Initial AVS and Cadmium Binding

Variable Identification a	nd Units for Data Table
NAME\$	Sediment identifier
NUM	Expt number
DATE\$	Date of the experiment
AVS	Initial AVS at the start of the titration ( $\mu$ mol/gm)
CD_G	Binding capacity of the sediment (µmol Cd/g sediment

		NAME\$	NUM	DATE\$	AVS	CD_G
CASE	1	BRH	1.0000	8/5/88	203.8500	122.9768
CASE	2	BRH	2.0000	9/22/88		105.8089
CASE	3	HR	1.0000	7/12/88		12.4965
CASE	ž	HR	2,0000	7/22/88	14.5600	6.3564
CASE	Ę	. HD	3 0000	10/03/88		7.9552
CASE	4		1 0000	7/5/88	•	6 1573
CASE	9		2 0000	7/0/00	•	5 5422
CASE	6	LIB	2.0000	7/17/00	41 2500	2.2622
CASE	8	LIB	5.0000	//15/88	14.2500	2.2021
CASE	9	LIB	4.0000	7/13/88	12.7500	7.1214
CASE	10	LIB	5.0000	7/14/88	12.2700	8.1084
CASE	11	LIB	6.0000	8/1/88		3.8762
CASE	12	LIB	7.0000	8/2/88	12,9000	4.7734
CASE	13	I TR	9.0000	8/3/88		6.4904
CASE	16	LID	10,0000	0/10/88	7 5900	1 5547
CASE	15	20	1 0000	7/8/88	2 1800	0 0381
LASE	10	33	2,0000	7/1/00	2.1000	2 0420
CASE	10	33	2.0000	7/1/00	•	2.0020
CASE	17	55	5.0000	//14/00	7'4500	1.012/
CASE	18	SS	4.0000	7/18/88	5.1500	0.7589
CASE	19	SS	5.0000	8/4/88	•	1.1768

# AVS in Long Island Sound Sediments<sup>#</sup>

Variable Identification and Units for Data Table							
ID\$	Sediment identifier						
TABLE\$	Table number in the reference from which the data were taken						
Т	Temperature °C						
DEPTH1	Starting depth of the core slice (cm)						
DEPTH2	Ending depth of the core slice (cm)						
DEPTH	Average depth of the core slice (cm)						
FES	Measured AVS in the slice $(\mu mol/g)$						
#[Aller, 1980]							

FES	7.5000 11.0000 8.1000 12.0000	23.0000 20.0000	13.0000 11.0000 12.0000	11.0000 7.2000 8.4000	6.6000 7.2000	10.0000	0.6000 9.0400	13.7000	9.3600	0.0000	19.0000	15.2000 8.4500 5.7100	4.8800 3.1800	1.7000	16.9000	10.1000	6.8600	4.4300	11.0000 22.7000	30.7000	24.5000 24.6000 15.9000
DEPTH	0.5000 1.5000 3.5000	4.5000 5.5000 6.5000	7.5000 8.5000 9.5000	10.5000 11.5000 12.5000	13-5000	16.5000	21.0000 0.5000 1.5000	2.5000	5-5000 6-5000	0.5000	2.5000	4.5000 5.5000	7.5000 8.5000	9.5000 0.5000	2.5000	5.5000	6.5000 7.5000	1.5000	3.5000	5.5000	7.5000 9.0000 11.0000
DEPTH2	1.0000 3.0000 4.0000	5.0000 7.0000	8.0000 9.0000 10.0000	11.0000 12.0000 13.0000	15.0000	18.0000	24.0000 1.0000 2.0000	7.0000 4.0000	20000 9.00000 2.00000	8.0000 1.0000 0000	3.0000	5.000 6.0000	8.0000 9.0000 9.0000	10.000	3,0000	5.0000	7.0000 8.0000	2.0000	5.0000	2.0000	10.000 12.0000 14.0000
DEPTH1	0.0000 2.0000 3.0000	4.0000 5.0000 6.0000	7.0000 8.0000 9.0000	10.0000 11.0000 12.0000	14.0000	16.0000	0.000 0.0000 1.0000	2-0000 3-0000	6-0000 10000 10000	0000	2.0000	4-0000 5-0000	8,0000	0000	2.0000	5.0000	6.0000 7.0000	1.0000	3.0000	5.0000	7.0000 8.0000 10.0000
L	20.0000 20.0000 20.0000 20.0000	20.0000 20.0000	20.0000 20.0000 20.0000	20.0000 20.0000 20.0000	20.000	20.000	20.0000 13.2000 13.2000	13.2000	13.2000	13.2000 3.0000 3.0000	3.0000 3.0000	3-0000 3-0000	3.0000 3.0000	3.0000 19.0000	19.0000	19.0000	19.0000 19.0000	18.5000	18.5000	18.5000	18.5000 18.5000 18.5000
TABLES	<u> </u>	888	888	8 8 8 7 7 7			81 87 87	87 87	87 87	83 88 88	8888	888	9888 8888 8888	888	888 8	288	89 89	9 9 7 7 7 7 7 7 7 7 7 7 7 7 7 7 7 7 7 7	812 812	812	812 812 812 812 812 812 812 812 812 812
ID\$	FOAM1 FOAM1 FOAM1	FOAM1 FOAM1 FOAM1	FOAM1 FOAM1 FOAM1	FOAM1 FOAM1 FDAM1	FOAM1 FOAM1	FOAM1	FOAM1 NWC-2 NWC-2	NWC-2	NWC-2	NWC-2 NWC-3	NWC-3	NHC-3 NHC-3	NWC-3 NWC-3 NWC-3	NWC-5	NWC-4 NUC-4	NWC-4 NWC-4	NWC-4 NWC-4	DEEP-1	DEEP-1	DEEP-1	DEEP-1 DEEP-1 DEEP-1
	-0104	502	<b>80</b> 0	200 201	1455	2645	2012	1222	5235	28 28 28	30,5	222	462%	38	64 70 70	435 435	4 <u>6</u> ;	0 7 7 7 7 7 7	2620	223	83283
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## TABLE 8

## AVS in Sapelo Island Sediments<sup>#</sup>

Variable Identification and Units for Data Table

STA\$	Sediment identifier
SEASON\$	Table number in the reference from which the data were taken
DEPTH1	Starting depth of the core slice (cm)
DEPTH2	Ending depth of the core slice (cm)
FES_W	Measured AVS in the slice (weight %)
FES	Measured AVS in the slice (µmol/g)

#[Reaves, 1984]

		STA\$	SEASON\$	DEPTH1	DEPTH2	FES_W	FES
CASE	1	MUD	W	0.000	1.000	0.006	1.875
CASE	2	MUD	W	1.000	2.000	0.005	1.563
CASE	3	MUD	W	2.000	3.000	0.012	3.750
CASE	4	MUD	W	5.000	4.000	0.000	20.027
CASE	5	MUU	W	4.000	5.000	0.077	18 125
CASE	9	MUD	W L	5.000	7 000	0.050	10 063
CASE	6	MUD	w	7 000	8 000	0.051	15,938
CASE	õ	MUD	ü	8,000	9,000	0.047	14.688
CASE	10	MUD	ü	9,000	10.000	0.060	18.750
CASE	11	MUD	ü	10.000	11.000	0.069	21.563
CASE	iż	MUD	ÿ	11.000	12.000	0.035	10.938
CASE	13	MUD	Ŵ	12.000	13.000	0.039	12.188
CASE	14	MUD	¥	13.000	14.000	0.046	14.375
CASE	15	MUD	W	14.000	15.000	0.036	11.250
CASE	16	MUD	S	0.000	1.000	0.075	23.438
CASE	17	MUD	S	1.000	2.000	0.058	18.125
CASE	18	MUD	S	2.000	5.000	0.090	28.125
CASE	19	MUD	S	5.000	4.000	0.097	30.313
CASE	20	MUD	S	4.000	5.000	0.120	37.500 71.543
CASE	21	MUD	S	5.000	7 000	0.229	51 543
CASE	22	MUD	5	7 000	8 000	0.153	67 813
LASE	23	MUD	3	8 000	0.000	0 164	51 250
CASE	25	MUD	s c	9,000	10,000	0 193	60.313
CASE	26	MUD	Š	10,000	11,000	0.176	55,000
CASE	27	MUD	š	11.000	12.000	0.222	69.375
CASE	28	MUD	š	12,000	13.000	0.201	62.813
CASE	29	MUD	Š	13.000	14.000	0.222	69.375
CASE	30	MUD	Š	14.000	15,000	0.220	68.750
CASE	31	CK	W	0.000	1.000	0.031	9.688
CASE	32	CK	¥	1.000	2.000	0.088	27.500
CASE	33	CK	W	2.000	3.000	0.077	24.063
CASE	34	CK	W	3.000	4.000	0.073	22.813
CASE	35	CK	W	4.000	5.000	0.103	32.188
CASE	36	CK		5.000	6.000	0.115	35.313
CASE	37	CK	. N	6.000	7.000	0.057	21.100
CASE	38	CK	W	7.000	0.000	0.055	74 250
LASE	29		W	0.000	10 000	0.133	61 563
CASE	40		w U	10 000	11 000	0.133	38 750
CASE	41			11 000	12 000	0 124	38 750
CASE	42		u u	12 000	13 000	0.070	21.875
CASE	44	CK CK	ü	13,000	14,000	0.090	28,125
CASE	25	ČK	ü	14.000	15,000	0.099	30,938
CASE	46	CK	ŝ	0.000	1.000	0.019	5,938
CASE	47	ČK	š	1.000	2.000	0.018	5.625
CASE	48	CK	Š	2.000	3.000	0.039	12.188
CASE	49	CK	S	3.000	4.000	0.075	23,438
CASE	50	CK	S	4.000	5.000	0.115	35.938
CASE	51	CK	S	5.000	6.000	0.196	61.250
CASE	52	CK	S	6.000	7.000	0.155	48.438
CASE	53	CK	S	7.000	8.000	0.144	45.000
CASE	54	CK	S	8.000	9.000	0.135	42.188
CASE	55	CK	S	9.000	10.000	0.125	39.063
CASE	56	CK	S	10.000	11.000	0.103	52.188
CASE	57	CK	S	11.000	12.000	0.102	51.8/5
CASE	20	CK	S	12.000	12.000	0.000	21.000
LASE	59 40		2	12.000	14.000	0.102	31.0/3
LASE	00	LK	2	14.000	12:000	0.105	32.100

## DEVELOPMENT OF EXPERIMENTAL METHODOLOGY

#### A. Measuring Cadmium Activity

The most direct method to measure activity of a metal is to use an ion specific electrode (Orion Model 94-48). Ion selective electrodes measure the activity of the target species only - in this case  $Cd^{2+}$ ). The determination of a reliable standard curve is the first step in the establishment of an analytical method. For the ion selective electrode this involves the measurement of relative potential in conjunction with a double function reference electrode as a function of activity, of the analyte  $(Cd^{2+})$ .

Because  $Cd^{2+}$  forms several stable complexes with Cl<sup>-</sup>, the standardization cannot be done in seawater. A noncomplexing matrix of the same ionic strength as seawater, 0.7 M NaNO<sub>3</sub>, was used. The results are shown in Fig. 1. The linear behavior of the potential versus cadmium activity to 0.1 mg/L with a slope of 28.2 is consistent with the Nernst equation. When a hydrogen carbonate buffer is added to mimic seawater conditions no significant change occurs in the standard curve (Fig. 2). If the 0.7 M NaNO<sub>3</sub> Cd<sup>2+</sup> potentials are compared with those obtained for the same total cadmium in seawater, the Cd<sup>2+</sup> fraction is about 5%, which is consistent with values calculated from simultaneous equilibrium models of cadmium speciation in seawater.

To further investigate the performance of the cadmium electrode, several titrations were performed using ions that are known to complex with  $Cd^{2+}$  and for which the formation constants are known. The results of a hydroxide titration are shown in Fig. 3. The value of  $pK_1 = 9.75$  is consistent with literature values. The results of a chloride titration are shown in Fig. 4. The initial total cadmium concentrations are 10 and 1 mg/L. The value of 26.4 which is obtained for the formation constant of CdCl<sup>+</sup> is consistent with reported values.

## **B. Water Only Exposure - Bioassay Results**

The above experiments demonstrated that the cadmium electrode was indeed measuring the cadmium activity in the systems of interest. During this period water-only bioassays were being performed at the Narraganset EPA Environmental Research Lab (ERL) to determine the total

cadmium (Cd<sub>T</sub>) LC<sub>50</sub> for the organism, <u>Ampelisca abdita</u>, that was to be used in the sediment exposure studies. The results are shown in Fig. 5 for both the definitive and range finding experiments.

Following this preliminary work, total cadmium samples bracketing the LC<sub>50</sub> values of 0.32 - 0.55 mg/L were prepared in seawater. The Cd<sup>2+</sup> concentration was determined by the electrode, while total cadmium was measured by anodic stripping voltametry and graphite furnace atomic absorption spectrometry using an ammonium phosphate matrix modification procedure. The results are shown in Fig. 6. The data from the two methods are combined in Fig. 7 and the ratio of cadmium activity to total cadmium concentration is found to be Cd<sup>2+</sup>/Cd<sub>T</sub> = 0.051. This result is used to convert the total cadmium concentrations in the water only exposure experiment to the cadmium activity.

## C. Interstitial Water Diffusion Sampler

The final task that had to be completed before sediment Bioassay studies could be undertaken was the design and construction of a suitable sampling device. The final design of the peeper is shown in Fig. 8. The device is constructed of acrylic material. The body is 1.5 in. thick with three 0.5 in. holes bored at six different levels 0.75 in. apart. A solid 0.25 in. base plate is fused to the back. The front contains a nucleopore membrane, on top of which is a 20 mil polyethylene gasket and a 0.5 in. cover plate. The entire assembly is held together with six 0.25 in. PVC nuts and bolts. The volume of each cell is 5.0 mL, which provides the necessary minimum sample volume of 15 mL required for the electrode measurement, at each sampling level. This sample volume size was determined by experiment to be adequate for reliable measurement.

The interstitial water sampling device depends upon diffusion across the membrane to measure the interstitial water concentrations. In order to establish the equilibration time for transfer across the membrane the cells of the assembled peeper were filled with distilled water. The device was then immersed in seawater. The cells were sampled periodically and the conductivity of the sample was measured. The results are shown in Fig. 9. As can be seen the 12 micron membrane equilibrated quite rapidly. However even the 1 micron membrane reached equilibrium within 24 hours. Since this equilibration time is well within the time scale of the toxicity experiments, it was decided to use this porosity membrane in subsequent experiments.

## **D. Initial Toxicity Experiments**

When sediment exposure experiments are performed it is necessary to be able to predict the interstitial cadmium concentration that will be obtained from initially combining a quantity of a cadmium salt with measured amount of reference sediment and seawater. To provide this information solutions were prepared with total cadmium concentrations of 200 to 6000 ppm. Each of these contained 1000 mL of control sediment (55% water). The peepers were then placed in the settled sediment-water systems. After two days the cadmium concentration was measured as a function of depth. The results are shown in Fig. 10. Initially it was believed that the interstitial cadmium concentration was being controlled by simple partitioning between a sediment bound component and the aqueous component. The amount of cadmium used in preparing the systems for the interstitial water Bioassay experiments was determined from these results. The attempt was made to bracket the cadmium  $LC_{50}$  obtained in the water only exposure. However, in all of the systems the final interstitial cadmium concentration was not sufficient to produce mortality.

The first bioassay results indicated that an additional process was operating in the cadmiumsediment water system. The observation of a yellow precipitate (probably CdS) when preparing the more concentrated systems suggested that sediment generated sulfide was depressing the aqueous cadmium concentration during the actual exposure.

## E. Development of Acid Volatile Sulfide Extraction Method

The most labile sulfide component of sediments is the acid volatile sulfide (AVS). It is the solid phase sulfide in the sediment that is soluble in cold acid. The measurement technique is to convert the sulfides to  $H_2S(aq)$ , purge it with a gas, and trap it [see Morse et al., 1987 for a review]. A 500 mL Erlenmeyer flask reaction vessel fitted with a three-hole stopper is followed by three sequentially connected 250 mL Erlenmeyer flask trapping vessels. The first is a chloride trap

with 200 mL of pH 4 buffer (0.05M potassium hydrogen phthlate) to prevent chloride carry over. The second and third traps contain 200 mL of a 0.1M silver nitrate solution for trapping H<sub>2</sub>S. The four flasks are connected with airtight appropriately shaped glass and Tygon tubing.

A nitrogen gas flow allows continuous purging of the system. In order to prevent oxidation the gas flows through an oxygen-scrubbing system consisting of a vanadous chloride solution in the first scrubbing tower and the matrix of the analyte (seawater) in the second tower. Vanadous chloride is prepared using four grams of ammonium metavanadate boiled with 50 mL of concentrated hydrochloric acid and diluted to 500 mL. Amalgamated zinc, prepared by taking about 15 grams of zinc, covering it with deionized water and adding 3 drops of concentrated hydrochloric acid before adding a small amount of mercury to complete the amalgamation, is then added to the vanadous chloride solution.

The sediment sample (10-15 grams of wet sediment) or standard to be analyzed is placed in the reaction vessel after the entire system has been purged with nitrogen for about an hour. The system is again purged for 5-10 minutes, and deaerated 6M hydrochloric acid is added from a thistle tube to achieve a final concentration in the vessel of 0.5M. The system is run at room temperature for one hour which has been found to be sufficient to complete the extraction. Fig. 11 present the results of an experiment in which the time course of AVS extraction from a sediment (Long Island Sediment) is followed. It is clear that one hour is sufficiently long for the extraction to be completed.

## **F.** Figure Captions

- Figure. 1 Cadmium electrode calibration curve in 0.7M NaNO<sub>3</sub>, pH = 8 corresponding to the ionic strength of seawater. Slope = 28.2. Sensitivity of the electrode is 0.1 mg/L
- Figure. 2 Cadmium electrode calibration curve in 0.7M NaNO<sub>3</sub>, 0.002M NaHCO<sub>3</sub>, pH = 8, corresponding to the ionic strength and bicarbonate concentration of seawater. Slope = 28.2. Sensitivity of the electrode is 0.1 mg/L.
- Figure 3. Hydroxide titration to determine  $pK_1$  for the reaction:  $Cd^{2+} + OH^- <-> CdOH^+$ .
- Figure 4. Chloride titrations to determine  $K_1$  for the reaction:  $Cd^{2+} + Cl^- <-> CdCl^+$ .
- Figure 5. Ampelisca Toxicity Test: Water Only Exposure. LC50 = 0.32 mg Cd/L (top) and 0.55 mg Cd/L
- Figure 6. Cadmium concentrations in the toxicity test samples (mg Cd/L) determined using a polaragraphic method (top) and using an AA graphite furnace method (bottom) versus electrode concentration (mg Cd<sup>2+</sup>/L).
- Figure 7. Cadmium concentrations in the toxicity test samples (mg Cd/L) determined using both a polaragraphic and AA graphite furnace method versus electrode concentration (mg Cd<sup>2+</sup>/L). Regression line is Cd<sup>2+</sup> = 0.051 Cd, which implies a K<sub>1</sub> of the chloride reaction of K<sub>1</sub> = 26.4 M<sub>-1</sub>.
- Figure 8. Design of diffusion sampler "peeper"

Figure 9. Time to equilibrium for membrane transfer.

Figure 10. Initial interstitial water sampling results. Static exposure.

Figure 11. Time course experiment for AVS extraction of Long Island Sound sediment.







Figure. 2 Cadmium electrode calibration curve in 0.7M NaNO<sub>3</sub>, 0.002M NaHCO<sub>3</sub>, pH = 8, corresponding to the ionic strength and bicarbonate concentration of seawater. Slope = 28.2. Sensitivity of the electrode is 0.1 mg/L.







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Figure 4. Chloride titrations to determine  $K_1$  for the reaction:  $Cd^{2+} + Cl^- <-> CdCl^+$ .



Figure 5. Ampelisca Toxicity Test: Water Only Exposure. LC50 = 0.32 mg Cd/L (top) and 0.55 mg Cd/L



Figure 6. Cadmium concentrations in the toxicity test samples (mg Cd/L) determined using a polaragraphic method (top) and using an AA graphite furnace method (bottom) versus electrode concentration (mg Cd<sup>2+</sup>/L).



Figure 7. Cadmium concentrations in the toxicity test samples (mg Cd/L) determined using both a polaragraphic and AA graphite furnace method versus electrode concentration (mg Cd<sup>2+</sup>/L). Regression line is Cd<sup>2+</sup> = 0.051 Cd, which implies a K<sub>1</sub> of the chloride reaction of K<sub>1</sub> = 26.4 M<sub>-1</sub>.

# PEEPER DESIGN DIMENSIONS (CM)



Figure 8. Design of diffusion sampler "peeper"



Figure 9. Time to equilibrium for membrane transfer.



Figure 10. Initial interstitial water sampling results. Static exposure.



EXTRACTION RATE (umol/gm-min)

## DEVELOPMENT AND VERIFICATION OF A KINETIC

## **MODEL OF POLLUTANT DESORPTION**

## FROM DREDGE SEDIMENT AND SEWAGE SLUDGE

#### I. Introduction

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The purpose of this project is to develop a kinetic model of heavy metal desorption from sludges and sediments. This reaction significantly influences the fate of these chemicals during the ocean disposal of sludge and dredged sediments since the rapidity with which metals desorb determines, to a large extent, their ultimate fate. If desorption is rapid then the chemical is primarily in the aqueous phase and water column transport determines its fate. However if the metal remains in particulate form then settling and sedimentation will occur. Thus the rate and degree of desorption greatly influence the fate of these chemicals.

Significant progress has been made in the development of the models for this reaction. An equilibrium desorption model for reversibly bound heavy metals and organic chemicals was developed in a previous project (Di Toro and Mahony, 1986). The model describes the initial desorption of heavy metals from sewage sludge and Black Rock Harbor sediment upon mixing with seawater at various particle concentrations. Only a small fraction of the total particulate metal initially desorbes. Consecutive desorptions confirmed that only a small fraction was available during short term (1 hour) desorptions. However, it was noted that although desorption of the reversibly bound (or labile) metal fraction was rapid there was a distinct and slower reaction which also released particle bound metal into the aqueous phase. The time scales of this reaction is days rather than hours. This slow release phenomena, which had been observed experimentally by Rohatgi and Chen (1975) for digested sewage sludge, was confirmed by us for Black Rock Harbor sediment. Rohatgi and Chen speculated that the release might be due to an oxidation reaction which liberated heavy metals that either were bound to organic carbon or which were present as metal sulfides.

These observations, and our own experimental experience, lead us to propose a three component model of heavy metal desorption. Particulate metal was assumed to exist as either reversibly sorbed metal, for which the reversible partition coefficient follows the particle interaction model which we had developed in the previous project, and two other com-

ponents. A resistant component which initially is not desorbed but is gradually released over a period of five to ten days; and a refractory fraction which resists release for the duration of the experiment. This model was fit to Rohatgi and Chen's data and it provides a reasonable fit to the observations. An example is shown in fig. 1.

However, Rohatgi and Chen's experiments could not be used to distinguish between a number of other possible models that could easily fit the data as well. For example it is possible that the increase in dissolved concentration of metals during the experiment is due to slow, reversible, desorption kinetics. That is, it is possible that the desorption reaction for these particles takes a number of days before it reaches equilibrium so that the slow release is just due to the slowness of the desorption reaction. Desorption kinetics which are initially rapid but are followed by a slow phase have been observed for hydrophobic organic chemicals (Karickhoff, 1980).

Alternately it may be that the partition coefficient is decreasing during the experiment due to a change in particle properties. If the particle surface properties are being altered by exposure to oxygenated seawater during the experiment then the desorption reaction might still be rapid but it is adjusting to the slowly changing surface properties of the particles. The oxidation of particlulate reactive organic carbon would influence the partitioning in this way.

The purpose of this research project is to experimentally distinguish between these possibilities and to determine, to the extent possible, the mechanism responsible for the slow release of metals and to incorporate any necessary changes into the desorption model. The experiments conducted to date, using Black Rock Harbor sediment and two digested sewage sludges from the Bergen County and Ridgewood sewage treatment plants, appear to confirm the three component model as originally formulated. It appears that the slow release of metals are from the fraction of the metal which is initially present as particulate metal sulfides. These are released as the particulate sulfides is oxidized to sulfate in the reactor. An initial experiment in sulfate free seawater exhibited an increase in sulfate over time confirming the oxidation of sulfide to sulfate. Additionally, an anaerobic reactor, for

which nitrogen gas bubbling was substituted for oxygen containing air bubbling, exhibited no increase in dissolved metal over time. Parallel experiments with radiotagged metal appear to confirm this interpretation although some short term (less than one day) effects are still to be explained.

Hence the data appears to confirm the three component desorption kinetic model. It remains to actually fit these data to the model equations in order to obtain the model parameters. The final version would then be applicable to the evaluation of the fate of heavy metals during the ocean disposal of sludges and dredged sediments.

## II. Experimental Design and Results to Date

The experiments are all conducted in small (350 mL) reaction vessels. Metal free seawater and sludge or sediment particles are initially added. The reactors are kept aerobic by bubbling filtered air and are mixed using magnetic stirring. Duplicate reactors at two particle concentrations, 300 and 3000 mg/L, are used. A particle free control reactor which receives the same air, stirring, and sampling, is monitored to detect possible contamination. At various times during the experiment samples are taken and analyzed for total and dissolved metal (the separation is by centrifugation). The duration of the experiment is twenty to thirty days.

Fig. 2 presents the results obtained for copper from Black Rock Harbor sediment and Bergen County sewage sludge for two particle concentrations (300 and 3000 mg/l). The log scales for the concentration are increased by a factor of ten for the 3000 mg/L reactor so that the results for both reactors can be directly compared. The increase in dissolved copper concentration is approximately one order of magnitude for both particle types with the larger dissolved concentrations associated with the smaller particle concentration. This is a result of the decreased fraction of the metal that reversibly partitions to the particles at lower particle concentrations. Fig. 3 presents the results for zinc and cadmium for Black Rock Harbor sediment. Again the release increases by approximately an order of magnitude. However, the chromium

results in fig. 4 show no significant increase for either Black Rock Harbor sediment or Bergen County sludge. The significance of this fact is discussed below.

Changes in pH and total organic carbon (TOC) are shown in fig. 5. For Black Rock Harbor sediment the pH and TOC are essentially constant. For Bergen County sludge the pH increases slightly during the first day and stayed approximately constant thereafter. Although this may have affected the reversible partition coefficient during this time (it would increase with increasing pH) the release continues during the period of constant pH. The TOC decreased from 400 (40) mgC/L to 200 (20) mgC/L in the 3000 (300) mg/L reactor during the experiment. However the release patterns of metals is similar for Black Rock Harbor sediment and Bergen County sludge. Since no change in TOC or pH was observed for Black Rock Harbor sediment, it is unlikely that the pH and TOC changes were responsible for the metal release.

The results of an anaerobic reactor experiment are shown in fig. 6. The arrangement is the same as the aerobic reactor, the only difference is that nitrogen gas is substituted for the air that is bubbled through the reactors. As can be seen the dissolved copper concentration stayed constant for Black Rock Harbor sediment and the two sludges. The TOC also remained constant as expected since the experiment did not last long enough to initiate significant anaerobic breakdown of the sludges. Thus the presence of oxygen is clearly necessary for the slow metal release to occur.

The fact that copper, cadmium, and zinc are slowly released while chromium concentrations are constant is consistent with the hypothesis that the slow release of metals is due to the oxidation of particulate metal sulfides. The solubility of these metal-sulfides is very low as indicated below (Lindsay, 1979):

Solut	Log Ksp	
CuS	(covellite)	-36.10
CdS	(greennokite)	-27.07
ZnS	(wurtzite)	-22.5

However no Cr(III) sulfide solid phase is reported in tabulations of sulfide mineral solubilities (Naumov et al., 1974).

A possible approach to a direct verification that particulate sulfide is being oxidized is to monitor the oxidation end product, sulfate, An increase in sulfate would and observe its concentration in time. indicate that reduced sulfur, presumably sulfide, was being oxidized in the reactor. The experiment is conducted in sulfate-free artificial seawater so that the large naturally occurring sulfate background does not obscure the increase. The results are shown in fig. 7. The upper panel presents the actual observations. The initial sulfate concentration is due to the sulfate in the supernatant of the Black Rock Harbor sediment stock which was added at the start of the experiment. Future experiments will separate the particles from the supernatant. The bottom panel presents the excess sulfate, defined as the difference between that initially present and that observed at the indicated time. It is interesting to note that the pattern of sulfate release is similar to that observed for the metal release.

In addition to measurements of the total and dissolved metal concentrations over time additional parallel experiments were conducted with radiotagged metals. The purpose of these experiments was to examine the question of the time scale of adsorption and the possible variation of the partition coefficient over time. The kinetic experiment used the same reactor setup. After the particles were added the reactor was tagged with radioactive metal. The total and dissolved radiotagged metal was monitored in time. Changes in the distribution between total and dissolved metal could be due to either slow adsorption kinetics or changes in the adsorption partition coefficient. But the absence of change would eliminate both mechanisms as significantly contributing to the release phenomena.

The results from these kinetic experiments are shown in fig. 8 for zinc. The Bergen County sludge results indicate that very little change occurred in the dissolved concentration suggesting that the variation of TOC did not significantly change the partitioning. However the Black Rock Harbor sediment results indicated a substantial increase in dissolved concentration and a decrease in total concentration that is

attributed to the accumulation of sediment particles on the walls of the reactor. However the changing total concentration casts some doubt on the validity of the experiment so we plan to repeat it.

The results for cadmium are shown in fig. 9. With the exception of the data for less than one day, the dissolved cadmium concentration remained constant for the duration of the experiment for both Black Rock Harbor sediment and Bergen County sludge.

These experiments do not yield a completely consistent interpretation. However they do eliminate one possible model - that the adsorption and desorption kinetics are slow and reversible. If sorption is slow but ultimately reversible then for an adsorption experiment the dissolved concentration should initially be large, since adsorption has not yet had an opportunity to occur, and it should decrease with time. For the experiments where changes were observed the reverse pattern was observed so that slow reversible sorption kinetics appear to be ruled out.

In order to discriminate between adsorption kinetics and partition coefficient changes, an additional set of parallel experiments were performed in order to examine if the partition coefficient is varying in time. A parallel reactor was used which was not initially tagged with radioactive metal. Rather, at various times during the experiment a small sample was taken. This sample was then spiked with a small amount of radioactive metal. The sample was agitated for one hour after which the total and dissolved radioactive metal concentration was determined. Following the adsorption step, a desorption step was performed in order to examine the behavior of the reversibly sorbed metal. If the distribution of total and dissolved metal was changing with time, that would indicate that the partition coefficients were changing with time.

The results from these spiking experiments are shown in fig. 10 for zinc. With the exception of the changing adsorption partition coefficient for Black Rock Harbor sediment, the other partition coefficients are constant. It is interesting that the desorption partition coefficient is constant for both Black Rock Harbor sediment and Bergen County sludge.

The results of the spiking experiments for cadmium are shown in fig. 11. Once again the partition coefficients appear to be quite constant for times greater than one day. But both the adsorption and desorption partition coefficients are changing initially.

The results of the kinetic and spiking experiments are not consistent with a single explanation. On the one hand, the Bergen County sludge zinc kinetic and the spike adsorption-desorption data, the Black Rock Harbor sediment spike desorption data, and both the kinetic and spike adsorption-desorption Black Rock Harbor sediment and Bergen County sludge cadmium data for time greater than one day suggest that no partition coefficient changes are occurring since the dissolved concentrations are essentially constant. However, the Black Rock Harbor sediment zinc kinetic and spike adsorption data, and all the less than one day cadmium data, suggest that something was changing during these time periods. Additional experiments are planned to investigate the cause of these unexpected and as yet unexplained results.

## III. Future Directions

The major focus of this portion of the research will be to complete the development of the kinetic model for metal desorption from sediments and sludges. The reactor data will be fit to the model equations to estimate the parameters. Additional radiotag experiments will be performed in an attempt to understand the somewhat anomalous results described above. However the focus of the work will be to complete the development with a minimum of experimental effort so that the Sediment Criteria work can begin. A project report will be written that includes all the experimental data as well as the modeling results. A journal article will also be prepared.

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#### FIGURE CAPTIONS

- 1. Experimental data (Rohatgi and Chen, 1975) and the three component model fit for cadmium and chromium.
- Total and dissolved copper concentrations versus time for Black Rock Harbor sediment and Bergen County sludge at 3000 mg/L (left side) and 300 mg/L (right side). Control reactor concentrations (C) are also shown.
- 3. Total and dissolved zinc (top) and cadmium (bottom) concentrations versus time for Black Rock Harbor sediment and Bergen County sludge at 3000 mg/L (left side) and 300 mg/L (right side).
- Total and dissolved chromium concentrations versus time for Black Rock Harbor sediment and Bergen County sludge at 3000 mg/L (left side) and 300 mg/L (right side).
- 5. pH (top) and Total Organic Carbon (bottom) versus time for Black Rock Harbor sediment (left) and Bergen County sludge (right) reactors.
- 6. Anaerobic Reactor dissolved copper concentrations versus time for Black Rock Harbor (top left), Ridgewood (bottom left) and Bergen County (top right). Total organic carbon concentrations versus time for the three reactors are also shown (bottom right).
- 7. Sulfate (top) and excess sulfate (bottom) concentration versus time for Black Rock Harbor sediment.
- 8. Concentration of total and dissolved radioactive zinc versus time. Radioactive zinc is added at the start (t=0) of the experiment.
- Concentration of total and dissolved radioactive cadmium versus time. Radioactive cadmium is added at the start (t=0) of the experiment.
- 10. Concentration of total and dissolved radioactive zinc at various times. Radioactive zinc is added to a subsample taken at the indicated time and a one hour - one hour adsorption-desorption measurement is made. The total and dissolved zinc at adsorption (top) and desorption (bottom) is shown for Black Rock Harbor (left) and Bergen County (right).
- 11. Concentration of total and dissolved radioactive cadmium at various times. Radioactive cadmium is added to a subsample taken at the indicated time and a one hour one hour adsorption-desorption measurement is made. The total and dissolved cadmium at adsorption (top) and desorption (bottom) is shown for Black Rock Harbor (left) and Bergen County (right).



Experimental data (Rohatgi and Chen, 1975) and the three component model fit for cadmium and chromium.





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7. Sulfate (top) and excess sulfate (bottom) concentration versus time for Black Rock Harbor sediment.

## BLACK ROCK HARBOR SEDIMENT - #2

## PARTITION COEFFICIENT KINETICS





PARTITION COEFFICIENT KINETICS




## BLACK ROCK HARBOR SEDIMENT - #6

## PARTITION COEFFICIENT KINETICS



BERGEN COUNTY SLUDGE - #7

PARTITION COEFFICIENT KINETICS



 Concentration of total and dissolved radioactive cadmium versus time. Radioactive cadmium is added at the start (t=0) of the experiment.







