

**DEVELOPMENT AND VERIFICATION OF A KINETIC
MODEL OF POLLUTANT DESORPTION
FROM DREDGE SEDIMENT AND SEWAGE SLUDGE**

FINAL REPORT

by

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OVERVIEW

This final report encompasses a two phase research effort. The initial direction was toward the development and verification of a kinetic model of heavy metal desorption from dredge sediment and sewage sludge. The second phase used the insights and results gained from that effort to begin an investigation of the toxicity of cadmium in sediments. This change of direction was agreed upon by both the Manhattan College and EPA Narragansett Laboratory researchers and was reflected in the renewal application for the second year of funding. The report is divided into distinct parts, reflecting each topic.

The first part reports on the role of solid phase sulfide in determining the toxicity of cadmium in sediments. For marine sediments the importance of sulfide and the possibility of the formation of insoluble metal sulfides has often been pointed out. We have shown that acid volatile sulfide - solid phase amorphous FeS(s) and MnS(s) which are soluble in cold acid - is the dominant sediment property that controls the cadmium binding capacity of marine sediments and provides the proper normalization for establishing the toxicity of cadmium in a variety of marine sediments. This is a major finding of our research project. The results are presented as a paper which is being submitted for publication. An additional Appendix II is included which presents all the experimental data in tabular form.

The second part of this report presents the results of the development of the methodology that was needed for performing the experiments and measurements to assess the toxicity of cadmium in sediments. The experimental calibration of the cadmium electrode and the design and testing of a diffusional sampler is presented. Also a validation experiment for the Acid Volatile Sulfide extraction method is presented.

The third part of this report presents the results of the sludge and sediment desorption experiments. These data confirm that the original model proposed for this reaction was correct. A three phase model is appropriate: A reversibly sorbed component; a metal sulfide component that is released via oxidation; and a refractory component that is not released within the time scale of the experiment.

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**TOXICITY OF CADMIUM IN SEDIMENTS:
THE ROLE OF ACID VOLATILE SULFIDE**

by

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INTRODUCTION

The toxicity of chemicals in sediments is strongly influenced by the extent to which the chemical binds to the sediment. This modifies the chemical potential to which the organisms are subjected. As a consequence different sediment types will exhibit different degrees of toxicity for the same total quantity of chemical. These differences have been reconciled by relating organism response to the chemical concentration in the interstitial water of the sediments [see the review in EPA, 1989]. As a consequence the relevant sediment properties are those which influence the distribution of chemical between the solid and aqueous phases.

The varying toxicity of non-ionic organic chemicals in different sediments has been found to be primarily determined by the organic carbon content of the sediments [EPA, 1989]. The purpose of this paper is to establish the importance of another sediment phase: the acid volatile sulfide phase - the sediment sulfides that are soluble in cold acid - in determining the toxicity of cadmium in sediments. By implication, this phase is likely to be important for all metals which form insoluble sulfides. Most freshwater and marine sediments - completely aerobic sediments are the exception - contain sufficient acid volatile sulfide for this phase to be the predominant determinant of toxicity.

METHODS AND MATERIALS

A. Exposure System Design

Sediment dwelling amphipods were exposed for 10 days to control and cadmium-spiked sediments in a 900 mL flow-through chamber with 200 mL of sediment (3.5 cm depth) and 600 mL of overlying seawater. Lighting was continuous to inhibit the amphipods' swimming behavior. Filtered air and seawater flow (10 volume replacements/day) ensured acceptable dissolved oxygen concentrations and cadmium free overlying water.

A diffusion sampler ("peeper") [Hesslein, 1976; Carignan et al., 1984; 1985], designed to fit within the exposure chamber and sample the interstitial and overlying water concentrations, was constructed of Plexiglas G grade unshrunken cast acrylic sheet: 6 x 3 x 2 in. deep with 6 rows of 3 3/4 in. diameter 1 1/2 in. deep holes, each of which has a volume of about 5 mL. The open side of the

peeper is covered by a sheet of 1 micron polycarbonate membrane (Nucleopore), followed by a 30 mil low density polyethylene gasket and a 1/2 inch Plexiglas cover plate, both of which have the same hole pattern as the body and secured with PVC-1 cap screws and nuts. Equilibration time was measured to be less than one day.

B. Organism Collection and Holding

Ampelisca abdita were collected from tidal flats in the Pettaquamscutt (Narrow) River, a small estuary flowing into Narragansett Bay, RI, transferred to the laboratory within one half hour, and sieved through a 0.5mm mesh screen. *Ampelisca* were collected with a dip net after flotation on the air/water interface. *Rhepoxynius hudsoni* were collected in shallow water at Ninigret Pond, RI. Adult animals were sieved from the sediment through a 1 mm mesh screen in the field, transported to the laboratory within an hour, sieved again and transferred to holding containers. The amphipods were maintained in presieved uncontaminated collection site sediment and flowing filtered seawater, and acclimated to the assay temperature at the rate of 1 to 3°C per day. During acclimation, the *Ampelisca* were fed, *ad libitum*, the laboratory cultured diatom *Phaeodactylum tricornutum*. *Rhepoxynius* were not fed.

C. Sediment Acid Volatile Sulfide

The principal property of concern of the sediments used in these experiments was the acid volatile sulfide (AVS) concentration. It is the solid phase sulfide in the sediment that is soluble in cold acid. The measurement technique is to convert the sulfides to H₂S(aq), purge it with a gas, and trap it [see Morse et al., 1987 for a review]. A 500 mL Erlenmeyer flask reaction vessel fitted with a three-hole stopper is followed by three sequentially connected 250 mL Erlenmeyer flask trapping vessels. The first is a chloride trap with 200 mL of pH 4 buffer (0.05M potassium hydrogen phthalate) to prevent chloride carry over. The second and third traps contain 200 mL of a 0.1M silver nitrate solution for trapping H₂S. The four flasks are connected with airtight appropriately shaped glass and Tygon tubing.

A nitrogen gas flow allows continuous purging of the system. In order to prevent oxidation the gas flows through an oxygen-scrubbing system consisting of a vanadous chloride solution in the

first scrubbing tower and the matrix of the analyte (seawater) in the second tower. Vanadous chloride is prepared using four grams of ammonium metavanadate boiled with 50 mL of concentrated hydrochloric acid and diluted to 500 mL. Amalgamated zinc, prepared by taking about 15 grams of zinc, covering it with deionized water and adding 3 drops of concentrated hydrochloric acid before adding a small amount of mercury to complete the amalgamation, is then added to the vanadous chloride solution.

The sediment sample (10-15 grams of wet sediment) or standard to be analyzed is placed in the reaction vessel after the entire system has been purged with nitrogen for about an hour. The system is again purged for 5-10 minutes, and deaerated 6M hydrochloric acid is added from a thistle tube to achieve a final concentration in the vessel of 0.5M. The system is run at room temperature for one hour which has been found to be sufficient to complete the extraction. The nitrogen gas flows at a bubble rate of about four per second. The sample vessel is swirled every five or ten minutes. At completion all hydrogen sulfide produced has been converted to silver sulfide in the first silver nitrate trap and no precipitate is found the second trap. The suspension in the first silver nitrate trap is passed through a 1.2 micron GF fiber filter, dried at 102°C, and weighed.

Standards prepared from appropriate quantities of iron(II) sulfate and sodium sulfide (the latter being added from a solution standardized against lead perchlorate), typically gave yields of 95-103%. Silver sulfide precipitates were usually in the range 20-30 mg. When a blank was run (sample without acid), about 0.9 mg silver sulfide was obtained. When the acid was run without a sample, about 0.6 mg silver chloride was obtained. This corresponds to a detection limit of ~ 0.5 $\mu\text{mol/g}$.

D. Sediment Characterization and Spiking Procedure

Sediments of three different acid-volatile sulfide concentrations were used in the toxicity tests. The LI Sound sediment, with a high AVS concentration, was collected from an uncontaminated site in central Long Island Sound ($40^{\circ}7.95'N$ and $72^{\circ}52.7'W$) with a Smith-MacIntyre grab sampler, returned to the laboratory, press sieved wet through a 2 mm mesh stainless steel screen, homogenized, and stored at 4°C. *A. abdita* has been tested many times in this sediment and both its survival and reproduction have been good (Scott and Redmond, in press). The Ninigret Pond

sediment was a low AVS sand collected from the *Rhepoxyinius* collection site. The upper few inches of sediment were collected with a shovel, returned to the laboratory, sieved wet through a 2 mm stainless steel screen, rinsed several times to remove high-organic fine particles, homogenized, and stored at 4°C. The third sediment was a 50/50 (volume) mixture of LI Sound and Ninigret Pond sediments.

Sediments were spiked by adding 1000 mL of wet sediment to 1500 mL of 20°C filtered seawater into which a weighted amount of cadmium chloride had been dissolved. An additional 500 mL of filtered seawater was used to rinse the sediment container. The mixture was stirred with a nylon spatula, capped and placed on a paint shaker for 5 minutes to ensure complete mixing, and held at ambient temperature (~15°C) water bath for 7 days to ensure equilibrium of the cadmium and sediment. A thin layer of cadmium sulfide precipitate that had formed on the surface of the sediment was removed, the test sediments were then homogenized, and 200 mL were transferred to each of three replicate exposure containers. For the experiments with peepers, they were inserted at this time. Exposure containers were placed in the water bath with air and seawater delivery.

E. Toxicity Experiment

The amphipods were sieved from holding containers through a 0.5 mm stainless steel screen and distributed sequentially into 100 mL plastic beakers. After sorting and eliminating dead or outsized animals, the beakers were randomized, air delivery in the exposure system was halted, and one beaker of amphipods was added to the two replicate exposure containers in each treatment. *Rhepoxyinius* were added to the Ninigret Pond treatments, and *Ampelisca* to the LI Sound and mixture treatments. The third replicate exposure container in each treatment received no amphipods and was used as a chemical control. Salinity and temperature of the overlying seawater remained relatively constant at $20.7 \pm 0.3^\circ\text{C}$ ($n=11$) and $30.3 \pm 0.5\%$ ($n=11$) during the 10 day exposure period.

After termination the contents of each exposure container were sieved through a 0.5 mm screen. For *Ampelisca*, material retained on the sieve was preserved in 5% buffered formalin with

Rose Bengal stain for later sorting. For *Rhepoxynius*, material retained on the sieve was examined immediately after sieving. In both cases, recovered animals were counted, and any missing individuals were counted as mortalities. The peeper interstitial and overlying water cadmium concentrations were determined as was the AVS and solid phase cadmium in the chemical control vessels.

F. Cadmium Determinations and Titrations

The cadmium ion concentration in both the peeper samples and the titrations described below was measured as Cd²⁺ activity using an Orion 94-48 cadmium ion selective electrode and a double junction reference electrode (Orion 90-02). The electrode was standardized with a serial dilution of a 1 g/L cadmium solution that was also used as the titrant. Sediment cadmium was determined using a cold concentrated nitric acid (16M, 5mL) digestion of 10mL wet sediment followed by a peroxide oxidation (10mL 30%) and evaporation to dryness. The residue is reconstitute to 20mL using 0.1M nitric acid and the cadmium measured using an AA.

Cadmium titrations of FeS suspensions (prepared in the same manner as the AVS standards) and sediments were performed using sample sizes of 5 to 10 gm dry wt. added to 50 mL seawater which was constantly stirred. Cadmium chloride was added and dissolved cadmium was monitored using the electrode. Anaerobic conditions were maintained using a nitrogen atmosphere provided by a glove box or by constantly bubbling nitrogen through the covered titration vessel. In the sediment titrations where electrode response was slow, a uniform differential response-time procedure was employed to obtain consistent voltage readings.

EXPERIMENTAL RESULTS AND INTERSTITIAL WATER CORRELATIONS

A. Dry weight normalization

The toxicity of cadmium to *Rhepoxynius hudsoni* in Ninigret Pond sediment; and to *Ampe-lisca* in Long Island Sound sediment and an equal parts mixture of the two sediments, is shown in Fig 1. The curves are log-logistic concentration response functions with the same slope parameter (Table 1). The LC50s range from 318 µmol/gm to 3200 µmol/gm on a sediment dry weight basis. As shown below these two organisms have virtually the same LC50s in water only exposures: 0.012

mg Cd²⁺/L (0.23 mg Cd/L) and 0.017 mg Cd²⁺/L (0.34 mg Cd/L) for *Rhepoxynius* and *Ampelisca* respectively. The concentrations are for free cadmium, Cd²⁺, and total dissolved cadmium, Cd, respectively. Hence the differences in the cadmium toxicity are likely to be attributable to varying sediment properties. In addition Swartz et al., (1985) reported the *Rhepoxynius abronius* cadmium LC50 for a Yaquina Bay sediment to be 25 $\mu\text{mol/gm}$. Thus a factor of ten separates each of the LC50s for these three marine sediments: Long Island Sound (3200 $\mu\text{mol/gm}$), Ninigret Pond (318 $\mu\text{mol/gm}$) and Yaquina Bay (25 $\mu\text{mol/gm}$). An explanation for the over two order of magnitude variation in LC50s would surely be useful.

B. Correlation to Interstitial Water concentration

The correlation between organism toxicity and interstitial water concentration for sediments with different dry weight sediment toxicity has been reported (Adams et al., 1985; Swartz et al., 1985; Kemp and Swartz, 1986). In addition the evidence suggests [Borgmann, 1983] that biological response correlates to chemical activity, in particular to the divalent metal activity, {Me²⁺} [Sunda and Guillard, 1976; Sunda et al., 1978; Zamuda and Sunda, 1982]. The claim is not that the only bioavailable form of the metal is Me²⁺ - for example MeOH⁺ may also be bioavailable - but that the DOC or other ligand complexed fractions are not bioavailable.

These two hypotheses are examined in Fig. 2a, a comparison of the observed mortality to the observed interstitial water cadmium activity, measured with the specific ion electrode, for the three sediments in Fig. 1. The concentration response curves for *Ampelisca* and *Rhepoxynius* in water only exposures are nearly identical. The interstitial water concentration data from the sediment exposures are somewhat scattered. However the grouped data, presented in Fig. 2b as medians (50th percentile) and interquartile ranges (25th to 75th percentiles) parallel the water only exposure curve. These results conform to previous observations that the concentration response curves for sediment exposures, which are quite different on a sediment cadmium dry weight basis (Fig. 1), are quite comparable on an interstitial water basis. Table 1 presents the results.

C. Sediment Cadmium vs. Interstitial Water

The prediction of the toxicity of cadmium in sediments requires that the relationship between sediment cadmium concentration and interstitial water concentration be established. A plot of solid phase versus aqueous phase cadmium concentrations - which is regularly used for the analysis of sorption data - is shown in Fig. 3.

The data can be envisioned as a titration in which cadmium is added incrementally to the sediment and the resulting aqueous and solid phase cadmium distribution is measured. Initially the solid phase concentration increases but the aqueous phase concentration remains below the detection limit of the cadmium electrode. Then a critical sediment concentration is reached at which point the aqueous concentration increases sharply - in the region marked "transition" in Fig. 3. Note that the increase is over two orders of magnitude in aqueous concentration while the sediment concentration remains nearly constant. As more cadmium is added, the data then appear to follow a linear trend which is characteristic of a sorption reaction.

It is apparent that the critical part of the relationship between solid and aqueous phase cadmium is the onset of the transition region. There is a sudden increase in interstitial water cadmium activity ($\text{mg Cd}^{2+}/\text{L}$) and total dissolved concentration (mg Cd/L) from nonlethal levels below $0.001 \text{ mg Cd}^{2+}/\text{L}$ (0.02 mg Cd/L), passing the water only LC50: $0.015 \text{ mg Cd}^{2+}/\text{L}$ (0.30 mg Cd/L) for *Rhepoxynius* and *Ampelisca*, to concentrations in excess of $0.1 \text{ mg Cd}^{2+}/\text{L}$ (2.0 mg Cd/L). This marks the transition between nontoxic and toxic sediments. The solid phase - aqueous phase relationship at the lower sediment cadmium concentrations is unclear since the aqueous concentrations are below detection. However the data do not appear to conform to a straight line sorption isotherm that would be inferred by extrapolation from the high concentration data since detectable dissolved concentrations would have been present. The more likely possibility is that a precipitation reaction is maintaining the aqueous phase concentration at below detectable values in the region of low sediment concentrations. Since these are marine sediments the possibility of the formation of a cadmium sulfide precipitate is suggested.

METAL SULFIDES AND CADMIUM TITRATIONS

The importance of sulfide in the control of interstitial water metal concentrations of marine sediments is well documented (Boulegue, 1983; Emerson et al., 1983; Davies-Cooley et al., 1985; Morse et al., 1987). Metal sulfides are very insoluble and the equilibrium interstitial water metal concentrations in their presence are small. It is possible that the interstitial water sulfide concentration in the sediment samples used for these toxicity tests was initially high enough that so that as cadmium was added to the sediment, cadmium sulfide was precipitating following the reaction:



However direct measurements of the interstitial water sulfide activity, $\{S^{2-}\}$, with a sulfide electrode failed to detect any free sulfide in the unspiked sediments. This was a most puzzling result since it was visually clear that a bright yellow cadmium sulfide precipitate was forming as cadmium was added to the sediment.

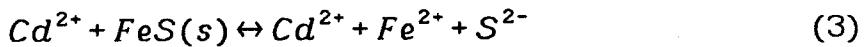
The lack of significant quantity of dissolved sulfide in the interstitial water and the evident formation of solid phase cadmium sulfide suggested the following possibility. Most of the sulfide in sediments is in the form of solid phase iron sulfides. Perhaps the source of the sulfide is the *solid phase* sulfide initially present. Then as cadmium is added to the sediment it causes the solid phase iron sulfide to dissolve releasing sulfide which is available for the formation of cadmium sulfide. This possibility is examined below.

A. Solubility Relationships and Displacement Reactions

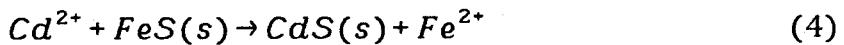
The majority of sulfide in sediments is in the form of iron monosulfides (mackinawite and greigite) and iron bisulfide (pyrite) of which the former are most reactive [see the review by Morse et al., 1987]. Iron monosulfide, $FeS(s)$, is in equilibrium with aqueous phase sulfide via the reaction:



If cadmium is added to the aqueous phase, then the result is:



As the cadmium concentration increases, $[Cd^{2+}][S^{2-}]$ will exceed the solubility product of cadmium sulfide and $CdS(s)$ will start to form. Since cadmium sulfide is more insoluble than iron monosulfide, $FeS(s)$ should start to dissolve in response to the lowered sulfide concentration in the interstitial water. The overall reaction is:



where the iron in $FeS(s)$ is displaced by cadmium to form soluble iron and solid cadmium sulfide, $CdS(s)$. A theoretical analysis of the Cd(II)-Fe(II)-S(II) system, presented in Appendix I, supports this conclusion. The relevant parameter, which can be termed the metal sulfide solubility parameter for any metal, Me, is $\alpha_{Me^{2+}} K_{MeS}$. It is the product of $\alpha_{Me^{2+}} = [\Sigma Me(aq)] / [Me^{2+}]$, the ratio of total dissolved Me to the divalent species concentration; and $K_{MeS} = [Me^{2+}][S^{2-}]$, the metal sulfide solubility product. These are given in Table 2. The sulfide solubility parameters, shown in Fig. 4, determines the behavior of $[FeS(s)]$ and $[MeS(s)]$ as the metal is added to the sediment. For example since the cadmium sulfide solubility parameter is less than the iron sulfide solubility parameter, cadmium will form a sulfide at the expense of the iron sulfide which will dissolve. Note that all the metals examined in Fig. 4 are predicted to dissolve FeS and MnS .

B. Experimental Results - FeS

The calculations presented above reflect the chemical composition expected at thermodynamic equilibrium. However many solid phase reactions are not at equilibrium with respect to either the aqueous phase or other solid phases because of the slow kinetics involved in the necessary transformations. Therefore a direct test of the extent to which this reaction takes place has been performed.

A quantity of freshly precipitated iron sulfide is titrated by adding dissolved cadmium. The resulting aqueous cadmium activity, measured with the cadmium electrode versus the ratio of cadmium added, $[Cd]_A$, to the amount of FeS initially present, $[FeS(s)]_i$, is shown in Fig. 5. The electrode potentials (left) correspond to a very low cadmium concentration during the initial portion of

the titration. Then a sharp upward inflection occurs near $[Cd]_A \sim [FeS]_i$ indicating that all the iron sulfide has dissolved to form CdS and any additional cadmium added appears as free cadmium. The plot of dissolved cadmium versus cadmium added (right) illustrates the rapid increase in dissolved cadmium that occurs near $[Cd]_A / [FeS]_i = 1$. A similar experiment has been performed for amorphous MnS with comparable results. It is interesting to note that such a replacement reaction was postulated by Pankow (1979) to explain an experimental result using copper and FeS.

These experiments plainly demonstrate that solid phase amorphous iron and manganese sulfide can readily be dissolved by adding cadmium. As a consequence it is a source of available sulfide which must be taken into account in evaluating the relationship between solid phase and aqueous phase cadmium in sediments.

C. Titration results - Sediments

A similar titration procedure has been used to evaluate the behavior of sediment samples taken from four quite different marine environments: Black Rock Harbor; the Long Island Sound and Ninigret Pond sediments used in the toxicity tests; and the Hudson River. The results are shown in Fig. 6. The binding capacity for cadmium is estimated by extrapolating a straight line fit to the dissolved cadmium data. The equation is:

$$[\Sigma Cd(aq)] = \max \{0, m([Cd]_A - [Cd]_B)\} \quad (5)$$

where $[\Sigma Cd(aq)]$ is the total dissolved cadmium, $[Cd]_A$ is the cadmium added, $[Cd]_B$ is the bound cadmium, and m is the slope of the straight line. The sediments exhibit quite different binding capacities for cadmium, listed in Table 3, ranging from approximately 1 $\mu\text{mol/gm}$ to more than 100 $\mu\text{mol/gm}$. The question is whether this binding capacity is explained by the solid phase acid volatile sulfide present in the samples.

D. Correlation to Sediment AVS

Sulfides in sediments can be partitioned into three broad classes which reflect the techniques used for quantification [Berner, 1971; Goldhaber and Kaplan, 1974; Morse et al., 1987]. The most labile fraction, acid volatile sulfide (AVS), is associated with the more soluble iron and manganese

monosulfides. The more resistant sulfide mineral phase, iron pyrite, is not soluble in the cold acid extraction used to measure AVS. Neither is the third compartment, organic sulfide associated with the organic matter in sediments [Landers et al., 1983].

The possibility that acid volatile sulfide is a direct measure of the solid phase sulfide that reacts with cadmium is examined in Fig. 7 (left): a plot of the sediment binding capacity for cadmium versus the measured initial AVS for each sediment. The line of perfect agreement is shown. The sediment cadmium binding capacity appears to be somewhat less than the initial AVS for the sediments tested. However a comparison between the initial AVS of the sediments and that remaining after the cadmium titration is completed, Fig. 7 (right), suggests that some AVS is lost during the titration procedure. It is possible that a portion is oxidized even though deoxygenated N₂ gas is passed through the reactor. Or it is possible that some of the AVS is lost as H₂S(g) via stripping into the gas stream. In any case the correlation of sediment binding capacity and final AVS is apparent in Fig. 7. This strongly suggests that AVS is the proper quantification of the solid phase sulfides that can be dissolved by cadmium.

SEDIMENT TOXICITY AND AVS NORMALIZATION

The toxicity experiment illustrated in Fig. 1 was designed to test the utility of AVS as a predictor of the cadmium binding capacity of sediments and therefore a predictor of the concentration of cadmium that would cause sediment toxicity.

A. Experimental Results

Fig. 8 presents the AVS and cadmium concentration data at the start and end of the experiment. The initial concentrations of AVS are averages of multiple measurements from the stock supply. The initial concentrations of cadmium are calculated from the weighted amounts added to the sediments. The final cadmium and AVS concentrations are measured in parallel chemical control vessels.

The AVS results are interesting. A constant amount of AVS, ~2 $\mu\text{mol/gm}$, is lost from each vessel. Since each sediment is exposed to aerobic overlying water for the same amount of time it is probable that the loss is via oxidation. The final cadmium concentrations are within 70% of the initial values reflecting the loss during initial preparation and via the flowing overlying water.

The toxicity experimental results are shown in Fig. 9. The sediment cadmium is normalized by the AVS for that sediment. The averages of the initial and final values are used for AVS. The relationship in Fig. 8 is used to estimate the final cadmium concentration from the initial concentration if it was not measured. Note that the increase in mortality occurs at the point where the sediment cadmium begins to exceed the sediment AVS on a molar basis. Total mortality occurs at $[\text{Cd}]/[\text{AVS}] > 3$. The LC50 that results from a combined fit of the data is 1.97 $\mu\text{mol Cd}/\mu\text{mol AVS}$.

The critical point is that the sediment AVS can be used to normalize the sediment cadmium concentration in the same way that sediment organic carbon is used to normalize non-ionic organic chemicals. The reason that both methods work is that they properly account for the chemical activity of the chemical in both the aqueous and sediment phases. Below 1 $\mu\text{mol Cd}/\mu\text{mol AVS}$ the cadmium is all precipitated as CdS(s) and the activity of Cd is very low. Above 1 $\mu\text{mol Cd}/\mu\text{mol AVS}$ there exists free cadmium in the interstitial water, sorbed cadmium in the sediment phase, as well as CdS(s) . The activity of cadmium in the system is now high enough to cause mortality. This is true for sediments with an appreciable amount of AVS, >1 $\mu\text{mol/gm}$. The reason is that the additional cadmium added in excess of 1 $\mu\text{mol Cd/gm}/\mu\text{mol AVS/gm}$ is large enough to exceed the activity of cadmium in the system that causes mortality even in the presence of some sorption phases - see Fig.5.

IMPLICATIONS FOR METAL TOXICITY IN SEDIMENTS

The first order importance of AVS in determining the toxicity of cadmium in sediments has important implications. These are discussed below.

A. Application to Other Metals and Mixtures

The other potentially toxic metals all form metal sulfide precipitates that are more insoluble than iron sulfide. Fig. 4 presents the sulfide solubility parameters for divalent metals. The iron and manganese sulfides have $\log(\alpha K_{sp}) > -25$ whereas the remaining sulfides have $\log(\alpha K_{sp}) < -25$. The implication is that the results found for cadmium are applicable to these other metals as well since, at equilibrium, they can displace iron and manganese sulfide to form a more insoluble sulfide precipitate.

In particular it is likely that the LC50 for any metal is at least $1 \mu\text{mol}/\mu\text{mol AVS}$. Given the high concentrations of AVS in most sediments, the LC50s of these metals are likely to be large concentrations. For a molecular weight range of Ni ~ 50 to Pb ~ 200 gm/mol, the LC50s for an AVS of 1 (10) $\mu\text{mol AVS/gm}$ would range from 50 (500) to 200 (2000) $\mu\text{g/gm}$.

A additional conjecture is that the molar AVS normalized toxicity of metals is additive. Since all the divalent metals in Fig. 4 have lower sulfide solubility parameters than FeS, they would all exist as metal sulfides if their molar sum is less than the AVS. For this case no metal toxicity would be expected and:

$$\frac{\sum_i [Me_T]_i}{[AVS]} < 1 \quad (6)$$

where $[Me_T]_i$ is the total cold acid extractable metal concentration in the sediment. On the other hand if their molar sum is greater than the AVS concentration then a portion of the metals with the lowest sulfide solubility parameters would exist as free metal and presumably exert a toxicity. For this case the following would be true:

$$\frac{\sum_i [Me_T]_i}{[AVS]} > 1 \quad (7)$$

But these two equations are precisely the formulas that one would employ to determine the extent of metal toxicity in sediments assuming additive behavior and neglecting the effect of partitioning. Whether the normalized sum is less than or greater than one discriminates between non toxic and

toxic sediments. The additivity does not come from the nature of the mechanism that causes toxicity. Rather it results from their equal ability of the metals to form metal sulfides with the same stoichiometric ratio of Me and S.

This discussion is predicated on the assumption that all the metal sulfides behave similarly to cadmium sulfide. Further it has been assumed that only acid soluble metals are reactive enough to affect the free metal activity. At present no experimental data to support either of these conjectures exists so that this discussion purely speculative.

B. AVS and Sediment Quality Criteria

Since AVS can bind cadmium and presumably metals and thereby reduce their toxicity AVS will obviously play a role in the determination of sediment quality criteria for metals. For sediments with very little or no AVS - fully oxidized sediments for example - an AVS normalization would not be appropriate. Rather the partitioning would be controlled by other sediment phases such as iron and manganese oxides and organic carbon (Jenne et al., 1986). An estimate of when partitioning to other phases can be important can be made using the proposed sediment quality criteria formula [EPA, 1989]:

$$r_{SQC} = K_p c_{WQC} \quad (8)$$

where r_{SQC} is the sediment quality criteria, K_p is the partition coefficient, and c_{WQC} is the chronic water quality criteria. For the case where there is only one metal competing for the AVS, the molar equivalent of the AVS would not bioavailable. Therefore it should be added to the allowable concentration so that:

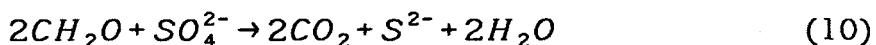
$$[r_{SQC}] = [AVS] + K_p [c_{WQC}] \quad (9)$$

where $[r_{SQC}]$ is the molar sediment quality criteria ($\mu\text{mol/gm}$), K_p is the partition coefficient (L/gm), and $[c_{WQC}]$ is the molar chronic water quality criteria ($\mu\text{mol/L}$). The range for freshwater acute (chronic) criteria for the metals in Fig. 4 (hardness = 100 mg/L) is 0.01 to 31 (.0001 to 1.6) $\mu\text{mol/L}$. The marine criteria are 0.01 to 3.8 (.0001 to 0.88) $\mu\text{mol/L}$ [EPA, 1986]. The

importance of partitioning can be judged by comparing the product $K_p[c_{WOC}]$ to the AVS concentration. Consider an AVS concentration of $1 \mu\text{mol/gm}$. If the partition coefficient is $K_p = 1 \text{ L/gm}$ then a metal with a criteria concentration of $1 \mu\text{mol/L}$ would have its sediment quality criteria doubled due to the partitioning. For $K_p = 10 \text{ L/gm}$ the criteria concentration at which partitioning doubles the sediment quality criteria drops to $0.1 \mu\text{mol/L}$. Hence the effect of partitioning only becomes significant for relatively low AVS concentrations ($\sim 1 \mu\text{mol AVS/gm}$) and for the metals with larger partition coefficients and criteria concentrations. For all cases with only a single metal involved, the minimum molar sediment quality criterion is [AVS].

C. AVS in Freshwater Sediments

Acid volatile sulfide is commonly found in marine sediments. It is produced by the diagene-sis of particulate organic carbon, represented as CH_2O , with sulfate as the electron acceptor [Goldhaber and Kaplan, 1974]:



and the precipitation of iron sulfide [Berner, 1971]:



It might be expected that AVS is significant only in marine sediments since the concentration of sulfate in seawater is $28 \text{ mM} = 2700 \text{ mg SO}_4/\text{L}$. By contrast average river water sulfate concentration is $0.12 \text{ mM} = 11.5 \text{ mg SO}_4/\text{L}$ [Stumm and Morgan, 1981]. However sedimentary organic matter is present in either locale and the sulfate in freshwater may be sufficient to produce a significant quantity of AVS. This is confirmed by the observations reported in Table 4. Surprisingly large values are found for sediments from the Great Lakes, rivers and other freshwater lakes. The magnitudes are nearly $1 \mu\text{mol/gm}$ to more than $100 \mu\text{mol/gm}$. This strongly suggests that the AVS concentration in freshwater sediments must be considered when addressing cadmium and other metal toxicity.

D. Vertical and Temporal AVS Profiles

The normal method for sediment preparation in sediment bioassays is to produce a uniform distribution of chemical and sediment by careful mixing. For these systems the AVS is uniformly distributed and the concentration to be used for normalization is unambiguous.

However the distribution of AVS in intact sediment cores exhibits both vertical and temporal variation over the annual cycle. Table 4 presents a summary of some observations [Aller 1980; Reaves, 1984]. There is a seasonal variation in the surface concentration of AVS at the Long Island Sound NWC station and all stations exhibit a strong vertical gradient between the surface 1 cm. and the average of the top 10 cm.

This variation in AVS concentration makes it more difficult to decide what AVS concentration should be used in evaluating the potential toxicity of metals in natural sediments. This is in contrast to the distribution of sediment organic carbon which is more spatially uniform and temporally stable. Hence it appears that intact cores should be used for sediment toxicity testing if metal toxicity is suspected. Indigenous predators such as *Nephtys incisa* should be eliminated, however, perhaps by asphyxiation [Scott and Redmond, in press].

E. Sediment sampling and interstitial water generation

Ferrous sulfide oxidizes very rapidly in aerobic environments. For suspensions, oxidation is virtually complete within a few hours [Nelson, 1978]. We also have noted a decline in AVS for sediments that are held for a long period or are exposed to air. It is clear, therefore, that care should be taken to keep sediments anaerobic before AVS measurements or toxicity testing.

The use of elutrates as a surrogate for interstitial water is also suspect since oxidation of metal sulfides and release of soluble metals can occur. Procedures for producing large volumes of "pore" water by equilibrating suspensions of sediments must be checked for the extent of AVS oxidation that occurs.

CONCLUSIONS

It has been shown that AVS is the proper normalization parameter for cadmium toxicity in sediments. The observed amphipod mortality versus normalized cadmium concentration, $[Cd]/[AVS]$, is the same for sediments with over an order of magnitude difference in dry weight normalized cadmium LC50s. The correlation between mortality and interstitial water metal activity has also been confirmed. Although the fact that metals can form insoluble sulfides is well known, it apparently has not been recognized that FeS and MnS, quantified as AVS, is a reactive pool of *solid phase* sulfide that is available to bind with metals which have sulfide solubility parameters smaller than FeS.

Titrations of FeS and MnS with cadmium demonstrate that the displacement reaction, Eq. 4. does occur. Further, titrations of sediments with cadmium indicates that an abrupt increase of dissolved cadmium occurs when the added cadmium exceeds the measured AVS. However, these data are not as certain since AVS appears to be lost during the titration and the relationship is only approximate (Fig. 7). Nevertheless, the AVS normalized toxicity data (Fig. 9) does demonstrate that the normalization is quantitative.

Surprisingly, the AVS of freshwater sediments is in the same range as marine sediments. Therefore, AVS should also be the proper normalization for these sediments. The other sorption phases are expected to be important for low AVS sediments and for metals with large partition coefficients and water effect concentrations.

ACKNOWLEDGEMENTS

This research was sponsored by an EPA Cooperative Agreement CR812824-01 between Manhattan College and EPA Environmental Research Laboratory, Narragansett RI. The assistance and encouragement of Christopher Zarba, EPA Criteria and Standards Division; Herbert Allen, Drexel University; and our research assistants: Indra Sweeney, Paul Morgan, Clare Sydlik, Luisa Milevoj, and Christine Begley is gratefully acknowledged.

APPENDIX I

Solubility Relationships for Metal Sulfides

The behavior of iron sulfide during a titration with cadmium can be analyzed using a simplified equilibrium model of the Cd(II)-Fe(II)-S(II) system. The mass action laws for the sulfide solubilities are:

$$\gamma_{Cd^{2+}}[Cd^{2+}]\gamma_{S^{2-}}[S^{2-}] = K_{CdS} \quad (12)$$

$$\gamma_{Fe^{2+}}[Fe^{2+}]\gamma_{S^{2-}}[S^{2-}] = K_{FeS} \quad (13)$$

where $[Cd^{2+}]$, $[Fe^{2+}]$, and $[S^{2-}]$ are the molar concentrations; $\gamma_{Cd^{2+}}$, $\gamma_{Fe^{2+}}$, and $\gamma_{S^{2-}}$ are the activity coefficients; and K_{FeS} and K_{CdS} are the sulfide solubility products. The mass balance equations for total cadmium, iron(II), and sulfide are:

$$\alpha_{Cd^{2+}}[Cd^{2+}] + [CdS(s)] = [Cd]_A \quad (14)$$

$$\alpha_{Fe^{2+}}[Fe^{2+}] + [FeS(s)] = [FeS(s)]_i \quad (15)$$

$$\alpha_{S^{2-}}[S^{2-}] + [CdS(s)] + [FeS(s)] = [FeS(s)]_i \quad (16)$$

where $\alpha_{Cd^{2+}} = [\Sigma Cd(aq)]/[Cd^{2+}]$, $\alpha_{Fe^{2+}} = [\Sigma Fe(aq)]/[Fe^{2+}]$, and $\alpha_{S^{2-}} = [\Sigma S(aq)]/[S^{2-}]$

are the ratios of the total dissolved Cd, Fe(II), and S(II) to the divalent species concentrations, respectively. $[CdS(s)]$ and $[FeS(s)]$ are the concentration of solid phase cadmium and iron sulfide; $[FeS]_i$ is the initial iron sulfide in the sediment, and $[Cd]_A$ is the added cadmium.

The solution of these equations begins with substituting Eqs.(14) and (15) into Eq.(16). Noting that $\alpha_{S^{2-}}[S^{2-}] = [\Sigma S(aq)] \ll [Cd]_A$, which states that the total dissolved sulfide in the interstitial water is much less than the cadmium added, it follows that:

$$\gamma_{S^{2-}}[S^{2-}] \approx \frac{\alpha_{Fe^{2+}} K_{FeS} / \gamma_{Fe^{2+}} + \alpha_{Cd^{2+}} K_{CdS} / \gamma_{Cd^{2+}}}{[Cd]_A} \quad (17)$$

Then substituting Eqs. (12), (13) and (17) into Eqs.(14) and (15) yields the concentrations of solid phase sulfides:

$$[CdS(s)] \approx [Cd]_A \left(1 - \frac{\alpha_{Cd^{2+}} K_{CdS}}{\alpha_{Cd^{2+}} K_{CdS} + \alpha_{Fe^{2+}} K_{FeS}} \right) \quad (18)$$

$$[FeS(s)] \approx [FeS]_i - \frac{\alpha_{Fe^{2+}} K_{FeS}}{\alpha_{Cd^{2+}} K_{CdS} + \alpha_{Fe^{2+}} K_{FeS}} [Cd]_A \quad (19)$$

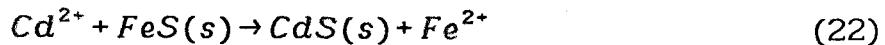
where it has been assumed that the activity coefficients for Cd^{2+} and Fe^{2+} are equal, $\gamma_{Cd^{2+}} \approx \gamma_{Fe^{2+}}$, since they are both divalent cations. The relative magnitudes of $\alpha_{Fe^{2+}} K_{FeS}$ and $\alpha_{Cd^{2+}} K_{CdS}$ determines the behavior of $[FeS(s)]$ and $[CdS(s)]$ as cadmium is added to the sediment. For this reason they are termed sulfide solubility parameters. Table 2 presents reported values. Since the cadmium solubility parameter is much less than the iron sulfide solubility parameter, i.e., $\alpha_{Cd^{2+}} K_{CdS} \ll \alpha_{Fe^{2+}} K_{FeS}$, Eqs. (18) and (19) become:

$$[CdS(s)] \approx Cd_A \quad (20)$$

and:

$$[FeS(s)] \approx [FeS]_i - Cd_A \quad (21)$$

Hence as cadmium is added to this system cadmium sulfide forms at the expense of iron sulfide. The overall reaction is:



Note that if $\alpha_{Cd^{2+}} K_{CdS} \gg \alpha_{Fe^{2+}} K_{FeS}$ then $[FeS(s)] \approx [FeS]_i$; $[CdS(s)] \approx 0$ and no cadmium sulfide would form.

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TABLE 1
Log-Logistic Parameters^(a)

Experiment	R ₀ (%)	β	LC50	Units	Fig. No.
LI Sound ^(b)	3.32	4.50	3200.	µg Cd/gm	(1)
Mixture ^(b)	"	"	1130.	"	"
Ninigret Pd ^(b)	"	"	318.	"	"
Yaquina Bay	0.0	3.34	25.5		
Water Only ^(c)					
<i>A. abdita</i>	0.0	2.34	0.34	mg Cd/L ^(d)	(2)
<i>R. hudsoni</i>	0.0	2.33	0.24	"	"
Joint ^(f)	0.0	2.12	0.29	"	"
All Sediments AVS Normalized	5.88	4.48	1.97	µmol Cd/ µmol AVS	(9)

(a) Concentration - response formula:

$$R = R_0 + \frac{100 - R_0}{1 + (LC50/c)^\beta}$$

R = mortality (%) at concentration c

R₀ = control mortality (%)

LC50 = concentration for 50% mortality

β = population sensitivity parameter

(b) The three sediments are fit assuming one value of R₀ and β.

(c) Water only exposures - no sediment present in the exposure vessels. Exposure for 96 hrs.

(d) Divide by 20 to obtain mg Cd²⁺/L.

(e) Data from Swartz et al., (1985) is fit to the log-logistic function.

(f) Joint fit of the *Ampelisca abdita* and *Rhepoxynius hudsoni* water only exposure data.

TABLE 2
Metal Sulfide Solubility and
Ratio of Total Dissolved to Free Cation Metal Concentration

Metal Sulfide	$\log K_{sp,2}$	$\log K_{sp}$	$\log \alpha$		$\log(\alpha K_{sp})$
			pH=7.6	pH=8.2	
HgS	-38.50	-57.25	15.10	15.10	-42.15
CuS	-22.19	-40.94	0.50	0.92	-40.23
PbS	-14.67	-33.42	1.12	1.32	-32.20
CdS	-14.10	-32.85	1.50	1.50	-31.35
ZnS	-9.64	-28.39	0.12	0.14	-28.26
NiS	-9.23	-27.98	0.11	0.17	-27.84
FeS	-3.64	-22.39	0.10	0.12	-22.28
FeS(am)	-3.05	-21.80	0.10	0.12	-21.69
MnS	-0.40	-19.15	0.13	0.13	-19.02

Solubility products, $K_{sp,2}$, for the reaction: $Me^{2+} + HS^- \leftrightarrow MeS(s) + H^+$ for CdS (Greenockite), FeS(amorphous) and Mackinawite, MnS (Alabandite), and NiS (Millerite), from Emerson et al., (1983). Solubility products for CuS (Covellite), HgS (Metacinnabar), PbS (Galena), and ZnS (Wurtzite), and $pK_2 = 18.57$ for the reaction $HS^- \leftrightarrow H^+ + S^{2-}$, from Schoonen and Barnes, (1988). K_{sp} is for the reaction: $Me^{2+} + S^{2-} \leftrightarrow MeS(s)$ is computed from $\log K_{sp,2}$ and pK_2 . Ratios of total to free metal concentrations: $\alpha = [\Sigma Me(aq)]/[Me^{2+}]$, from Byrne et al., (1988) at $T = 5^\circ C$. $\log(\alpha K_{sp}) = \log \alpha + \log K_{sp}$. All logs are \log_{10} .

TABLE 3
Cadmium Binding Capacity and AVS of Sediments

Sediment	Initial AVS ($\mu\text{mol/g}$) ^(a)	Final AVS ($\mu\text{mol/g}$) ^(b)	Cd Binding Capacity ($\mu\text{mol/g}$)
Black Rock Harbor	175.	-	114.
Hudson River	12.6	-	8.58
LI Sound	15.9	13.9	4.57
Mixture	5.45	3.23	-
Ninigret Pond	2.34	0.28	1.12

(a) Average AVS of repeated measurements of the stock

(b) AVS after the sediment toxicity experiment

TABLE 4
AVS in Freshwater and Marine Sediments

Location	T°C(a)	AVS ($\mu\text{mol/gm}$)		Reference
		Depth Interval	(0 - 1 cm) (0 - 10 cm)	
Fresh Water Sediments				
Everglades peat basin	-	-	0.31 - 1.3	Altschuler et al., 1983
Lake Mendota	-	-	8.7 - 112.	Nriagu, 1968
Lake Ontario	-	11.6	27.1	Nriagu et al., 1976
Lake Erie	(W)	15.0	7.5	Matisoff et al., 1981
Marine Sediments				
Long Island Sound				Aller, 1980
NWC	3.0	0.0	8.35	
NWC	13.2	0.60	10.5	
NWC	19.0	0.097	10.3	
DEEP-1	18.5	0.62	17.4	
FOAM-1	20.0	7.50	13.3	
Sapelo Island				Reaves, 1984
Mud Flat	(W)	1.88	14.6	
Mud Flat	(S)	3.44	43.2	
Tidal Ck.	(W)	9.69	28.4	
Tidal Ck.	(S)	5.94	31.9	

(a) (W) = Winter; (S) = Summer

FIGURE CAPTIONS

Figure 1. Toxicity test results for sediments from Ninigret Pond (*Ampelisca*) Long Island Sound and the mixture (*Rhepoxynius*). Cadmium concentrations on a sediment dry weight basis.

Figure 2. Mortality versus interstitial water cadmium activity. Water only exposure data for *Ampelisca* and *Rhepoxynius*. The line is a joint fit to both data sets (Table 1). Toxicity test results for the sediments in Fig. 1. Top panel - individual data. Bottom panel - statistical summary of all the sediment interstitial water data

Figure 3. Sediment cadmium versus interstitial water cadmium activity for LI Sound sediment.

Figure 4. Metal sulfide solubility parameters for seawater: $\alpha_{M_s^{2+}} \cdot K_{M_sS}$ for the metals as indicated. pH = 7.2 to 8.2; T = 20°C (Table 2).

Figure 5. Cadmium titrations of amorphous FeS. Abscissa is cadmium added normalized by FeS initially present. Ordinate is cadmium electrode response (left panel) and total dissolved cadmium (right panel).

Figure 6. Cadmium titration of sediments: Black Rock Harbor, Long Island Sound, Hudson River, Ninigret Pond. Cadmium added per unit dry weight of sediment versus total dissolved cadmium.

Figure 7. Sediment binding capacity - from the x axis intercepts of the data in Fig. 6. versus sediment AVS concentration at the start of the titration (left). Initial AVS versus final AVS at the end of the cadmium titration.

Figure 8. AVS and cadmium concentrations for the sediments used in the toxicity test (Fig. 1). Initial and final AVS (left); initial and final cadmium (right).

Figure 9. Mortality versus AVS normalized sediment cadmium for Long Island Sound, Ninigret Pond, and a 50/50 volume mixture.

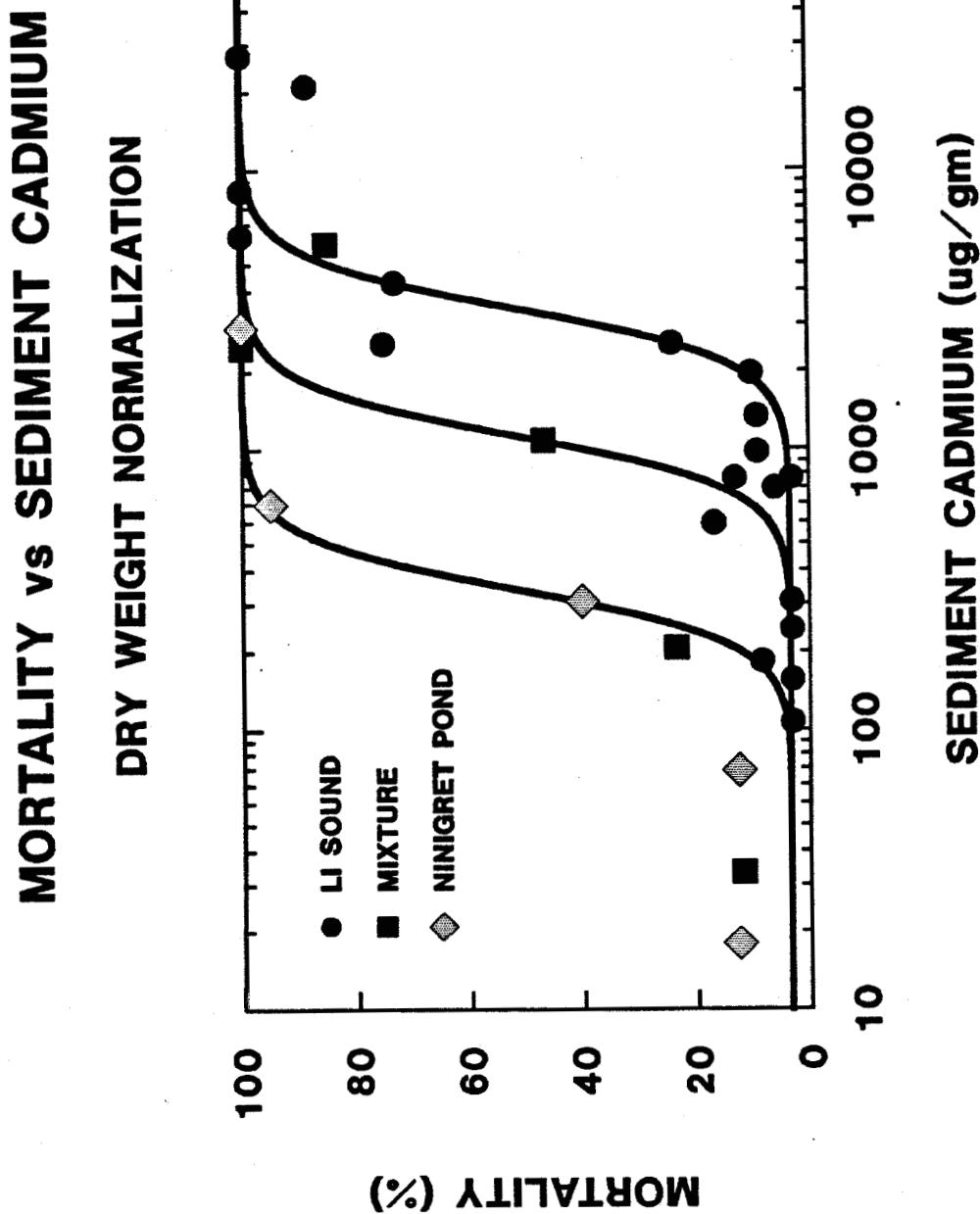


Figure 1. Toxicity test results for sediments from Ninigret Pond (*Ampelisca*) Long Island Sound and the mixture (*Rhepoxynius*). Cadmium concentrations on a sediment dry weight basis.

MORTALITY vs INTERSTITIAL WATER CADMIUM

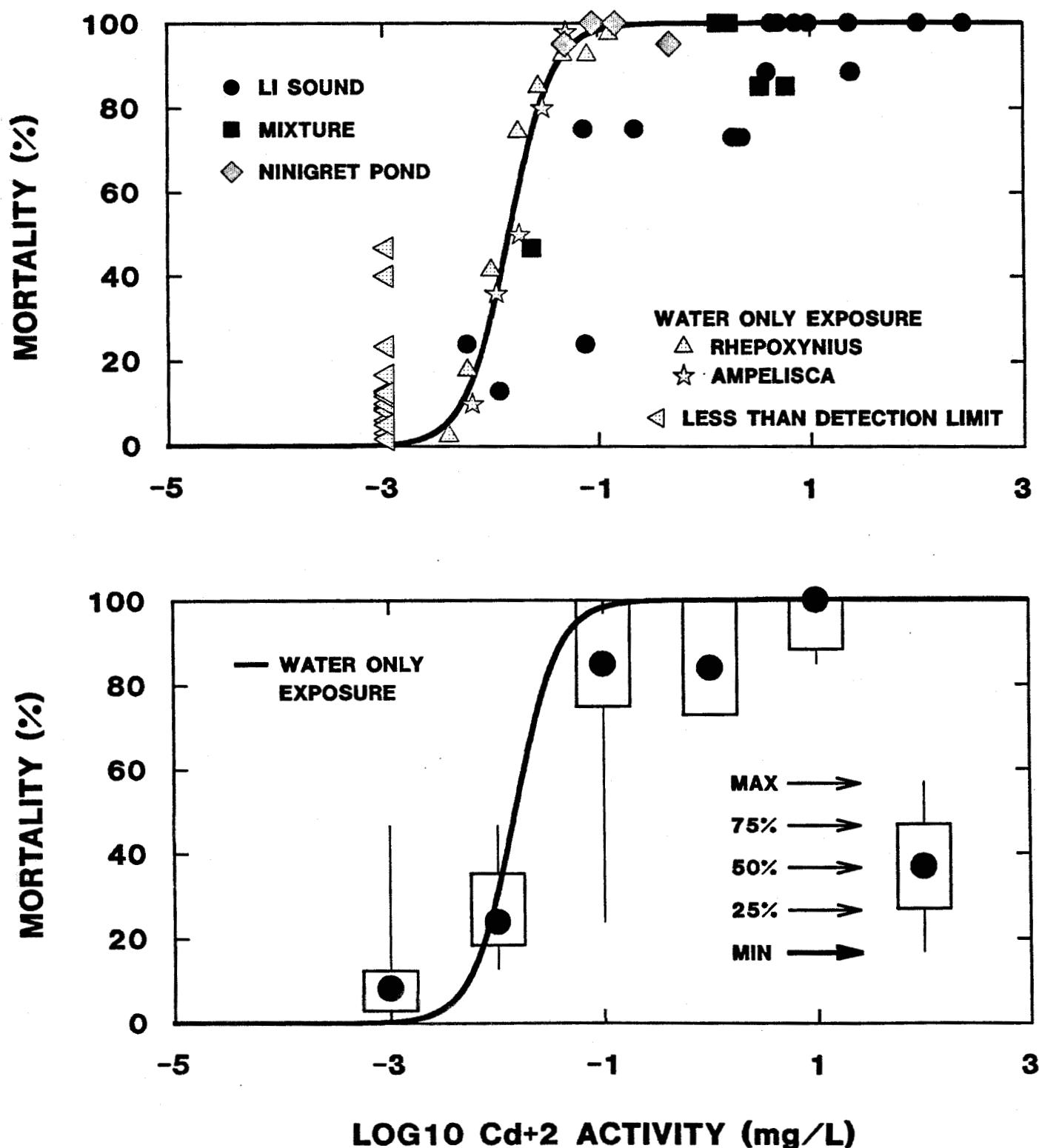


Figure 2. Mortality versus interstitial water cadmium activity. Water only exposure data for *Ampelisca* and *Rhepoxynius*. The line is a joint fit to both data sets (Table 1). Toxicity test results for the sediments in Fig. 1. Top panel - individual data. Bottom panel - statistical summary of all the sediment interstitial water data

SEDIMENT vs INTERSTITIAL WATER CADMIUM

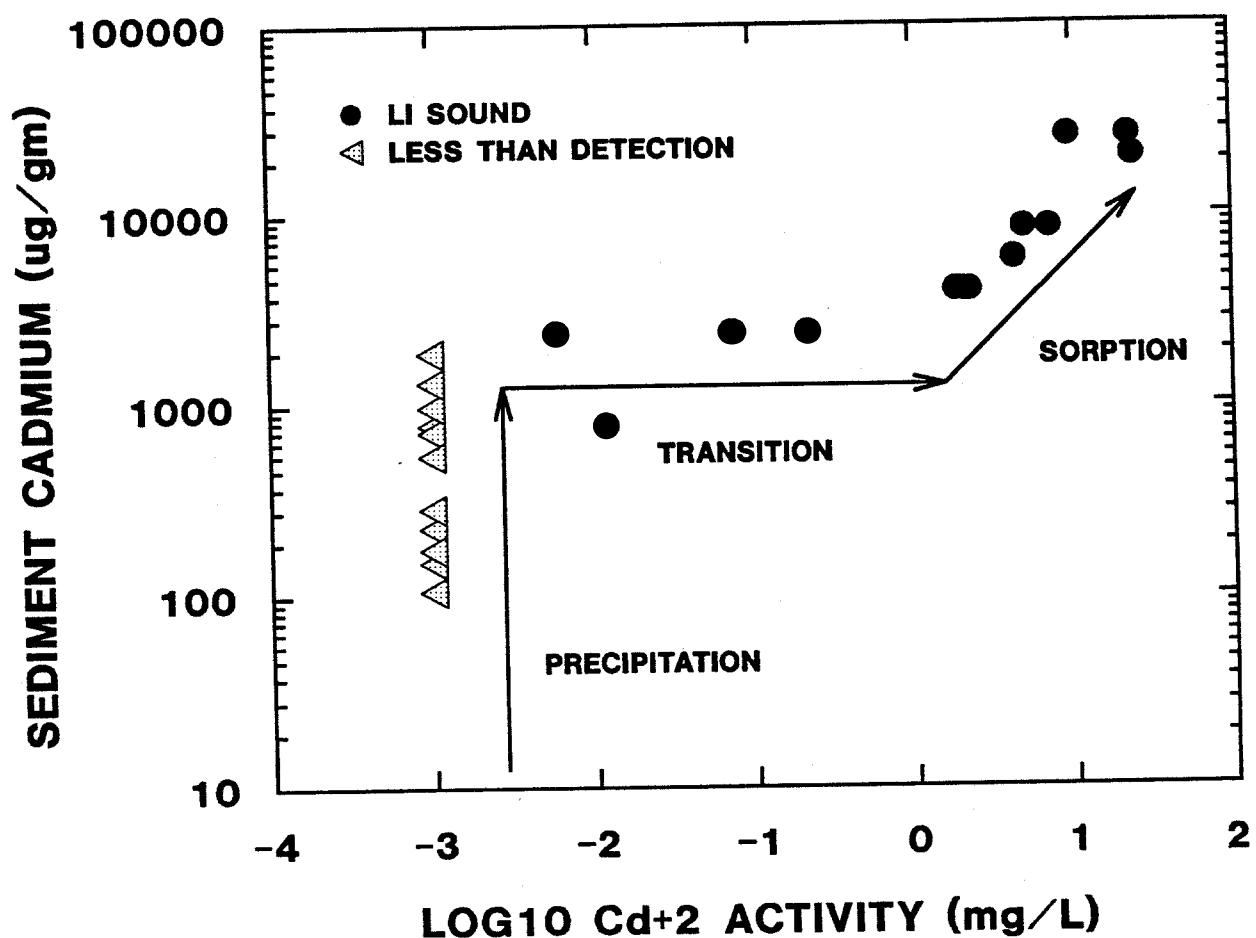


Figure 3. Sediment cadmium versus interstitial water cadmium activity for LI Sound sediment.

METAL SULFIDE SOLUBILITY PARAMETERS

SEAWATER

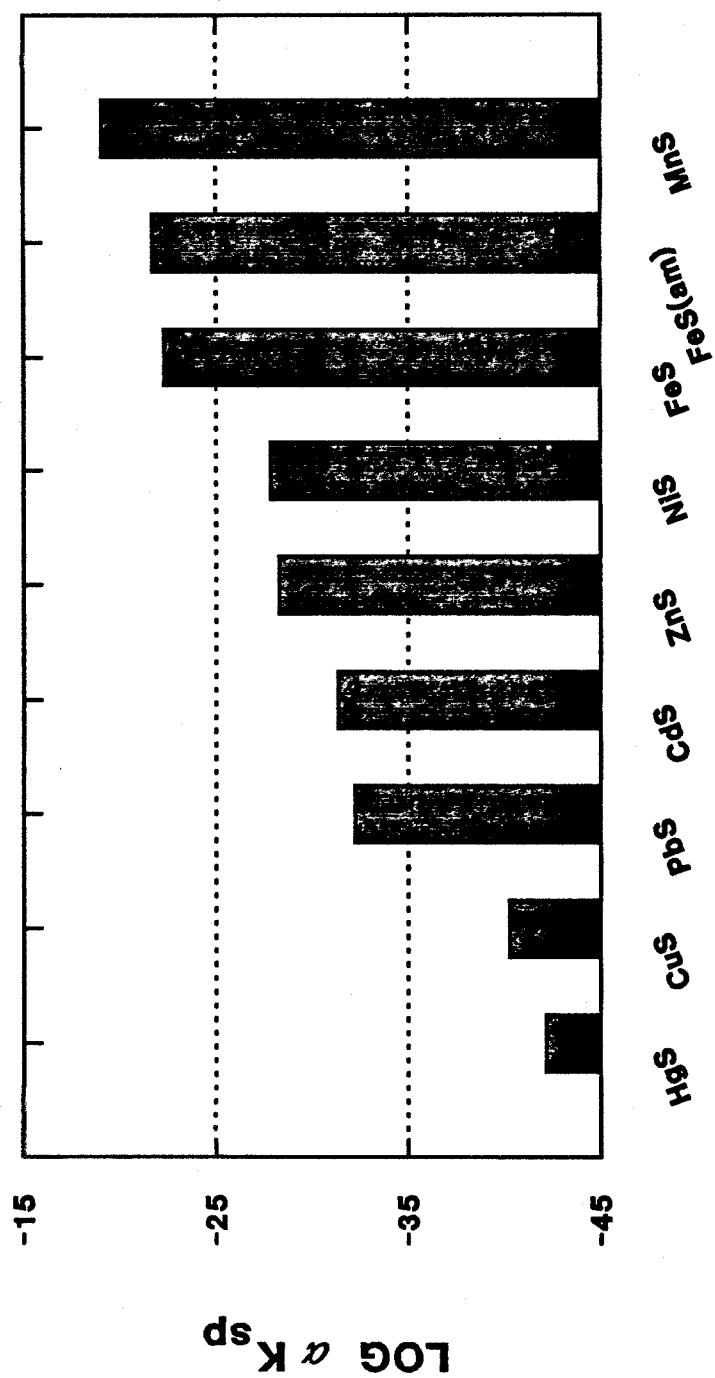


Figure 4. Metal sulfide solubility parameters for seawater: α_{K_s} , K_{M_s} for the metals as indicated.

pH = 7.2 to 8.2; T = 20°C (Table 2).

CADMIUM TITRATION OF IRON SULFIDE

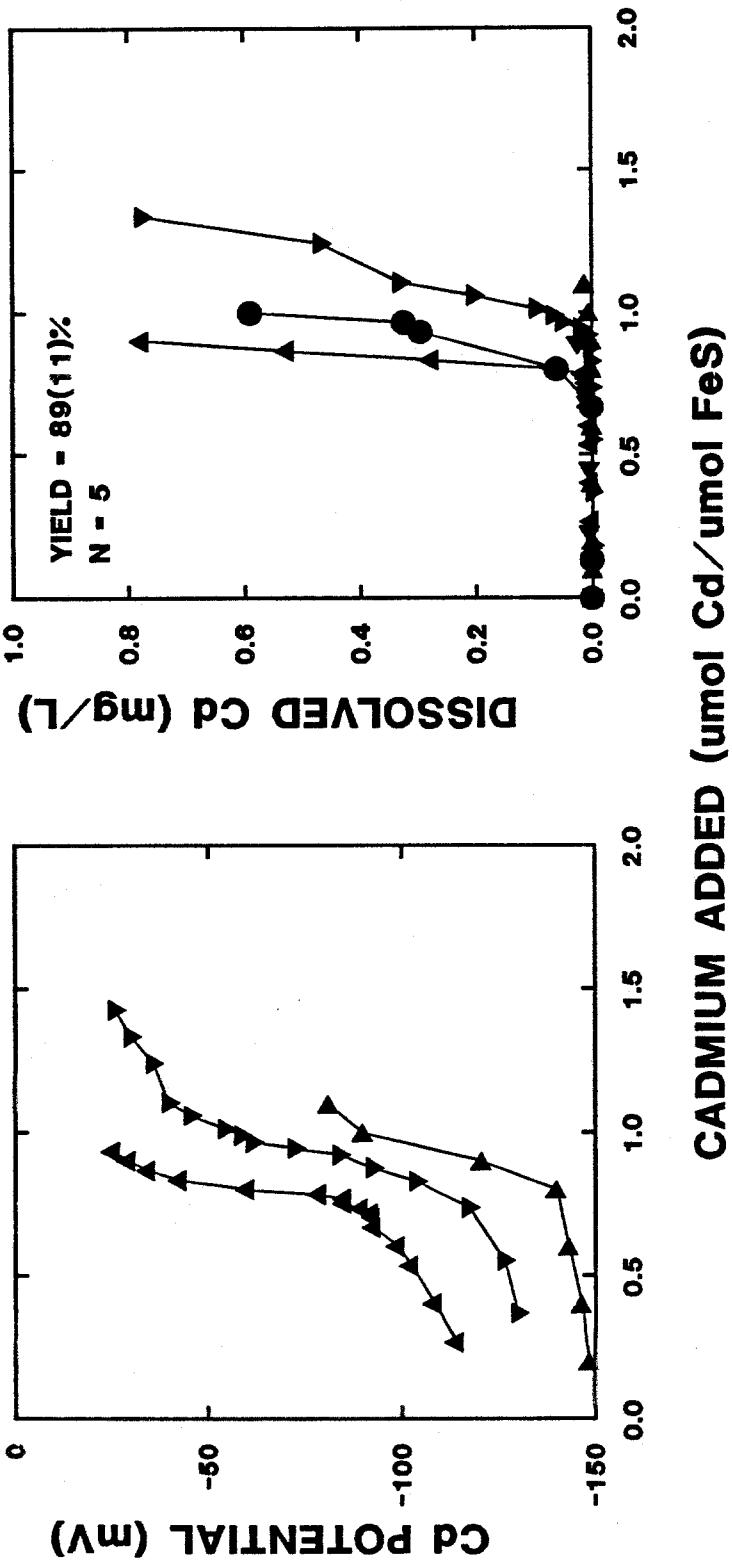


Figure 5. Cadmium titrations of amorphous FeS. Abscissa is cadmium added normalized by FeS initially present. Ordinate is cadmium electrode response (left panel) and total dissolved cadmium (right panel).

CADMIUM TITRATION OF SEDIMENTS

DRY WEIGHT NORMALIZATION

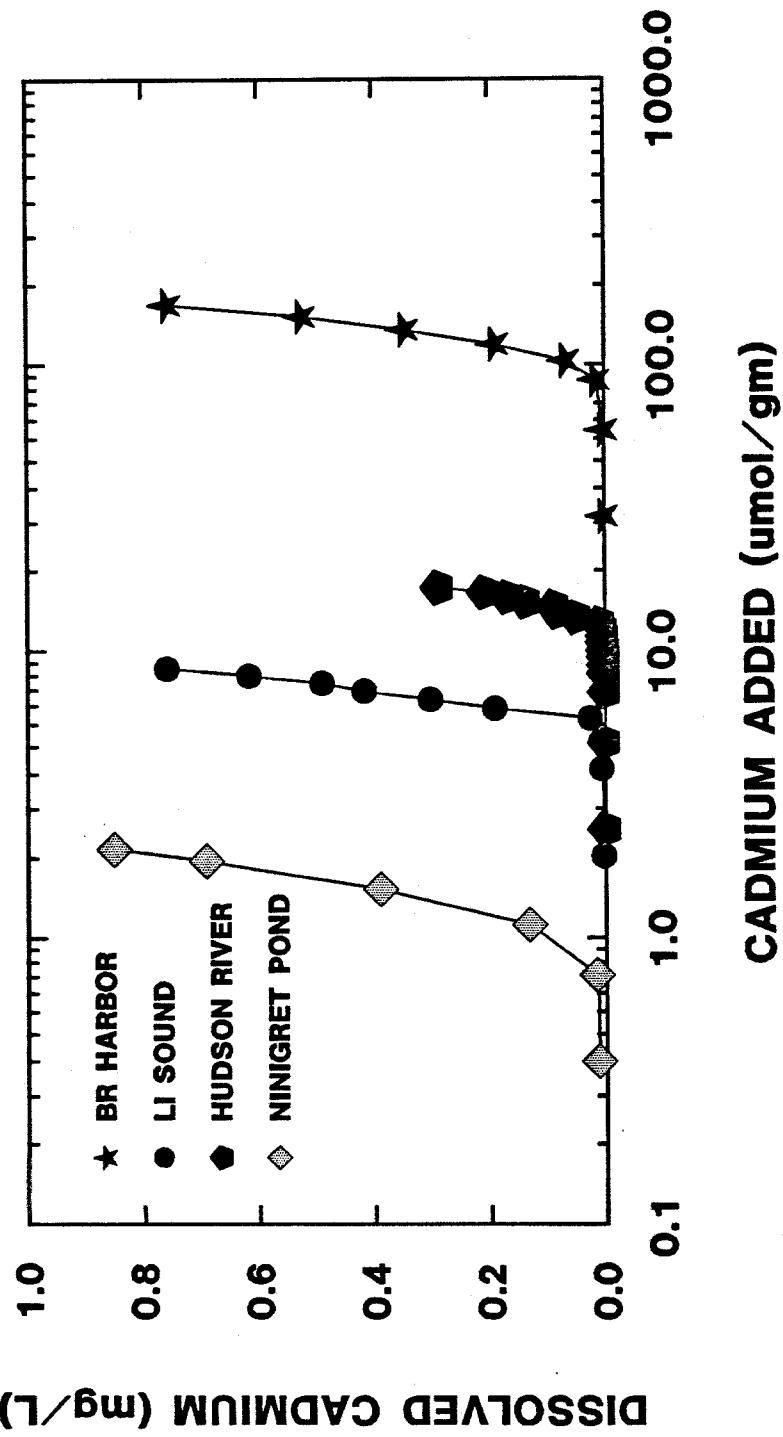


Figure 6. Cadmium titration of sediments: Black Rock Harbor, Long Island Sound, Hudson River, Ninigret Pond. Cadmium added per unit dry weight of sediment versus total dissolved cadmium.

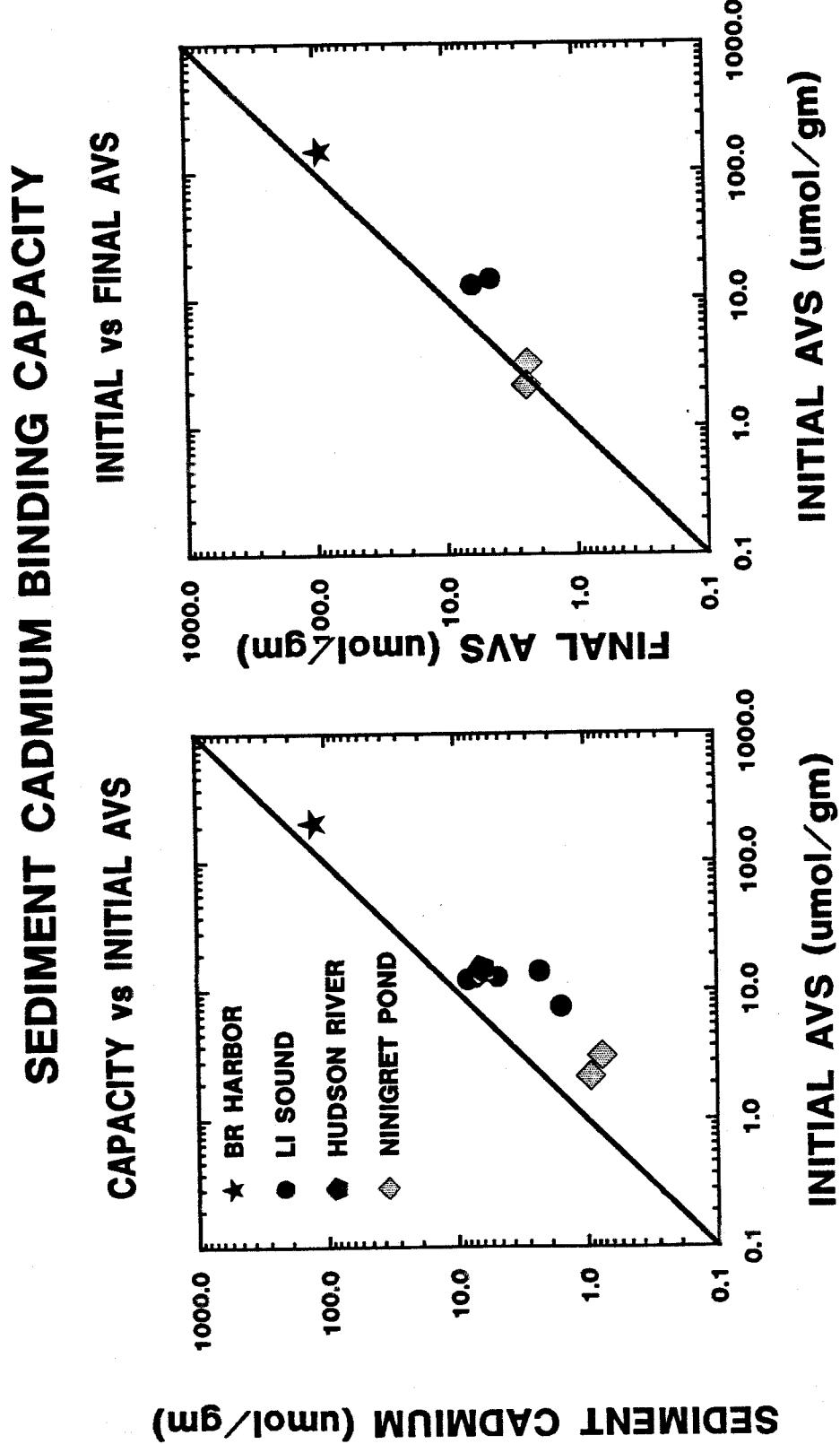
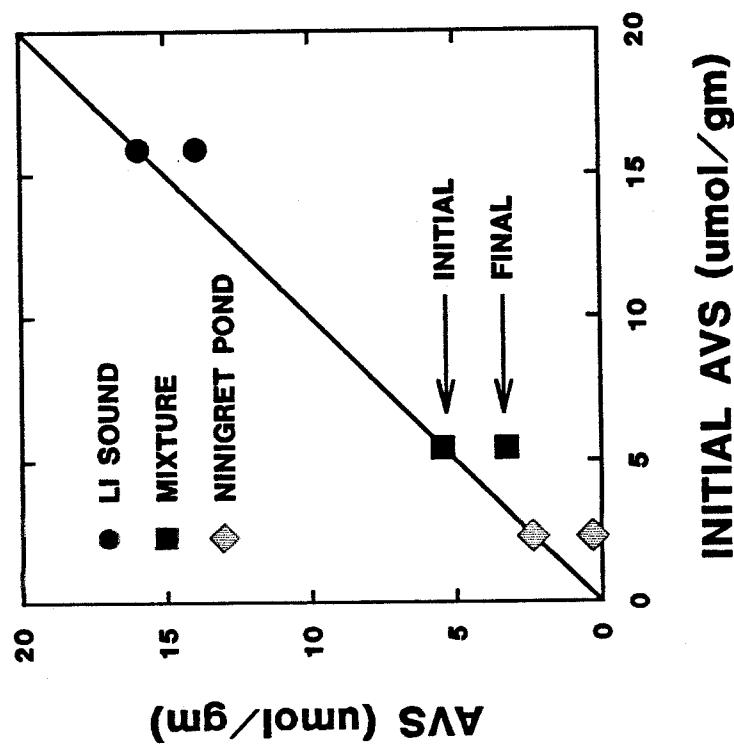


Figure 7. Sediment binding capacity - from the x axis intercepts of the data in Fig. 6. versus sediment AVS concentration at the start of the titration (left). Initial AVS versus final AVS at the end of the cadmium titration.

AVS AND CADMIUM CONCENTRATIONS

INITIAL AND FINAL AVS



INITIAL AND FINAL Cd

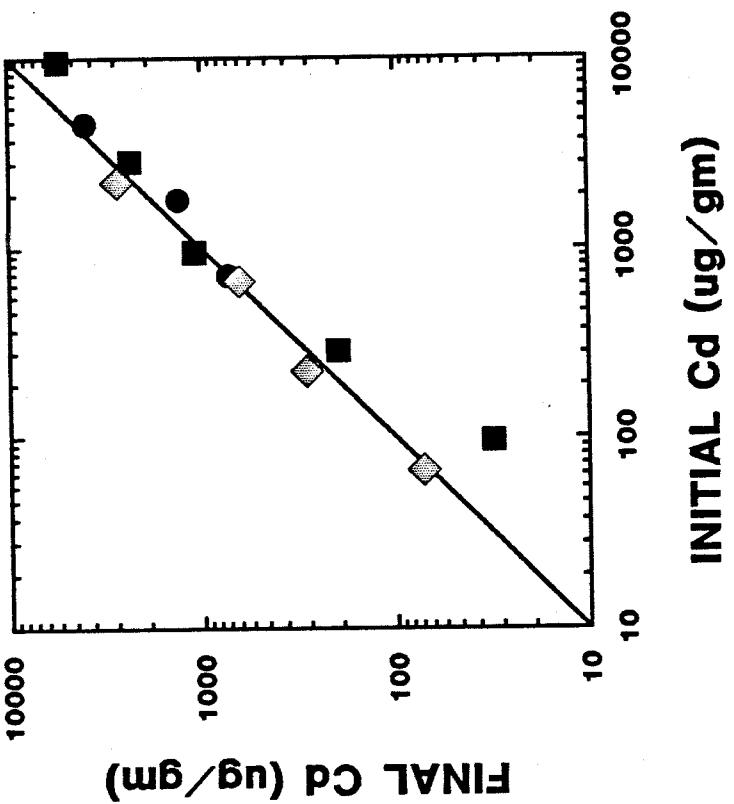


Figure 8. AVS and cadmium concentrations for the sediments used in the toxicity test (Fig. 1). Initial and final AVS (left); initial and final cadmium (right).

MORTALITY vs SEDIMENT CADMIUM
ACID VOLATILE SULFIDE NORMALIZATION

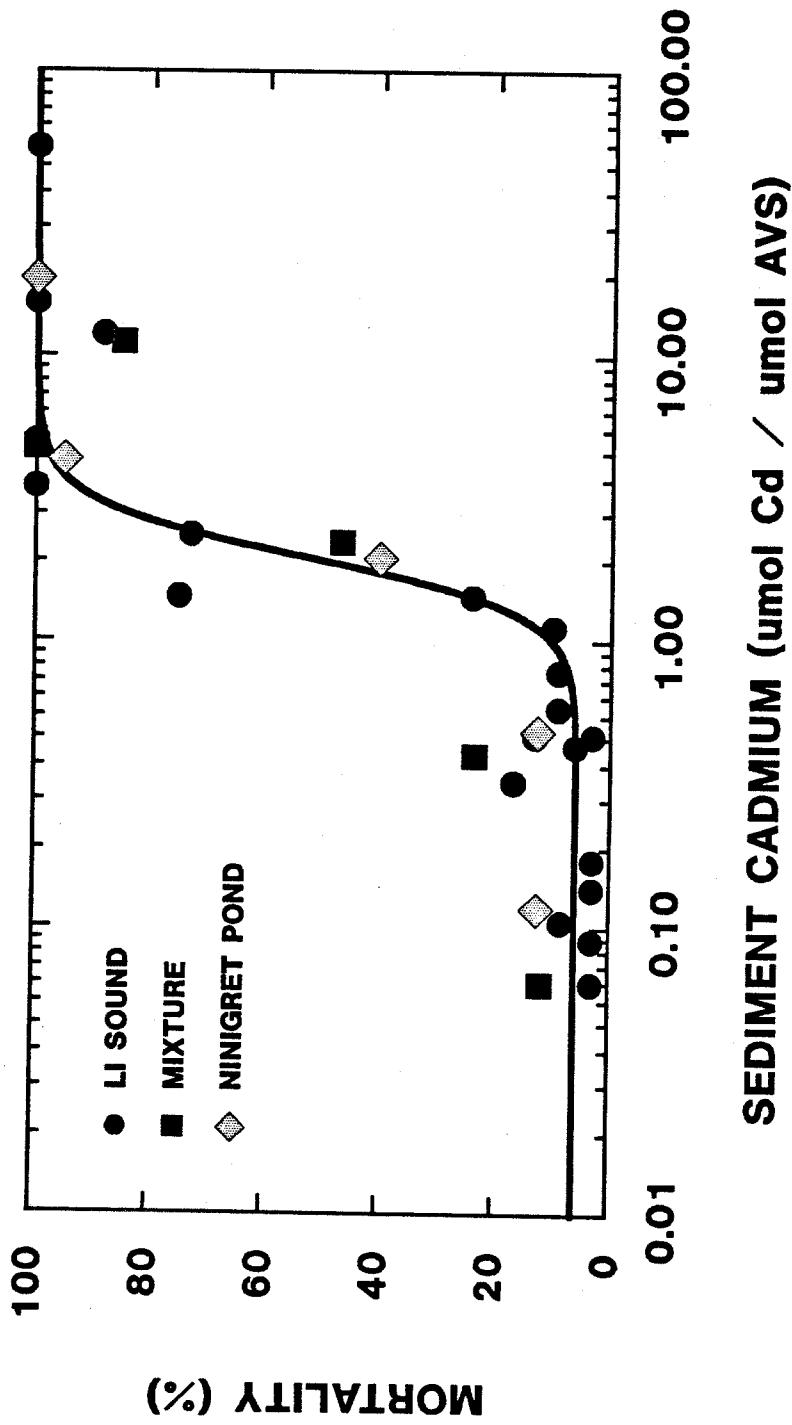


Figure 9. Mortality versus AVS normalized sediment cadmium for Long Island Sound, Ninigret Pond, and a 50/50 volume mixture.

APPENDIX II

DATA TABLES

Table 1
Final Data for Toxicity Experiments

Expt.#	Sediment	r (ug/g)	[Cd]/[AVS] (mol/mol)	Dissolved Cd(a)			Mortality (%)
				1 cm	4 cm	centrif.(b)	
1.000	LI Sound	0.0	0.0	0.001	0.001	.	3.000
1.000	"	106.748	0.064	0.001	0.001	.	3.000
1.000	"	150.995	0.090	0.001	0.001	.	3.000
1.000	"	229.153	0.137	0.001	0.001	.	3.000
1.000	"	288.237	0.172	0.001	0.001	.	3.000
1.000	"	789.775	0.472	0.001	0.001	.	3.000
2.000	"	0.0	0.0	0.001	0.001	.	3.000
2.000	"	789.775	0.472	0.012	0.001	.	13.000
2.000	"	2396.442	1.431	0.222	0.074	.	75.000
2.000	"	8423.713	5.030	7.210	5.030	.	100.000
2.000	"	25560.363	15.262	22.890	9.580	.	100.000
2.000	"	89846.995	53.648	268.260	101.380	.	100.000
3.000	"	0.0	0.0	0.001	0.001	.	6.000
3.000	"	726.606	0.434	0.001	0.001	.	6.000
3.000	"	980.801	0.586	0.001	0.001	.	9.000
3.000	"	1315.596	0.786	0.001	0.001	.	9.000
3.000	"	2396.442	1.431	0.006	0.076	.	24.000
3.000	"	3961.468	2.365	1.880	2.280	.	73.000
4.000	"	0.0	0.0	0.001	.	0.001	1.65
4.000	"	175.533	0.105	0.001	.	0.001	8.350
4.000	"	544.771	0.325	0.001	.	0.001	16.700
4.000	"	1872.224	1.118	0.001	.	0.001	10.000
4.000	"	5810.512	3.469	4.330	.	1.420	100.000
4.000	"	19969.073	11.924	24.200	.	3.940	88.400
4.000	Mixture	0.000	0.000	0.001	.	0.001	16.700
4.000	"	31.078	0.064	0.001	.	0.001	11.700
4.000	"	196.491	0.403	0.001	.	0.001	23.400
4.000	"	1082.707	2.219	0.001	.	0.024	46.700
4.000	"	2325.815	4.768	1.740	.	1.350	100.000
4.000	"	5443.609	11.159	5.900	.	3.400	85.000
4.000	Ninigret	0.000	0.000	0.000	.	0.000	5.000
4.000	"	17.195	0.117	0.001	.	0.001	12.500
4.000	"	71.788	0.488	0.001	.	0.001	12.500
4.000	"	287.150	1.950	0.001	.	0.001	40.000
4.000	"	638.191	4.334	0.475	.	0.050	95.000
4.000	"	2727.925	18.527	0.146	.	0.090	100.000

(a) Detection limit = 0.001 mg Cd⁺²/L

(b) Centrifugate from the sediment of the chemical control sediment

Table 2
Toxicity Tests - Water Only Exposure

Organism	Dissolved Cd (mg Cd ²⁺ /L)	Mortality (%)
<i>Ampelisca</i>	0.000	2.000
	" 0.007	10.000
	" 0.011	36.000
	" 0.018	50.000
	" 0.031	80.000
	" 0.051	98.000
<i>Rhepoxynius</i>	0.000	0.000
	" 0.010	2.500
	" 0.015	17.900
	" 0.025	41.500
	" 0.045	74.400
	" 0.070	85.000
	" 0.120	92.500
	" 0.200	92.500
	" 0.320	97.500

Table 2
Toxicity Tests - Water Only Exposure

Organism	Dissolved Cd (mg Cd ²⁺ /L)	Mortality (%)
<i>Ampelisca</i>	0.000	2.000
	" 0.007	10.000
	" 0.011	36.000
	" 0.018	50.000
	" 0.031	80.000
	" 0.051	98.000
<i>Rhepoxynius</i>	0.000	0.000
	" 0.004	2.500
	" 0.006	17.900
	" 0.010	41.500
	" 0.018	74.400
	" 0.028	85.000
	" 0.048	92.500
	" 0.080	92.500
	" 0.128	97.500

TABLE 3
Chemistry and Toxicity Data for Sediment Toxicity Tests

Variable Identification and Units for Data Table

EXPT	Experiment number
SED\$	Sediment ID
DEPTH	Depth of peeper sampling. Depth = 0 corresponds to water only exposures.
CDCL2	Concentration of CdCl ₂ in the sediment mixture (g CdCl ₂ /L) based on the initial weighed sample.
CDCL2H ₂ O	Concentration of CdCl ₂ · 2.5H ₂ O in the sediment mixture (g CdCl ₂ · 2.5H ₂ O/L) based on the initial weighed sample.
CT	Concentration of Cd in the sediment mixture (g Cd/L) based on the initial weighed sample.
CT_OBS	Observed concentration of Cd in the sediment mixture (g Cd/L)
CT_FNL	Final concentration of Cd in the sediment mixture (g Cd/L). CT_FNL = CT_OBS if it was measured. If not then $CT_FNL = 10^{(-0.1568 + 1.028 \log 10(CT))}$ which is the regression of log CT_OBS versus log CT.
M	Concentration of sediment solids in the sediment mixture (g/L).
R	Cd concentration on the solids based on CT (μg/gm). R = CT/M
R_OBS	Cd concentration on the solids based on CT_OBS (μg/gm). R = CT_OBS/M
R_FNL	Cd concentration on the solids based on CT_FNL (μg/gm). R = CT_FNL/M
RM	Molar Cd concentration on the solids based on CT_FNL (μmol/gm). R = CT_FNL/M/112.4
CD	Dissolved cadmium activity (mg Cd ²⁺ /L) for the water only experiments (EXPT = 0).
CD_PEEP	Dissolved cadmium activity (mg Cd ²⁺ /L) for the peeper cavities within the sediment. All cavities in the overlying water were below detection.

CD_CENTR	Dissolved cadmium activity ($\text{mg Cd}^{2+}/\text{L}$) for the centrifugate separated from the sediment in the chemical controls at the end of the experiment.
LCD_PEEP	$\text{Log}_{10}(\text{CD_PEEP})$
LCD_CENT	$\text{Log}_{10}(\text{CD_CEMTR})$
AVS1	Final AVS at the end of the experiment ($\mu\text{mol/gm}$)
AVS2	Initial AVS at the start of the experiment ($\mu\text{mol/gm}$)
CD_AVIS1	Sediment cadmium concentration normalized by AVS1. $\text{CD_AVS1} = \text{R_FNL/AVS1}$
LCD_AVIS1	$\text{Log}_{10}(\text{CD_AVS1})$
CD_AVIS2	Sediment cadmium concentration normalized by AVS2. $\text{CD_AVS2} = \text{R_FNL/AVS2}$
LCD_AVIS2	$\text{Log}_{10}(\text{CD_AVS2})$
CD_AVIS	Sediment cadmium concentration normalized by AVS. $\text{CD_AVS} = \text{R_FNL/AVS}$
LCD_AVIS	$\text{Log}_{10}(\text{CD_AVS})$
MORT	Organism mortality (%)
SURVIVOR	Organism survival (%). $\text{SURVIVOR} = 100 - \text{MORT}$

CDFNL4

	CD_AV_S	LCD_AV_S	MORT	SURVIVOR
CASE 1	2.0000	98.0000		
CASE 2	10.0000	90.0000		
CASE 3	36.0000	64.0000		
CASE 4	50.0000	50.0000		
CASE 5	20.0000	2.0000		
CASE 6	80.0000	97.0000		
CASE 7	98.0000	97.0000		
CASE 8	35.0000	97.0000		
CASE 9	0.0637	1.1956		
CASE 10	0.0902	-1.0450		
CASE 11	0.0902	-1.0450		
CASE 12	0.1368	-0.8638		
CASE 13	0.1368	-0.8638		
CASE 14	0.1368	-0.7642		
CASE 15	0.1721	-0.3264		
CASE 16	0.4716	-0.3264		
CASE 17	0.4716	-0.3264		
CASE 18	0.4716	-0.3264		
CASE 19	0.4716	-0.3264		
CASE 20	0.4716	-0.3264		
CASE 21	0.4716	-0.3264		
CASE 22	0.4716	-0.3264		
CASE 23	1.4309	0.1556	13.0000	87.0000
CASE 24	1.4309	0.1556	75.0000	25.0000
CASE 25	5.0298	0.7016	100.0000	0.0000
CASE 26	5.0298	0.016	100.0000	0.0000
CASE 27	15.2621	1.1836	100.0000	0.0000
CASE 28	53.6477	1.7296	100.0000	0.0000
CASE 29	53.6477	1.7296	94.0000	6.0000
CASE 30	15.2621	1.1836	91.0000	9.0000
CASE 31	0.3359	-0.3627	91.0000	9.0000
CASE 32	0.3359	-0.3627	91.0000	9.0000
CASE 33	0.5856	-0.2324	91.0000	9.0000
CASE 34	0.5856	-0.2324	91.0000	9.0000
CASE 35	0.5856	-0.1048	91.0000	9.0000
CASE 36	0.7855	-0.1048	91.0000	9.0000
CASE 37	0.7855	-0.1048	76.0000	24.0000
CASE 38	1.4309	0.1556	24.0000	24.0000
CASE 39	1.4309	0.1556	73.0000	27.0000
CASE 40	2.3654	0.3739	73.0000	27.0000
CASE 41	2.3654	0.3739	73.0000	27.0000
CASE 42	2.3654	0.3739	73.0000	27.0000
CASE 43	2.3654	0.3739	73.0000	27.0000
CASE 44	0.1048	-0.9796	8.3500	91.6500
CASE 45	0.3253	-0.8777	16.7000	83.3000
CASE 46	1.1179	0.0484	10.0000	90.0000
CASE 47	3.4695	0.5403	100.0000	0.0000
CASE 48	11.9235	1.0764	88.4000	11.6000
CASE 49	11.1591	-1.1958	16.7000	83.3000
CASE 50	0.0637	-0.3949	23.4000	76.6000
CASE 51	0.4028	-0.2463	46.7000	53.3000
CASE 52	2.2195	0.6783	100.0000	0.0000
CASE 53	4.7678	1.0476	85.0000	15.0000
CASE 54	11.1591	1.0476	85.0000	15.0000
CASE 55	0.1168	-0.9326	12.5000	87.5000
CASE 56	0.4875	-0.3120	40.0000	60.0000
CASE 57	1.9902	0.2901	95.0000	5.0000
CASE 58	4.3342	0.3369	1.2678	100.0000
CASE 59	18.5266			
CASE 60				
CASE 61				

TABLE 4
Chemistry Data for Cadmium Titrations of FeS and Sediments

Variable Identification and Units for Data Table

NUM	Expt number
NAME\$	Sediment name
CAL_CD	Calibration - cadmium concentration (mg/L)
LCAL_CD	" log10 " "
CAL_POT	Calibration - electrode potential mv
ADD_CD	Volume of cadmium stock solution added (mL)
UMCD_UMS	$\mu\text{mol Cd added} / \mu\text{mol AVS}$
POT	Dissolved total cadmium - electrode potential (mv)
LOGCD	Log10 Dissolved Cd - from electrode potential and calibration
PH	pH during the titration
CD	Dissolved total Cadmium - from electrode (mg/L)
M	Mass of sediment in the titration (gm)
V	Initial volume of seawater (mL)
CD_MOL	Cd concentration of the titrant (M)
UMCD_G	$\mu\text{mol Cd added} / \text{g sediment}$
UMCDDS_G	$\mu\text{mol dissolved Cd} / \text{sediment}$
CD_TOT	Total Cd added (mg/L)

NAME\$	NUM	CAL_CD	LCAL_CD	CAL_POT	ADD_CD	UMCD_UMS	POT	LOGCD
CASE	1.0000	BRH	BRH	-55.0000	1.0000	0.0160	-161.0000	-4.8400
CASE	1.0000	BRH	BRH	-53.8000	2.0000	0.0310	-123.8000	-3.4900
CASE	1.0000	BRH	BRH	-42.0000	3.0000	0.0660	-118.3000	-3.2900
CASE	1.0000	BRH	BRH	-40.6000	4.0000	0.0930	-121.2000	-3.0100
CASE	1.0000	BRH	BRH	-27.8000	6.0000	0.1090	-113.2000	-3.3900
CASE	1.0000	BRH	BRH	-14.8000	7.0000	0.1550	-119.3000	-3.1300
CASE	1.0000	BRH	BRH	0.2000	10.0000	0.1870	-120.3000	-3.3300
CASE	1.0000	BRH	BRH	13.9000	12.0000	0.2650	-118.3000	-3.3600
CASE	1.0000	BRH	BRH	28.5000	17.0000	0.3450	-110.2000	-3.2900
CASE	1.0000	BRH	BRH	.	22.0000	0.4210	-102.2000	-3.0100
CASE	1.0000	BRH	BRH	.	27.0000	0.4990	-102.2000	-2.7100
CASE	1.0000	BRH	BRH	.	32.0000	0.5760	-102.2000	-1.7100
CASE	1.0000	BRH	BRH	.	37.0000	0.6540	-17.5000	0.3500
CASE	1.0000	BRH	BRH	.	42.0000	0.7320	-10.9000	0.5900
CASE	1.0000	BRH	BRH	.	52.0000	0.8090	-4.2000	0.8300
CASE	1.0000	BRH	BRH	.	57.0000	0.8880	-0.3000	0.9700
CASE	1.0000	BRH	BRH	.	62.0000	0.9650	3.2000	1.0900
CASE	1.0000	BRH	BRH	.	67.0000	1.0430	5.6000	1.1800
CASE	1.0000	BRH	BRH	.	72.0000	0.0000	-116.4000	-3.2500
CASE	1.0000	BRH	BRH	.	72.0000	0.2020	-104.4000	-2.8300
CASE	1.0000	BRH	BRH	.	72.0000	0.4040	-98.4000	-2.6100
CASE	1.0000	BRH	BRH	.	72.0000	0.6060	-76.7000	-1.8200
CASE	1.0000	BRH	BRH	.	72.0000	0.7070	-58.9000	-1.1700
CASE	1.0000	BRH	BRH	.	72.0000	0.8080	-46.5000	-0.7200
CASE	1.0000	BRH	BRH	.	72.0000	0.9090	-39.4000	-0.4600
CASE	1.0000	BRH	BRH	.	72.0000	1.0110	-34.3000	-0.2800
CASE	1.0000	BRH	BRH	.	72.0000	1.1110	-29.9000	-0.1200
CASE	1.0000	BRH	BRH	.	72.0000	1.2120	-26.0000	0.0200
CASE	1.0000	BRH	BRH	.	72.0000	1.3130	-25.4000	0.0500
CASE	1.0000	BRH	BRH	.	72.0000	0.0000	-296.3000	-11.3800
CASE	1.0000	BRH	BRH	.	72.0000	0.5220	-238.2000	-9.3700
CASE	1.0000	BRH	BRH	.	72.0000	0.8220	-19.8000	-8.5700
CASE	1.0000	BRH	BRH	.	72.0000	0.8970	-206.7000	-8.0100
CASE	1.0000	BRH	BRH	.	72.0000	0.9660	-189.5000	-7.2600
CASE	1.0000	BRH	BRH	.	72.0000	1.0340	-100.9000	-3.4400
CASE	1.0000	BRH	BRH	.	72.0000	1.1030	-74.8000	-2.3100
CASE	1.0000	BRH	BRH	.	72.0000	1.3780	-48.4000	-1.1700
CASE	1.0000	BRH	BRH	.	72.0000	1.6550	-38.4000	-0.7300
CASE	1.0000	BRH	BRH	.	72.0000	0.0000	-297.4000	-11.1100
CASE	1.0000	BRH	BRH	.	72.0000	0.1330	-254.7000	-9.3700
CASE	1.0000	BRH	BRH	.	72.0000	0.6670	-127.5000	-4.2100
CASE	1.0000	BRH	BRH	.	72.0000	0.8010	-53.7000	-1.2200
CASE	1.0000	BRH	BRH	.	72.0000	0.9350	-36.7000	-0.5300
CASE	1.0000	BRH	BRH	.	72.0000	0.9670	-35.5000	-0.4900
CASE	1.0000	BRH	BRH	.	72.0000	1.0000	-29.3000	-0.2300
CASE	1.0000	BRH	BRH	.	72.0000	0.0000	-324.6000	-10.4100
CASE	1.0000	BRH	BRH	.	72.0000	0.2670	-114.3000	-3.0700
CASE	1.0000	BRH	BRH	.	72.0000	0.4010	-108.6000	-2.8700
CASE	1.0000	BRH	BRH	.	72.0000	0.5330	-102.6000	-2.6600
CASE	1.0000	BRH	BRH	.	72.0000	0.6910	-99.1000	-2.5300
CASE	1.0000	BRH	BRH	.	72.0000	0.6670	-92.8000	-2.3100
CASE	1.0000	BRH	BRH	.	72.0000	0.7010	-92.1000	-2.2900
CASE	1.0000	BRH	BRH	.	72.0000	0.7170	-89.5000	-2.2100
CASE	1.0000	BRH	BRH	.	72.0000	0.7350	-85.4000	-2.0600
CASE	1.0000	BRH	BRH	.	72.0000	0.7670	-85.2000	-2.0500
CASE	1.0000	BRH	BRH	.	72.0000	1.17500	-78.8000	-1.8300
CASE	1.0000	BRH	BRH	.	72.0000	0.0000	-10.4100	-10.4100
CASE	1.0000	BRH	BRH	.	72.0000	0.2670	-114.3000	-114.3000
CASE	1.0000	BRH	BRH	.	72.0000	0.4010	-108.6000	-108.6000
CASE	1.0000	BRH	BRH	.	72.0000	0.5330	-102.6000	-102.6000
CASE	1.0000	BRH	BRH	.	72.0000	0.6910	-99.1000	-99.1000
CASE	1.0000	BRH	BRH	.	72.0000	0.6670	-92.8000	-92.8000
CASE	1.0000	BRH	BRH	.	72.0000	0.7010	-92.1000	-92.1000
CASE	1.0000	BRH	BRH	.	72.0000	0.7170	-89.5000	-89.5000
CASE	1.0000	BRH	BRH	.	72.0000	0.7350	-85.4000	-85.4000
CASE	1.0000	BRH	BRH	.	72.0000	0.7670	-85.2000	-85.2000
CASE	1.0000	BRH	BRH	.	72.0000	1.17500	-78.8000	-78.8000
CASE	1.0000	BRH	BRH	.	72.0000	0.0000	-10.4100	-10.4100
CASE	1.0000	BRH	BRH	.	72.0000	0.2670	-114.3000	-114.3000
CASE	1.0000	BRH	BRH	.	72.0000	0.4010	-108.6000	-108.6000
CASE	1.0000	BRH	BRH	.	72.0000	0.5330	-102.6000	-102.6000
CASE	1.0000	BRH	BRH	.	72.0000	0.6910	-99.1000	-99.1000
CASE	1.0000	BRH	BRH	.	72.0000	0.6670	-92.8000	-92.8000
CASE	1.0000	BRH	BRH	.	72.0000	0.7010	-92.1000	-92.1000
CASE	1.0000	BRH	BRH	.	72.0000	0.7170	-89.5000	-89.5000
CASE	1.0000	BRH	BRH	.	72.0000	0.7350	-85.4000	-85.4000
CASE	1.0000	BRH	BRH	.	72.0000	0.7670	-85.2000	-85.2000
CASE	1.0000	BRH	BRH	.	72.0000	1.17500	-78.8000	-78.8000
CASE	1.0000	BRH	BRH	.	72.0000	0.0000	-10.4100	-10.4100
CASE	1.0000	BRH	BRH	.	72.0000	0.2670	-114.3000	-114.3000
CASE	1.0000	BRH	BRH	.	72.0000	0.4010	-108.6000	-108.6000
CASE	1.0000	BRH	BRH	.	72.0000	0.5330	-102.6000	-102.6000
CASE	1.0000	BRH	BRH	.	72.0000	0.6910	-99.1000	-99.1000
CASE	1.0000	BRH	BRH	.	72.0000	0.6670	-92.8000	-92.8000
CASE	1.0000	BRH	BRH	.	72.0000	0.7010	-92.1000	-92.1000
CASE	1.0000	BRH	BRH	.	72.0000	0.7170	-89.5000	-89.5000
CASE	1.0000	BRH	BRH	.	72.0000	0.7350	-85.4000	-85.4000
CASE	1.0000	BRH	BRH	.	72.0000	0.7670	-85.2000	-85.2000
CASE	1.0000	BRH	BRH	.	72.0000	1.17500	-78.8000	-78.8000
CASE	1.0000	BRH	BRH	.	72.0000	0.0000	-10.4100	-10.4100
CASE	1.0000	BRH	BRH	.	72.0000	0.2670	-114.3000	-114.3000
CASE	1.0000	BRH	BRH	.	72.0000	0.4010	-108.6000	-108.6000
CASE	1.0000	BRH	BRH	.	72.0000	0.5330	-102.6000	-102.6000
CASE	1.0000	BRH	BRH	.	72.0000	0.6910	-99.1000	-99.1000
CASE	1.0000	BRH	BRH	.	72.0000	0.6670	-92.8000	-92.8000
CASE	1.0000	BRH	BRH	.	72.0000	0.7010	-92.1000	-92.1000
CASE	1.0000	BRH	BRH	.	72.0000	0.7170	-89.5000	-89.5000
CASE	1.0000	BRH	BRH	.	72.0000	0.7350	-85.4000	-85.4000
CASE	1.0000	BRH	BRH	.	72.0000	0.7670	-85.2000	-85.2000
CASE	1.0000	BRH	BRH	.	72.0000	1.17500	-78.8000	-78.8000
CASE	1.0000	BRH	BRH	.	72.0000	0.0000	-10.4100	-10.4100
CASE	1.0000	BRH	BRH	.	72.0000	0.2670	-114.3000	-114.3000
CASE	1.0000	BRH	BRH	.	72.0000	0.4010	-108.6000	-108.6000
CASE	1.0000	BRH	BRH	.	72.0000	0.5330	-102.6000	-102.6000
CASE	1.0000	BRH	BRH	.	72.0000	0.6910	-99.1000	-99.1000
CASE	1.0000	BRH	BRH	.	72.0000	0.6670	-92.8000	-92.8000
CASE	1.0000	BRH	BRH	.	72.0000	0.7010	-92.1000	-92.1000
CASE	1.0000	BRH	BRH	.	72.0000	0.7170	-89.5000	-89.5000
CASE	1.0000	BRH	BRH	.	72.0000	0.7350	-85.4000	-85.4000
CASE	1.0000	BRH	BRH	.	72.0000	0.7670	-85.2000	-85.2000
CASE	1.0000	BRH	BRH	.	72.0000	1.17500	-78.8000	-78.8000
CASE	1.0000	BRH	BRH	.	72.0000	0.0000	-10.4100	-10.4100
CASE	1.0000	BRH	BRH	.	72.0000	0.2670	-114.3000	-114.3000
CASE	1.0000	BRH	BRH	.	72.0000	0.4010	-108.6000	-108.6000
CASE	1.0000	BRH	BRH	.	72.0000	0.5330	-102.6000	-102.6000
CASE	1.0000	BRH	BRH	.	72.0000	0.6910	-99.1000	-99.1000
CASE	1.0000	BRH	BRH	.	72.0000	0.6670	-92.8000	-92.8000
CASE	1.0000	BRH	BRH	.	72.0000	0.7010	-92.1000	-92.1000
CASE	1.0000	BRH	BRH	.	72.0000	0.7170	-89.5000	-89.5000
CASE	1.0000	BRH	BRH	.	72.0000	0.7350	-85.4000	-85.4000
CASE	1.0000	BRH	BRH	.	72.0000	0.7670	-85.2000	-85.2000
CASE	1.0000	BRH	BRH	.	72.0000	1.17500	-78.8000	-78.8000
CASE	1.0000	BRH	BRH	.	72.0000	0.0000	-10.4100	-10.4100
CASE	1.0000	BRH	BRH	.	72.0000	0.2670	-114.3000	-114.3000
CASE	1.0000	BRH	BRH	.	72.0000	0.4010	-108.6000	-108.60

NAME	NUM	CAL_CD	LCAL_CD	CAL_POT	ADD_CD	UMCD_IMS	POT	LOGD
CASE	121	1.0000	.	.	-44.0000	-44.0000	-46.5000	-0.6800
CASE	122	1.0000	2.0000	.	-42.6000	-42.6000	-42.5000	-0.5400
CASE	123	2.0000	2.0000	.	-41.0000	-41.0000	-41.3000	-3.4100
CASE	124	0.0300	-1.5200	.	-38.8000	-38.8000	-38.4000	-2.3800
CASE	125	0.1000	-1.0000	.	-33.8000	-33.8000	-33.5000	-2.1200
CASE	126	0.3000	-0.5200	.	-26.4000	-26.4000	-26.0000	-2.0600
CASE	127	1.0000	0.0000	.	-12.5000	-12.5000	-12.0000	-1.7500
CASE	128	3.0000	2.0000	.	-12.5000	-12.5000	-12.0000	-1.5600
CASE	129	10.0000	2.0000	.	-2.6000	-2.6000	-1.4900	-1.4900
CASE	130	30.0000	1.4700	.	16.1000	16.1000	-0.8200	-0.8200
CASE	131	100.0000	2.0000	.	30.5000	30.5000	-0.5200	-0.5200
CASE	132	-0.2900	-0.2900
CASE	133	-0.2100	-0.2100
CASE	134	-0.1400	-0.1400
CASE	135	-0.1100	-0.1100
CASE	136	-0.0700	-0.0700
CASE	137	-0.0100	-0.0100
CASE	138	0.0200	0.0200
CASE	139	0.1700	0.1700
CASE	140	0.0100	0.0300	3.0000	-48.0000	-48.0000	-64.6000	-0.6800
CASE	141	0.1000	0.5200	3.0000	-43.2000	-43.2000	-51.5000	-1.5600
CASE	142	0.3000	1.0000	3.0000	-38.4000	-38.4000	-46.9000	-2.5400
CASE	143	1.0000	0.0000	3.0000	-28.9000	-28.9000	-35.7000	-3.5400
CASE	144	3.0000	0.4700	3.0000	-19.7000	-19.7000	-25.0000	-2.0600
CASE	145	10.0000	1.0000	3.0000	-14.5000	-14.5000	-16.8000	-1.6800
CASE	146	30.0000	1.4700	3.0000	-10.0000	-10.0000	-12.0000	-1.2000
CASE	147	100.0000	2.0000	3.0000	-2.0000	-2.0000	-4.0000	-0.6000
CASE	148	-0.4200	-0.4200
CASE	149	-0.3370	-0.3370
CASE	150	-0.2420	-0.2420
CASE	151	-0.1900	-0.1900
CASE	152	-0.1100	-0.1100
CASE	153	-0.0700	-0.0700
CASE	154	-0.0100	-0.0100
CASE	155	0.0100	0.0300	3.0000	-42.6000	-42.6000	-53.5000	-1.9200
CASE	156	0.1000	0.5200	3.0000	-37.8000	-37.8000	-46.1000	-4.5100
CASE	157	0.3000	1.0000	3.0000	-28.7000	-28.7000	-35.3000	-3.6500
CASE	158	1.0000	0.0000	3.0000	-20.7000	-20.7000	-27.7000	-2.7800
CASE	159	3.0000	0.4700	3.0000	-12.1000	-12.1000	-17.9000	-2.5800
CASE	160	10.0000	1.0000	3.0000	-10.4000	-10.4000	-13.1000	-2.2400
CASE	161	30.0000	1.4700	3.0000	-10.5000	-10.5000	-13.2000	-2.2500
CASE	162	100.0000	2.0000	3.0000	-2.0000	-2.0000	-4.0000	-0.6000
CASE	163	-0.4770	-0.4770
CASE	164	-0.3760	-0.3760
CASE	165	-0.2480	-0.2480
CASE	166	-0.1900	-0.1900
CASE	167	-0.1100	-0.1100
CASE	168	-0.0700	-0.0700
CASE	169	-0.0100	-0.0100
CASE	170	-0.0370	-0.0370
CASE	171	-0.0100	-0.0100
CASE	172	-0.0340	-0.0340
CASE	173	-0.0160	-0.0160
CASE	174	-0.0340	-0.0340
CASE	175	-0.0100	-0.0100
CASE	176	-0.0340	-0.0340
CASE	177	-0.0160	-0.0160
CASE	178	-0.0340	-0.0340
CASE	179	-0.0100	-0.0100
CASE	180	-0.0340	-0.0340

	NAME\$	NUM	CAL_CD	LCAL_CD	CAL_POT	ADD_CD	UMCD_UMS	POT	LOGCD
CASE	181	3.0000	LIB	1.4700	14.6000	0.7000	0.4910	-26.1000	0.0100
CASE	182	3.0000	LIB	2.0000	29.6000	0.8000	0.5610	-24.7000	0.0500
CASE	183	3.0000	LIB	0.0100	-2.0000	-47.2000	0.0000	-23.5000	0.0900
CASE	184	3.0000	LIB	0.0300	-4.5200	0.2000	0.1470	-21.9000	0.1600
CASE	185	4.0000	LIB	0.1000	-1.0000	-45.0000	4.0000	-2.9500	-3.7300
CASE	186	4.0000	LIB	0.3000	-0.5200	-38.4000	5.0000	-88.8000	-2.3700
CASE	187	4.0000	LIB	1.0000	0.0000	-28.3000	6.5000	-82.0000	-2.3700
CASE	188	4.0000	LIB	3.0000	0.4700	-15.8000	7.0000	-76.7000	-2.1100
CASE	189	4.0000	LIB	10.0000	1.0000	-1.5000	7.5000	-58.7000	-1.2100
CASE	190	4.0000	LIB	30.0000	1.4700	11.3000	8.0000	-51.7000	-0.9500
CASE	191	4.0000	LIB	100.0000	2.0000	26.9000	8.5000	-45.5000	-0.7100
CASE	192	4.0000	LIB	0.0100	-2.0000	-4.7000	9.0000	-41.8000	-0.5700
CASE	193	4.0000	LIB	0.0300	-0.5200	-48.8000	10.5000	-36.8000	-0.3800
CASE	194	4.0000	LIB	1.0000	0.0000	-41.8000	11.5000	-35.0000	-0.3100
CASE	195	4.0000	LIB	3.0000	0.4700	-11.2000	7.5000	-53.0000	-0.2700
CASE	196	4.0000	LIB	10.0000	1.0000	-1.3000	8.0000	-41.8000	-0.2700
CASE	197	4.0000	LIB	30.0000	1.4700	1.3000	8.5000	-37.9000	-0.2700
CASE	198	4.0000	LIB	100.0000	2.0000	16.6000	9.0000	-29.0000	-0.2700
CASE	199	5.0000	LIB	0.0100	-2.0000	-48.8000	5.0000	-27.0000	-0.1600
CASE	200	5.0000	LIB	0.0300	-0.5200	-41.8000	6.0000	-132.0000	-0.1500
CASE	201	5.0000	LIB	1.0000	0.0000	-23.9000	7.0000	-1.7300	-2.7100
CASE	202	5.0000	LIB	3.0000	0.4700	-11.2000	7.5000	-37.0000	-0.5400
CASE	203	5.0000	LIB	10.0000	1.0000	-1.3000	8.0000	-32.9000	-0.5400
CASE	204	5.0000	LIB	30.0000	1.4700	1.3000	8.5000	-29.0000	-0.5400
CASE	205	5.0000	LIB	100.0000	2.0000	16.6000	9.0000	-29.0000	-0.5400
CASE	206	5.0000	LIB	0.0100	-2.0000	-41.8000	5.0000	-27.0000	-0.5400
CASE	207	5.0000	LIB	0.0300	-0.5200	-45.5000	6.0000	-53.9000	-0.5400
CASE	208	5.0000	LIB	1.0000	0.0000	-37.8000	7.0000	-78.9000	-0.5400
CASE	209	6.0000	LIB	0.0100	-2.0000	-50.4000	0.5000	-100.7000	-0.6200
CASE	210	6.0000	LIB	0.0300	-1.5200	-45.5000	2.0000	-7.7300	-0.6200
CASE	211	6.0000	LIB	1.0000	0.0000	-37.8000	3.0000	-31.2000	-0.6200
CASE	212	6.0000	LIB	3.0000	0.4700	-12.6000	3.5000	-4.0000	-0.6200
CASE	213	6.0000	LIB	10.0000	1.0000	-1.8000	4.0000	0.4900	-0.6200
CASE	214	6.0000	LIB	30.0000	1.4700	1.60000	4.50000	-25.3000	-0.6200
CASE	215	6.0000	LIB	100.0000	2.0000	16.6000	5.0000	-22.7000	-0.6200
CASE	216	6.0000	LIB	0.0100	-2.0000	-47.2000	5.5000	-19.1000	-0.6200
CASE	217	6.0000	LIB	0.0300	-1.5200	-45.5000	6.0000	-17.2000	-0.6200
CASE	218	6.0000	LIB	1.0000	0.0000	-37.8000	7.0000	-15.5000	-0.6200
CASE	219	6.0000	LIB	3.0000	0.4700	-1.8000	7.5000	-8.8270	-0.6200
CASE	220	6.0000	LIB	10.0000	1.0000	-1.3000	8.0000	-8.8270	-0.6200
CASE	221	6.0000	LIB	30.0000	1.4700	1.3000	8.5000	-13.9000	-0.6200
CASE	222	6.0000	LIB	100.0000	2.0000	16.6000	9.0000	-134.0000	-0.6200
CASE	223	6.0000	LIB	0.0100	-2.0000	-48.6000	0.5000	-109.6000	-0.6200
CASE	224	6.0000	LIB	0.0300	-1.5200	-44.9000	1.0000	-100.9000	-0.6200
CASE	225	7.0000	LIB	0.1000	-1.0000	-50.5000	1.5000	-0.2510	-2.7000
CASE	226	7.0000	LIB	0.3000	-0.5200	-39.5000	2.5000	-0.3010	-2.7000
CASE	227	7.0000	LIB	1.0000	0.0000	-28.4000	3.5000	-0.3510	-2.7000
CASE	228	7.0000	LIB	3.0000	0.4700	-16.0000	4.0000	-0.4010	-2.7000
CASE	229	7.0000	LIB	10.0000	1.0000	-1.6000	4.5000	-0.4510	-2.7000
CASE	230	7.0000	LIB	30.0000	1.4700	1.7000	5.0000	-0.5010	-2.7000
CASE	231	7.0000	LIB	100.0000	2.0000	26.3000	5.5000	-0.5510	-2.7000
CASE	232	7.0000	LIB	0.0100	-2.0000	-49.2000	0.5000	-30.5000	-0.6010
CASE	233	7.0000	LIB	0.0300	-1.5200	-48.6000	1.0000	-62.4000	-0.6010
CASE	234	7.0000	LIB	1.0000	0.0000	-28.4000	1.5000	-46.9000	-0.6010
CASE	235	7.0000	LIB	3.0000	0.4700	-1.6000	2.0000	-39.5000	-0.6010
CASE	236	7.0000	LIB	10.0000	1.0000	-1.7000	2.5000	-0.6510	-2.7000
CASE	237	7.0000	LIB	30.0000	1.4700	1.7000	3.0000	-0.7010	-2.7000
CASE	238	7.0000	LIB	100.0000	2.0000	26.3000	3.5000	-0.7510	-2.7000
CASE	239	7.0000	LIB	0.0100	-2.0000	-49.2000	0.5000	-34.3000	-0.6010
CASE	240	7.0000	LIB	0.0300	-1.5200	-48.6000	1.0000	-62.4000	-0.6010

	NAME\$	CAL_CD	LCAI_CD	CAL_POT	ADD_CD	UNCD_WMS	POT	LOGED
CASE	241	9.0000	9.0000	-56.9000	-3.8200	-3.4600	-122.4000	-3.2300
CASE	242	9.0000	9.0000	-55.3000	-3.4600	-3.1600	-115.7000	-3.0900
CASE	243	9.0000	9.0000	-48.5000	-3.2300	-3.0900	-112.1000	-2.9300
CASE	244	9.0000	9.0000	-37.9000	-2.9300	-2.8000	-107.4000	-2.6900
CASE	245	9.0000	9.0000	-26.7000	-2.6900	-2.5000	-100.7000	-2.4000
CASE	246	9.0000	9.0000	-17.4000	-2.4000	-2.2000	-97.7000	-2.2000
CASE	247	9.0000	9.0000	-1.2000	-2.2000	-2.0000	-70.7000	-1.6300
CASE	248	9.0000	9.0000	14.9000	-1.6300	-1.4500	-62.5000	-1.3400
CASE	249	9.0000	9.0000	28.9000	-1.3400	-1.0000	-47.8000	-0.8200
CASE	250	9.0000	9.0000	55.9000	-0.8200	-0.5000	-41.2000	-0.5800
CASE	251	9.0000	9.0000	65.0000	-0.5800	-0.2500	-36.1000	-0.4100
CASE	252	9.0000	9.0000	70.0000	-0.4100	-0.2500	-32.0000	-0.2500
CASE	253	9.0000	9.0000	75.0000	-0.2500	-0.2500	-27.2000	-0.2500
CASE	254	10.0000	10.0000	-53.1000	0.0000	0.0000	-99.0000	0.0000
CASE	255	10.0000	10.0000	-53.8000	0.0000	0.0000	-80.5000	0.0000
CASE	256	10.0000	10.0000	-63.8000	0.0000	0.0000	-67.8000	0.0000
CASE	257	10.0000	10.0000	-57.0000	0.0000	0.0000	-47.3000	0.0000
CASE	258	10.0000	10.0000	-52.5000	0.0000	0.0000	-36.0000	0.0000
CASE	259	10.0000	10.0000	-0.7000	0.0000	0.0000	-36.0000	0.0000
CASE	260	10.0000	10.0000	-12.7000	0.0000	0.0000	-36.0000	0.0000
CASE	261	10.0000	10.0000	-2.4000	0.0000	0.0000	-30.2000	0.0000
CASE	262	10.0000	10.0000	1.5000	0.0000	0.0000	-28.1000	0.0000
CASE	263	10.0000	10.0000	2.4000	0.0000	0.0000	-24.0000	0.0000
CASE	264	10.0000	10.0000	1.4700	0.0000	0.0000	-99.3000	0.0000
CASE	265	10.0000	10.0000	2.0000	0.0000	0.0000	-95.8000	0.0000
CASE	266	10.0000	10.0000	1.0000	0.0000	0.0000	-50.7000	0.0000
CASE	267	10.0000	10.0000	0.5000	0.0000	0.0000	-40.3000	0.0000
CASE	268	10.0000	10.0000	0.0000	0.0000	0.0000	-38.8000	0.0000
CASE	269	10.0000	10.0000	0.4700	0.0000	0.0000	-35.7000	0.0000
CASE	270	10.0000	10.0000	1.7000	0.0000	0.0000	-32.5000	0.0000
CASE	271	10.0000	10.0000	1.4700	0.0000	0.0000	-27.6000	0.0000
CASE	272	10.0000	10.0000	1.5000	0.0000	0.0000	-24.5000	0.0000
CASE	273	10.0000	10.0000	2.0000	0.0000	0.0000	-21.6000	0.0000
CASE	274	10.0000	10.0000	1.49.7000	0.0000	0.0000	-138.0000	0.0000
CASE	275	10.0000	10.0000	-40.9000	0.0000	0.0000	-105.3000	0.0000
CASE	276	10.0000	10.0000	-27.8000	0.0000	0.0000	-42.0000	0.0000
CASE	277	10.0000	10.0000	-14.7000	0.0000	0.0000	-35.7000	0.0000
CASE	278	10.0000	10.0000	-0.7000	0.0000	0.0000	-32.5000	0.0000
CASE	279	10.0000	10.0000	1.2000	0.0000	0.0000	-27.6000	0.0000
CASE	280	10.0000	10.0000	1.4700	0.0000	0.0000	-24.5000	0.0000
CASE	281	10.0000	10.0000	1.5000	0.0000	0.0000	-21.6000	0.0000
CASE	282	10.0000	10.0000	1.5000	0.0000	0.0000	-18.7000	0.0000
CASE	283	10.0000	10.0000	2.0000	0.0000	0.0000	-13.8000	0.0000
CASE	284	10.0000	10.0000	2.0000	0.0000	0.0000	-10.9000	0.0000
CASE	285	10.0000	10.0000	2.0000	0.0000	0.0000	-8.0000	0.0000
CASE	286	10.0000	10.0000	2.0000	0.0000	0.0000	-6.1000	0.0000
CASE	287	10.0000	10.0000	2.0000	0.0000	0.0000	-4.2000	0.0000
CASE	288	10.0000	10.0000	2.0000	0.0000	0.0000	-2.3000	0.0000
CASE	289	10.0000	10.0000	2.0000	0.0000	0.0000	-0.5000	0.0000
CASE	290	10.0000	10.0000	2.0000	0.0000	0.0000	-0.5000	0.0000
CASE	291	10.0000	10.0000	2.0000	0.0000	0.0000	-0.5000	0.0000
CASE	292	10.0000	10.0000	2.0000	0.0000	0.0000	-0.5000	0.0000
CASE	293	10.0000	10.0000	2.0000	0.0000	0.0000	-0.5000	0.0000
CASE	294	10.0000	10.0000	2.0000	0.0000	0.0000	-0.5000	0.0000
CASE	295	10.0000	10.0000	2.0000	0.0000	0.0000	-0.5000	0.0000
CASE	296	10.0000	10.0000	2.0000	0.0000	0.0000	-0.5000	0.0000
CASE	297	10.0000	10.0000	2.0000	0.0000	0.0000	-0.5000	0.0000
CASE	298	10.0000	10.0000	2.0000	0.0000	0.0000	-0.5000	0.0000
CASE	299	10.0000	10.0000	2.0000	0.0000	0.0000	-0.5000	0.0000
CASE	300	10.0000	10.0000	2.0000	0.0000	0.0000	-0.5000	0.0000

NAME\$	NUM	CAL_CD	LCAL_CD	CAL_POT	CAL_CD	ADD_CD	UMCD	UMS	POT	LOGD
SS	30.0000	1.4700	13.9000	29.3000	6.0000	6.0000	-19.6000	0.0200	-26.2000	0.0200
SS	100.0000	2.0000	.	.	-41.5000	0.5000	-18.6000	0.0800	-22.6000	0.0800
SS	304	0.1000	-2.0000	-41.5000	0.5000	0.0000	-11.8000	0.1400	-20.9000	0.1400
CASE	305	0.0500	-1.5200	-38.5000	1.0000	0.0410	-11.4000	-4.1000	-11.8000	-4.1000
CASE	306	0.1000	-1.0000	-32.6000	1.5000	0.0810	-11.8000	-3.8400	-12.0000	-3.8400
CASE	307	0.3000	-0.5200	-28.7000	2.0000	0.1210	-12.3000	-4.1900	-12.0000	-4.1900
CASE	308	0.3000	0.4700	-10.0000	2.5000	0.2430	-11.5000	-3.8900	-11.6000	-3.8900
CASE	309	0.3000	0.4700	14.0000	3.0000	0.2830	-12.7000	-2.2400	-12.0000	-2.2400
CASE	310	0.3000	1.0000	17.3000	3.5000	0.3240	-13.0000	-1.7600	-13.0000	-1.7600
CASE	311	0.3000	1.4700	32.0000	4.0000	0.3630	-14.4000	-1.4800	-14.0000	-1.4800
CASE	312	0.3000	1.4700	32.0000	4.5000	0.4040	-15.0000	-1.3900	-15.0000	-1.3900
CASE	313	0.3000	1.4700	32.0000	5.0000	0.4450	-15.5000	-1.2900	-15.5000	-1.2900
CASE	314	0.3000	1.4700	32.0000	5.5000	0.4850	-16.0000	-1.2700	-16.0000	-1.2700
CASE	315	0.3000	1.4700	32.0000	6.0000	0.5260	-16.5000	-1.2700	-16.5000	-1.2700
CASE	316	0.3000	1.4700	32.0000	6.5000	0.5660	-17.0000	-1.2700	-17.0000	-1.2700
CASE	317	0.3000	1.4700	32.0000	7.0000	0.6000	-17.5000	-1.2700	-17.5000	-1.2700
CASE	318	0.3000	1.4700	32.0000	7.5000	0.1010	-18.0000	-1.2700	-18.0000	-1.2700
CASE	319	0.3000	1.4700	32.0000	8.0000	-2.0000	-18.5000	-0.3700	-18.5000	-0.3700
CASE	320	0.3000	1.4700	32.0000	8.5000	-51.4000	-19.0000	-0.3700	-19.0000	-0.3700
CASE	321	0.3000	1.4700	32.0000	9.0000	-50.7000	-19.5000	-0.3700	-19.5000	-0.3700
CASE	322	0.3000	1.4700	32.0000	9.5000	-49.0000	-20.0000	-0.3700	-20.0000	-0.3700
CASE	323	0.3000	1.4700	32.0000	10.0000	-47.3000	-20.5000	-0.3700	-20.5000	-0.3700
CASE	324	0.3000	1.4700	32.0000	10.5000	-45.6000	-21.0000	-0.3700	-21.0000	-0.3700
CASE	325	0.3000	1.4700	32.0000	11.0000	-43.9000	-21.5000	-0.3700	-21.5000	-0.3700
CASE	326	0.3000	1.4700	32.0000	11.5000	-42.2000	-22.0000	-0.3700	-22.0000	-0.3700
CASE	327	0.3000	1.4700	32.0000	12.0000	-40.5000	-22.5000	-0.3700	-22.5000	-0.3700
CASE	328	0.3000	1.4700	32.0000	12.5000	-38.8000	-23.0000	-0.3700	-23.0000	-0.3700
CASE	329	0.3000	1.4700	32.0000	13.0000	-37.1000	-23.5000	-0.3700	-23.5000	-0.3700
CASE	330	0.3000	1.4700	32.0000	13.5000	-35.4000	-24.0000	-0.3700	-24.0000	-0.3700
CASE	331	0.3000	1.4700	32.0000	14.0000	-33.7000	-24.5000	-0.3700	-24.5000	-0.3700
CASE	332	0.3000	1.4700	32.0000	14.5000	-32.0000	-25.0000	-0.3700	-25.0000	-0.3700
CASE	333	0.3000	1.4700	32.0000	15.0000	-30.3000	-25.5000	-0.3700	-25.5000	-0.3700
CASE	334	0.3000	1.4700	32.0000	15.5000	-28.6000	-26.0000	-0.3700	-26.0000	-0.3700
CASE	335	0.3000	1.4700	32.0000	16.0000	-26.9000	-26.5000	-0.3700	-26.5000	-0.3700
CASE	336	0.3000	1.4700	32.0000	16.5000	-25.2000	-27.0000	-0.3700	-27.0000	-0.3700
CASE	337	0.3000	1.4700	32.0000	17.0000	-23.5000	-27.5000	-0.3700	-27.5000	-0.3700
CASE	338	0.3000	1.4700	32.0000	17.5000	-21.8000	-28.0000	-0.3700	-28.0000	-0.3700
CASE	339	0.3000	1.4700	32.0000	18.0000	-20.1000	-28.5000	-0.3700	-28.5000	-0.3700
CASE	340	0.3000	1.4700	32.0000	18.5000	-18.4000	-29.0000	-0.3700	-29.0000	-0.3700
CASE	341	0.3000	1.4700	32.0000	19.0000	-16.7000	-29.5000	-0.3700	-29.5000	-0.3700
CASE	342	0.3000	1.4700	32.0000	19.5000	-15.0000	-30.0000	-0.3700	-30.0000	-0.3700
CASE	343	0.3000	1.4700	32.0000	20.0000	-13.3000	-30.5000	-0.3700	-30.5000	-0.3700
CASE	344	0.3000	1.4700	32.0000	20.5000	-11.6000	-31.0000	-0.3700	-31.0000	-0.3700
CASE	345	0.3000	1.4700	32.0000	21.0000	-10.0000	-31.5000	-0.3700	-31.5000	-0.3700
CASE	346	0.3000	1.4700	32.0000	21.5000	-8.3000	-32.0000	-0.3700	-32.0000	-0.3700
CASE	347	0.3000	1.4700	32.0000	22.0000	-6.6000	-32.5000	-0.3700	-32.5000	-0.3700
CASE	348	0.3000	1.4700	32.0000	22.5000	-5.0000	-33.0000	-0.3700	-33.0000	-0.3700
CASE	349	0.3000	1.4700	32.0000	23.0000	-3.3000	-33.5000	-0.3700	-33.5000	-0.3700
CASE	350	0.3000	1.4700	32.0000	23.5000	-1.6000	-34.0000	-0.3700	-34.0000	-0.3700

	CD TOT	UMCDDS_G	UMCD_G	CD_MOL	V	M	CD	PH
CASE 1	19.8333	38.9077	57.2604	0.0000	5.3380	9.5070	0.0001	7.5300
CASE 2	0.0000	0.0001	0.0002	0.0090	12.6761	0.0001	0.0001	7.6100
CASE 3	0.0000	0.0001	0.0002	0.0090	19.1841	0.0001	0.0001	7.7200
CASE 4	0.0000	0.0001	0.0002	0.0090	22.1831	0.0001	0.0001	7.7400
CASE 5	0.0000	0.0001	0.0002	0.0090	31.5901	0.0001	0.0001	7.7300
CASE 6	0.0000	0.0001	0.0002	0.0090	38.0282	0.0001	0.0001	7.7800
CASE 7	0.0000	0.0001	0.0002	0.0090	53.8732	0.0001	0.0001	7.8400
CASE 8	0.0000	0.0001	0.0002	0.0090	69.7183	0.0001	0.0001	7.8400
CASE 9	0.0000	0.0001	0.0002	0.0090	85.5634	0.0005	0.0005	7.8400
CASE 10	0.0000	0.0001	0.0002	0.0090	101.4085	0.0050	0.0050	7.8400
CASE 11	0.0000	0.0001	0.0002	0.0090	117.2535	0.2116	0.2116	7.8400
CASE 12	0.0000	0.0001	0.0002	0.0090	133.0986	0.6452	0.6452	7.8400
CASE 13	0.0000	0.0001	0.0002	0.0090	148.9437	1.1822	1.1822	7.8400
CASE 14	0.0000	0.0001	0.0002	0.0090	164.7887	2.1603	2.1603	7.8400
CASE 15	0.0000	0.0001	0.0002	0.0090	180.6338	3.1282	3.1282	7.8400
CASE 16	0.0000	0.0001	0.0002	0.0090	196.4789	4.3165	4.3165	7.8400
CASE 17	0.0000	0.0001	0.0002	0.0090	212.3239	5.5475	5.5475	7.8400
CASE 18	0.0000	0.0001	0.0002	0.0090	228.5794	0.0008	0.0008	7.8400
CASE 19	0.0000	0.0001	0.0002	0.0090	244.8251	0.0022	0.0022	7.8400
CASE 20	0.0000	0.0001	0.0002	0.0090	260.0708	0.0037	0.0037	7.8400
CASE 21	0.0000	0.0001	0.0002	0.0090	275.3165	0.0234	0.0234	7.8400
CASE 22	0.0000	0.0001	0.0002	0.0090	290.5622	0.1055	0.1055	7.8400
CASE 23	0.0000	0.0001	0.0002	0.0090	305.8079	0.3001	0.3001	7.8400
CASE 24	0.0000	0.0001	0.0002	0.0090	321.0536	0.5512	0.5512	7.8400
CASE 25	0.0000	0.0001	0.0002	0.0090	336.3093	0.8420	0.8420	7.8400
CASE 26	0.0000	0.0001	0.0002	0.0090	351.5550	1.2281	1.2281	7.8400
CASE 27	0.0000	0.0001	0.0002	0.0090	366.7907	1.6229	1.6229	7.8400
CASE 28	0.0000	0.0001	0.0002	0.0090	382.0364	1.7105	1.7105	7.8400
CASE 29	0.0000	0.0001	0.0002	0.0090	397.2821	1.8492	1.8492	7.8400
CASE 30	0.0000	0.0001	0.0002	0.0090	412.5278	1.91	1.91	7.8400
CASE 31	0.0000	0.0001	0.0002	0.0090	427.7735	2.0833	2.0833	7.8400
CASE 32	0.0000	0.0001	0.0002	0.0090	442.0192	2.2516	2.2516	7.8400
CASE 33	0.0000	0.0001	0.0002	0.0090	457.2649	2.4286	2.4286	7.8400
CASE 34	0.0000	0.0001	0.0002	0.0090	472.5106	2.5957	2.5957	7.8400
CASE 35	0.0000	0.0001	0.0002	0.0090	487.7563	2.7626	2.7626	7.8400
CASE 36	0.0000	0.0001	0.0002	0.0090	502.9920	2.9295	2.9295	7.8400
CASE 37	0.0000	0.0001	0.0002	0.0090	518.2377	3.0960	3.0960	7.8400
CASE 38	0.0000	0.0001	0.0002	0.0090	533.4834	3.2629	3.2629	7.8400
CASE 39	0.0000	0.0001	0.0002	0.0090	548.7291	3.4298	3.4298	7.8400
CASE 40	0.0000	0.0001	0.0002	0.0090	563.9748	3.5867	3.5867	7.8400
CASE 41	0.0000	0.0001	0.0002	0.0090	579.2205	3.7436	3.7436	7.8400
CASE 42	0.0000	0.0001	0.0002	0.0090	594.4662	3.9005	3.9005	7.8400
CASE 43	0.0000	0.0001	0.0002	0.0090	609.7119	4.0574	4.0574	7.8400
CASE 44	0.0000	0.0001	0.0002	0.0090	624.9576	4.2143	4.2143	7.8400
CASE 45	0.0000	0.0001	0.0002	0.0090	639.2033	4.3712	4.3712	7.8400
CASE 46	0.0000	0.0001	0.0002	0.0090	654.4490	4.5281	4.5281	7.8400
CASE 47	0.0000	0.0001	0.0002	0.0090	669.6948	4.6850	4.6850	7.8400
CASE 48	0.0000	0.0001	0.0002	0.0090	684.9405	4.8419	4.8419	7.8400
CASE 49	0.0000	0.0001	0.0002	0.0090	700.1862	5.0088	5.0088	7.8400
CASE 50	0.0000	0.0001	0.0002	0.0090	715.4319	5.1657	5.1657	7.8400
CASE 51	0.0000	0.0001	0.0002	0.0090	730.6776	5.3226	5.3226	7.8400
CASE 52	0.0000	0.0001	0.0002	0.0090	745.9233	5.4795	5.4795	7.8400
CASE 53	0.0000	0.0001	0.0002	0.0090	761.1690	5.6364	5.6364	7.8400
CASE 54	0.0000	0.0001	0.0002	0.0090	776.4147	5.7933	5.7933	7.8400
CASE 55	0.0000	0.0001	0.0002	0.0090	791.6604	5.9502	5.9502	7.8400
CASE 56	0.0000	0.0001	0.0002	0.0090	806.9061	6.1071	6.1071	7.8400
CASE 57	0.0000	0.0001	0.0002	0.0090	822.1518	6.2640	6.2640	7.8400
CASE 58	0.0000	0.0001	0.0002	0.0090	837.3975	6.4209	6.4209	7.8400
CASE 59	0.0000	0.0001	0.0002	0.0090	852.6432	6.5778	6.5778	7.8400

	PH	CD	M	V	CD_MOL	UMCD_G	UMCDDS_G	CD_TOT
CASE 61		0.0646						
CASE 62		0.2754						
CASE 63		0.5268						
CASE 64		0.7762						
CASE 65		1.0715						
CASE 66	8.7800	0.0000						
CASE 67	8.4100	0.0000						
CASE 68	8.1200	0.0001						
CASE 69	7.8900	0.0002						
CASE 70	7.8100	0.0004						
CASE 71	7.9400	0.0013						
CASE 72	7.7600	0.0035						
CASE 73	7.9500	0.0074						
CASE 74	7.9200	0.0195						
CASE 75	7.8700	0.0513						
CASE 76	7.8300	0.0661						
CASE 77	7.8100	0.0955						
CASE 78	7.7300	0.3311						
CASE 79	7.6300	0.4677						
CASE 80	7.5800	0.7762						
CASE 81	7.5500	1.0965						
CASE 82	6.9100	0.0000						
CASE 83	6.9600	0.0043						
CASE 84	6.5800	0.0049						
CASE 85	6.5100	0.0052						
CASE 86	6.3100	0.0245						
CASE 87	6.2900	2.0893						
CASE 88	6.2200	3.8905						
CASE 89	6.1900	6.1660						
CASE 90	6.1500	16.9824						
CASE 91	6.1100	42.6980						
CASE 92	6.0700	57.5440						
CASE 93		29.5121						
CASE 94		35.3700						
CASE 95		35.3700						
CASE 96		35.3700						
CASE 97		35.3700						
CASE 98		35.3700						
CASE 99		35.3700						
CASE 100		35.3700						
CASE 101		35.3700						
CASE 102		35.3700						
CASE 103		35.3700						
CASE 104	7.8900	0.0001						
CASE 105	8.0000	0.0004						
CASE 106	7.9500	0.0006						
CASE 107	7.8400	0.0011						
CASE 108	7.8000	0.0012						
CASE 109	7.7600	0.0014						
CASE 110	7.7500	0.0020						
CASE 111	7.7200	0.0032						
CASE 112	7.6900	0.0036						
CASE 113	7.6600	0.0049						
CASE 114	7.6100	0.0102						
CASE 115	7.5900	0.0513						
CASE 116	7.5700	0.0832						
CASE 117	7.5100	0.0891						
CASE 118	7.4800	0.1380						
CASE 119	7.4500	0.1698						
CASE 120		35.3700						

	CD	CD_MOL	UMCD_G	UMCDDDS_G	CD_TOT
121	7.4100	7.4100	7.4100	7.4100	125.1464
CASE	122	7.4800	7.4800	7.4800	128.571
CASE	123	5.8500	5.8500	5.8500	0.0000
CASE	124	5.8500	5.8500	5.8500	11.096
CASE	125	7.5700	7.6300	7.6300	21.654
CASE	126	7.6400	7.6400	7.6400	0.0002
CASE	127	7.6600	7.6600	7.6600	41.696
CASE	128	7.6500	7.6400	7.6400	0.0003
CASE	129	7.6400	7.6400	7.6400	51.099
CASE	130	7.6400	7.6400	7.6400	60.243
CASE	131	7.6200	7.6200	7.6200	69.0175
CASE	132	5.8500	5.8500	5.8500	0.0028
CASE	133	7.6000	7.5900	7.5900	0.0028
CASE	134	7.5700	7.5700	7.5700	0.0028
CASE	135	7.5700	7.5700	7.5700	0.0028
CASE	136	7.5500	7.5500	7.5500	0.0028
CASE	137	7.5500	7.5500	7.5500	0.0028
CASE	138	7.5500	7.5500	7.5500	0.0028
CASE	139	8.4800	8.4800	8.4800	0.0028
CASE	140	7.9300	7.9300	7.9300	0.0028
CASE	141	7.8500	7.8500	7.8500	0.0028
CASE	142	7.8500	7.8500	7.8500	0.0028
CASE	143	7.4500	7.4500	7.4500	0.0028
CASE	144	7.7900	7.7900	7.7900	0.0028
CASE	145	7.7400	7.7400	7.7400	0.0028
CASE	146	7.6400	7.6400	7.6400	0.0028
CASE	147	7.6000	7.5600	7.5600	0.0028
CASE	148	7.6000	7.5600	7.5600	0.0028
CASE	149	7.4500	7.4500	7.4500	0.0028
CASE	150	7.4700	7.4700	7.4700	0.0028
CASE	151	7.3300	7.3300	7.3300	0.0028
CASE	152	7.2700	7.2700	7.2700	0.0028
CASE	153	7.2700	7.2700	7.2700	0.0028
CASE	154	7.2700	7.2700	7.2700	0.0028
CASE	155	7.2700	7.2700	7.2700	0.0028
CASE	156	7.2700	7.2700	7.2700	0.0028
CASE	157	7.2700	7.2700	7.2700	0.0028
CASE	158	7.2700	7.2700	7.2700	0.0028
CASE	159	7.2700	7.2700	7.2700	0.0028
CASE	160	7.2700	7.2700	7.2700	0.0028
CASE	161	7.2700	7.2700	7.2700	0.0028
CASE	162	7.2700	7.2700	7.2700	0.0028
CASE	163	7.2700	7.2700	7.2700	0.0028
CASE	164	7.2700	7.2700	7.2700	0.0028
CASE	165	7.2700	7.2700	7.2700	0.0028
CASE	166	7.2700	7.2700	7.2700	0.0028
CASE	167	7.9200	7.9200	7.9200	0.0028
CASE	168	7.9200	7.9200	7.9200	0.0028
CASE	169	7.8900	7.8900	7.8900	0.0028
CASE	170	7.8600	7.8600	7.8600	0.0028
CASE	171	7.8300	7.8300	7.8300	0.0028
CASE	172	7.8100	7.8100	7.8100	0.0028
CASE	173	7.7800	7.7800	7.7800	0.0028
CASE	174	8.4100	8.4100	8.4100	0.0028
CASE	175	8.4400	8.4400	8.4400	0.0028
CASE	176	8.3700	8.3700	8.3700	0.0028
CASE	177	8.4000	8.4000	8.4000	0.0028
CASE	178	8.3400	8.3400	8.3400	0.0028
CASE	179	8.3300	8.3300	8.3300	0.0028
CASE	180	8.3300	8.3300	8.3300	0.0028

	CD	TOT
CASE	241	0.0000
CASE	242	12.889
CASE	243	24.672
CASE	244	36.5639
CASE	245	48.1714
CASE	246	70.5767
CASE	247	81.3951
CASE	248	102.2966
CASE	249	0.0000
CASE	250	0.0000
CASE	251	0.0001
CASE	252	0.0001
CASE	253	0.0014
CASE	254	0.0027
CASE	255	0.003
CASE	256	0.0007
CASE	257	0.0020
CASE	258	0.0037
CASE	259	0.0001
CASE	260	0.0001
CASE	261	0.0001
CASE	262	0.0001
CASE	263	0.0001
CASE	264	0.0002
CASE	265	0.0002
CASE	266	0.0003
CASE	267	0.0007
CASE	268	0.0001
CASE	269	0.0001
CASE	270	0.0002
CASE	271	0.0006
CASE	272	0.0006
CASE	273	0.0007
CASE	274	0.0007
CASE	275	0.0007
CASE	276	0.0009
CASE	277	0.0006
CASE	278	0.0009
CASE	279	0.0008
CASE	280	0.0011
CASE	281	0.0009
CASE	282	0.0012
CASE	283	0.0020
CASE	284	0.0024
CASE	285	0.0039
CASE	286	0.0019
CASE	287	0.0355
CASE	288	0.0123
CASE	289	0.0162
CASE	290	0.0219
CASE	291	0.0219
CASE	292	0.0009
CASE	293	0.0015
CASE	294	0.0004
CASE	295	0.0004
CASE	296	0.0005
CASE	297	0.0004
CASE	298	0.0004
CASE	299	0.0012
CASE	300	0.0284

TABLE 5
AVS Determinations

Variable Identification and Units for Data Table

NAME\$	Sediment identifier
DATE\$	Date of the experiment
DRYWT	Dry weight of sediment extracted
AG2S	Weight of Ag ₂ S in sulfide trap
UMS_G	µmol/gm AVS
UMS_G1	µmol/gm AVS - These were extractions after a cadmium titration was completed.
RATIO	Ratio of UMS_G1 to UMS_G

		NAME\$	DATE\$	DRYWT	AG2S	UMS_G	UMS_G1	RATIO
CASE	1	BRH	8/5/88	3.9554	0.1998	203.8500		
CASE	2	BRH	9/22/88	2.1800	0.0789	146.0600	92.4600	0.6330
CASE	3	HR	7/11/88	11.8200	0.0312	10.6500	.	.
CASE	4	HR	7/22/88	5.8500	0.0211	14.5600	.	.
CASE	5	HR1	10/11/88A	10.5300	0.0350	27.9100	.	.
CASE	6	HR1	10/13/88A	7.1300	0.0426	24.1100	.	.
CASE	7	HR1	9/27/88	13.0700	0.0340	23.9900	.	.
CASE	8	HR1	9/27/888	11.4800	0.0387	24.8800	.	.
CASE	9	LIB	7/11/88	8.9100	0.0339	15.3500	.	.
CASE	10	LIB	7/11/88	8.9200	0.0336	15.2100	.	.
CASE	11	LIB	7/13/88	6.7100	0.0237	14.2500	.	.
CASE	12	LIB	7/13/88	15.9300	0.0503	12.7500	.	.
CASE	13	LIB	7/13/88	15.8000	0.0467	11.9300	.	.
CASE	14	LIB	7/14/88	17.9200	0.0545	12.2700	.	.
CASE	15	LIB	7/15/88	7.9200	0.0279	14.2200	.	.
CASE	16	LIB	7/15/88	6.0500	0.0243	16.2100	.	.
CASE	17	LIB	7/18/88	13.0800	0.0431	12.6000	.	.
CASE	18	LIB	7/19/88	15.8000	0.0408	10.4200	.	.
CASE	19	LIB	7/7/88	11.5200	0.0400	13.9700	.	.
CASE	20	LIB	7/8/88	11.0100	0.0419	15.3500	.	.
CASE	21	LIB	8/1/88	.	.	14.4000	4.5000	0.3125
CASE	22	LIB1	7/24/88	16.2500	0.0581	14.4300	.	.
CASE	23	LIB1	8/2/88	7.1170	0.0228	12.9300	6.2100	0.4803
CASE	24	LIB1	9/20/88	12.6600	0.0238	7.5900	.	.
CASE	25	LIB1	9/23/88	10.7800	0.0305	11.4100	.	.
CASE	26	SS	7/18/88	40.0500	0.0313	3.1500	.	.
CASE	27	SS	7/19/88	41.1600	0.0302	2.9600	.	.
CASE	28	SS	7/8/88	18.7000	0.0101	2.1800	.	.
CASE	29	SS	8/3/88	11.5200	0.0090	3.1500	2.3700	0.7524
CASE	30	SS	8/4/88	15.1000	0.0079	2.1100	2.4400	1.1564
CASE	31	SS	9/23/88	25.2100	0.0199	3.1900	.	.
CASE	32	SS1	9/27/88	15.9400	0.0103	2.6100	.	.

TABLE 6
Comparison of Initial AVS and Cadmium Binding

Variable Identification and Units for Data Table

NAME\$	Sediment identifier
NUM	Expt number
DATE\$	Date of the experiment
AVS	Initial AVS at the start of the titration ($\mu\text{mol/gm}$)
CD_G	Binding capacity of the sediment ($\mu\text{mol Cd/g sediment}$)

		NAME\$	NUM	DATES	AVS	CD_G
CASE	1	BRH	1.0000	8/5/88	203.8500	122.9768
CASE	2	BRH	2.0000	9/22/88	.	105.8089
CASE	3	HR	1.0000	7/12/88	.	12.4965
CASE	4	HR	2.0000	7/22/88	14.5600	6.3564
CASE	5	HR	3.0000	10/03/88	.	7.9552
CASE	6	LIB	1.0000	7/5/88	.	6.1573
CASE	7	LIB	2.0000	7/8/88	.	5.5622
CASE	8	LIB	3.0000	7/13/88	14.2500	2.2627
CASE	9	LIB	4.0000	7/13/88	12.7500	7.1214
CASE	10	LIB	5.0000	7/14/88	12.2700	8.1084
CASE	11	LIB	6.0000	8/1/88	.	3.8762
CASE	12	LIB	7.0000	8/2/88	12.9000	4.7734
CASE	13	LIB	9.0000	8/3/88	.	6.4904
CASE	14	LIB	10.0000	9/19/88	7.5900	1.5547
CASE	15	SS	1.0000	7/8/88	2.1800	0.9381
CASE	16	SS	2.0000	7/1/88	.	2.0620
CASE	17	SS	3.0000	7/14/88	.	1.0127
CASE	18	SS	4.0000	7/18/88	3.1500	0.7589
CASE	19	SS	5.0000	8/4/88	.	1.1768

TABLE 7
AVS in Long Island Sound Sediments#

Variable Identification and Units for Data Table

ID\$	Sediment identifier
TABLE\$	Table number in the reference from which the data were taken
T	Temperature °C
DEPTH1	Starting depth of the core slice (cm)
DEPTH2	Ending depth of the core slice (cm)
DEPTH	Average depth of the core slice (cm)
FES	Measured AVS in the slice ($\mu\text{mol/g}$)

[Aller, 1980]

CASE	TABLE\$	IDS	T	DEPTH1	DEPTH2	DEPTH	FES
1	FOAM1	B1	20.0000	0.0000	1.0000	2.0000	7.5000
2	FOAM1	B1	20.0000	2.0000	3.0000	8.1000	11.0000
3	FOAM1	B1	20.0000	3.0000	4.0000	12.0000	2.5000
4	FOAM1	B1	20.0000	4.0000	5.0000	23.0000	4.5000
5	FOAM1	B1	20.0000	5.0000	6.0000	20.0000	5.5000
6	FOAM1	B1	20.0000	6.0000	7.0000	18.0000	6.5000
7	FOAM1	B1	20.0000	7.0000	8.0000	13.0000	7.2000
8	FOAM1	B1	20.0000	8.0000	9.0000	11.0000	8.4000
9	FOAM1	B1	20.0000	9.0000	10.0000	12.0000	8.5000
10	FOAM1	B1	20.0000	10.0000	11.0000	10.5000	14.5000
11	FOAM1	B1	20.0000	11.0000	12.0000	11.5000	15.0000
12	FOAM1	B1	20.0000	12.0000	13.0000	12.5000	16.5000
13	FOAM1	B1	20.0000	13.0000	14.0000	13.5000	17.5000
14	FOAM1	B1	20.0000	14.0000	15.0000	14.5000	18.0000
15	FOAM1	B1	20.0000	15.0000	16.0000	15.5000	19.0000
16	FOAM1	B1	20.0000	16.0000	17.0000	16.5000	21.0000
17	FOAM1	B1	20.0000	17.0000	18.0000	17.5000	21.5000
18	FOAM1	B1	20.0000	18.0000	19.0000	18.5000	22.0000
19	FOAM1	B1	20.0000	19.0000	20.0000	19.5000	22.5000
20	NWC-2	B7	13.0000	0.0000	1.0000	2.0000	10.6000
21	NWC-2	B7	13.0000	1.0000	2.0000	3.0000	9.3600
22	NWC-2	B7	13.0000	2.0000	3.0000	4.0000	13.7000
23	NWC-2	B7	13.0000	3.0000	4.0000	5.0000	16.5000
24	NWC-2	B7	13.0000	4.0000	5.0000	6.0000	17.5000
25	NWC-2	B7	13.0000	5.0000	6.0000	7.0000	18.5000
26	NWC-2	B7	13.0000	6.0000	7.0000	8.0000	19.5000
27	NWC-2	B7	13.0000	7.0000	8.0000	9.0000	20.5000
28	NWC-3	B8	3.0000	0.0000	1.0000	2.0000	0.0000
29	NWC-3	B8	3.0000	1.0000	2.0000	3.0000	2.1000
30	NWC-3	B8	3.0000	2.0000	3.0000	4.0000	3.1800
31	NWC-3	B8	3.0000	3.0000	4.0000	5.0000	4.1800
32	NWC-3	B8	3.0000	4.0000	5.0000	6.0000	5.1800
33	NWC-3	B8	3.0000	5.0000	6.0000	7.0000	6.1800
34	NWC-3	B8	3.0000	6.0000	7.0000	8.0000	7.1800
35	NWC-3	B8	3.0000	7.0000	8.0000	9.0000	8.1800
36	NWC-3	B8	3.0000	8.0000	9.0000	10.0000	9.1800
37	NWC-3	B8	3.0000	9.0000	10.0000	11.0000	10.1800
38	NWC-4	B9	19.0000	0.0000	1.0000	2.0000	16.9000
39	NWC-4	B9	19.0000	1.0000	2.0000	3.0000	17.9000
40	NWC-4	B9	19.0000	2.0000	3.0000	4.0000	18.6000
41	NWC-4	B9	19.0000	3.0000	4.0000	5.0000	19.3000
42	NWC-4	B9	19.0000	4.0000	5.0000	6.0000	20.0000
43	NWC-4	B9	19.0000	5.0000	6.0000	7.0000	20.7000
44	NWC-4	B9	19.0000	6.0000	7.0000	8.0000	21.4000
45	NWC-4	B9	19.0000	7.0000	8.0000	9.0000	22.1000
46	DEEP-1	B12	18.5000	0.0000	1.0000	2.0000	16.0000
47	DEEP-1	B12	18.5000	1.0000	2.0000	3.0000	16.7000
48	DEEP-1	B12	18.5000	2.0000	3.0000	4.0000	17.4000
49	DEEP-1	B12	18.5000	3.0000	4.0000	5.0000	18.1000
50	DEEP-1	B12	18.5000	4.0000	5.0000	6.0000	18.8000
51	DEEP-1	B12	18.5000	5.0000	6.0000	7.0000	19.5000
52	DEEP-1	B12	18.5000	6.0000	7.0000	8.0000	20.2000
53	DEEP-1	B12	18.5000	7.0000	8.0000	9.0000	20.9000
54	DEEP-1	B12	18.5000	8.0000	9.0000	10.0000	21.6000
55	DEEP-1	B12	18.5000	9.0000	10.0000	11.0000	22.3000
56	DEEP-1	B12	18.5000	10.0000	11.0000	12.0000	23.0000

TABLE 8
AVS in Sapelo Island Sediments[#]

Variable Identification and Units for Data Table

STA\$	Sediment identifier
SEASON\$	Table number in the reference from which the data were taken
DEPTH1	Starting depth of the core slice (cm)
DEPTH2	Ending depth of the core slice (cm)
FES_W	Measured AVS in the slice (weight %)
FES	Measured AVS in the slice ($\mu\text{mol/g}$)

#[Reaves, 1984]

	STA\$	SEASON\$	DEPTH1	DEPTH2	FES_W	FES	
CASE	1	MUD	W	0.000	1.000	0.006	1.875
CASE	2	MUD	W	1.000	2.000	0.005	1.563
CASE	3	MUD	W	2.000	3.000	0.012	3.750
CASE	4	MUD	W	3.000	4.000	0.066	20.625
CASE	5	MUD	W	4.000	5.000	0.079	24.688
CASE	6	MUD	W	5.000	6.000	0.058	18.125
CASE	7	MUD	W	6.000	7.000	0.061	19.063
CASE	8	MUD	W	7.000	8.000	0.051	15.938
CASE	9	MUD	W	8.000	9.000	0.047	14.688
CASE	10	MUD	W	9.000	10.000	0.060	18.750
CASE	11	MUD	W	10.000	11.000	0.069	21.563
CASE	12	MUD	W	11.000	12.000	0.035	10.938
CASE	13	MUD	W	12.000	13.000	0.039	12.188
CASE	14	MUD	W	13.000	14.000	0.046	14.375
CASE	15	MUD	W	14.000	15.000	0.036	11.250
CASE	16	MUD	S	0.000	1.000	0.075	23.438
CASE	17	MUD	S	1.000	2.000	0.058	18.125
CASE	18	MUD	S	2.000	3.000	0.090	28.125
CASE	19	MUD	S	3.000	4.000	0.097	30.313
CASE	20	MUD	S	4.000	5.000	0.120	37.500
CASE	21	MUD	S	5.000	6.000	0.229	71.563
CASE	22	MUD	S	6.000	7.000	0.165	51.563
CASE	23	MUD	S	7.000	8.000	0.153	47.813
CASE	24	MUD	S	8.000	9.000	0.164	51.250
CASE	25	MUD	S	9.000	10.000	0.193	60.313
CASE	26	MUD	S	10.000	11.000	0.176	55.000
CASE	27	MUD	S	11.000	12.000	0.222	69.375
CASE	28	MUD	S	12.000	13.000	0.201	62.813
CASE	29	MUD	S	13.000	14.000	0.222	69.375
CASE	30	MUD	S	14.000	15.000	0.220	68.750
CASE	31	CK	W	0.000	1.000	0.031	9.688
CASE	32	CK	W	1.000	2.000	0.088	27.500
CASE	33	CK	W	2.000	3.000	0.077	24.063
CASE	34	CK	W	3.000	4.000	0.073	22.813
CASE	35	CK	W	4.000	5.000	0.103	32.188
CASE	36	CK	W	5.000	6.000	0.113	35.313
CASE	37	CK	W	6.000	7.000	0.087	27.188
CASE	38	CK	W	7.000	8.000	0.053	16.563
CASE	39	CK	W	8.000	9.000	0.116	36.250
CASE	40	CK	W	9.000	10.000	0.133	41.563
CASE	41	CK	W	10.000	11.000	0.124	38.750
CASE	42	CK	W	11.000	12.000	0.124	38.750
CASE	43	CK	W	12.000	13.000	0.070	21.875
CASE	44	CK	W	13.000	14.000	0.090	28.125
CASE	45	CK	W	14.000	15.000	0.099	30.938
CASE	46	CK	S	0.000	1.000	0.019	5.938
CASE	47	CK	S	1.000	2.000	0.018	5.625
CASE	48	CK	S	2.000	3.000	0.039	12.188
CASE	49	CK	S	3.000	4.000	0.075	23.438
CASE	50	CK	S	4.000	5.000	0.115	35.938
CASE	51	CK	S	5.000	6.000	0.196	61.250
CASE	52	CK	S	6.000	7.000	0.155	48.438
CASE	53	CK	S	7.000	8.000	0.144	45.000
CASE	54	CK	S	8.000	9.000	0.135	42.188
CASE	55	CK	S	9.000	10.000	0.125	39.063
CASE	56	CK	S	10.000	11.000	0.103	32.188
CASE	57	CK	S	11.000	12.000	0.102	31.875
CASE	58	CK	S	12.000	13.000	0.088	27.500
CASE	59	CK	S	13.000	14.000	0.102	31.875
CASE	60	CK	S	14.000	15.000	0.103	32.188

DEVELOPMENT OF EXPERIMENTAL METHODOLOGY

A. Measuring Cadmium Activity

The most direct method to measure activity of a metal is to use an ion specific electrode (Orion Model 94-48). Ion selective electrodes measure the activity of the target species only - in this case Cd²⁺). The determination of a reliable standard curve is the first step in the establishment of an analytical method. For the ion selective electrode this involves the measurement of relative potential in conjunction with a double function reference electrode as a function of activity, of the analyte (Cd²⁺).

Because Cd²⁺ forms several stable complexes with Cl⁻, the standardization cannot be done in seawater. A noncomplexing matrix of the same ionic strength as seawater, 0.7 M NaNO₃, was used. The results are shown in Fig. 1. The linear behavior of the potential versus cadmium activity to 0.1 mg/L with a slope of 28.2 is consistent with the Nernst equation. When a hydrogen carbonate buffer is added to mimic seawater conditions no significant change occurs in the standard curve (Fig. 2). If the 0.7 M NaNO₃ Cd²⁺ potentials are compared with those obtained for the same total cadmium in seawater, the Cd²⁺ fraction is about 5%, which is consistent with values calculated from simultaneous equilibrium models of cadmium speciation in seawater.

To further investigate the performance of the cadmium electrode, several titrations were performed using ions that are known to complex with Cd²⁺ and for which the formation constants are known. The results of a hydroxide titration are shown in Fig. 3. The value of pK₁ = 9.75 is consistent with literature values. The results of a chloride titration are shown in Fig. 4. The initial total cadmium concentrations are 10 and 1 mg/L. The value of 26.4 which is obtained for the formation constant of CdCl⁺ is consistent with reported values.

B. Water Only Exposure - Bioassay Results

The above experiments demonstrated that the cadmium electrode was indeed measuring the cadmium activity in the systems of interest. During this period water-only bioassays were being performed at the Narragansett EPA Environmental Research Lab (ERL) to determine the total

cadmium (Cd_T) LC₅₀ for the organism, Ampelisca abdita, that was to be used in the sediment exposure studies. The results are shown in Fig. 5 for both the definitive and range finding experiments.

Following this preliminary work, total cadmium samples bracketing the LC₅₀ values of 0.32 - 0.55 mg/L were prepared in seawater. The Cd^{2+} concentration was determined by the electrode, while total cadmium was measured by anodic stripping voltammetry and graphite furnace atomic absorption spectrometry using an ammonium phosphate matrix modification procedure. The results are shown in Fig. 6. The data from the two methods are combined in Fig. 7 and the ratio of cadmium activity to total cadmium concentration is found to be $\text{Cd}^{2+}/\text{Cd}_T = 0.051$. This result is used to convert the total cadmium concentrations in the water only exposure experiment to the cadmium activity.

C. Interstitial Water Diffusion Sampler

The final task that had to be completed before sediment Bioassay studies could be undertaken was the design and construction of a suitable sampling device. The final design of the peeper is shown in Fig. 8. The device is constructed of acrylic material. The body is 1.5 in. thick with three 0.5 in. holes bored at six different levels 0.75 in. apart. A solid 0.25 in. base plate is fused to the back. The front contains a nucleopore membrane, on top of which is a 20 mil polyethylene gasket and a 0.5 in. cover plate. The entire assembly is held together with six 0.25 in. PVC nuts and bolts. The volume of each cell is 5.0 mL, which provides the necessary minimum sample volume of 15 mL required for the electrode measurement, at each sampling level. This sample volume size was determined by experiment to be adequate for reliable measurement.

The interstitial water sampling device depends upon diffusion across the membrane to measure the interstitial water concentrations. In order to establish the equilibration time for transfer across the membrane the cells of the assembled peeper were filled with distilled water. The device was then immersed in seawater. The cells were sampled periodically and the conductivity of the sample was measured. The results are shown in Fig. 9. As can be seen the 12 micron membrane

equilibrated quite rapidly. However even the 1 micron membrane reached equilibrium within 24 hours. Since this equilibration time is well within the time scale of the toxicity experiments, it was decided to use this porosity membrane in subsequent experiments.

D. Initial Toxicity Experiments

When sediment exposure experiments are performed it is necessary to be able to predict the interstitial cadmium concentration that will be obtained from initially combining a quantity of a cadmium salt with measured amount of reference sediment and seawater. To provide this information solutions were prepared with total cadmium concentrations of 200 to 6000 ppm. Each of these contained 1000 mL of control sediment (55% water). The peepers were then placed in the settled sediment-water systems. After two days the cadmium concentration was measured as a function of depth. The results are shown in Fig. 10. Initially it was believed that the interstitial cadmium concentration was being controlled by simple partitioning between a sediment bound component and the aqueous component. The amount of cadmium used in preparing the systems for the interstitial water Bioassay experiments was determined from these results. The attempt was made to bracket the cadmium LC₅₀ obtained in the water only exposure. However, in all of the systems the final interstitial cadmium concentration was not sufficient to produce mortality.

The first bioassay results indicated that an additional process was operating in the cadmium-sediment water system. The observation of a yellow precipitate (probably CdS) when preparing the more concentrated systems suggested that sediment generated sulfide was depressing the aqueous cadmium concentration during the actual exposure.

E. Development of Acid Volatile Sulfide Extraction Method

The most labile sulfide component of sediments is the acid volatile sulfide (AVS). It is the solid phase sulfide in the sediment that is soluble in cold acid. The measurement technique is to convert the sulfides to H₂S(aq), purge it with a gas, and trap it [see Morse et al., 1987 for a review]. A 500 mL Erlenmeyer flask reaction vessel fitted with a three-hole stopper is followed by three sequentially connected 250 mL Erlenmeyer flask trapping vessels. The first is a chloride trap

with 200 mL of pH 4 buffer (0.05M potassium hydrogen phthalate) to prevent chloride carry over. The second and third traps contain 200 mL of a 0.1M silver nitrate solution for trapping H₂S. The four flasks are connected with airtight appropriately shaped glass and Tygon tubing.

A nitrogen gas flow allows continuous purging of the system. In order to prevent oxidation the gas flows through an oxygen-scrubbing system consisting of a vanadous chloride solution in the first scrubbing tower and the matrix of the analyte (seawater) in the second tower. Vanadous chloride is prepared using four grams of ammonium metavanadate boiled with 50 mL of concentrated hydrochloric acid and diluted to 500 mL. Amalgamated zinc, prepared by taking about 15 grams of zinc, covering it with deionized water and adding 3 drops of concentrated hydrochloric acid before adding a small amount of mercury to complete the amalgamation, is then added to the vanadous chloride solution.

The sediment sample (10-15 grams of wet sediment) or standard to be analyzed is placed in the reaction vessel after the entire system has been purged with nitrogen for about an hour. The system is again purged for 5-10 minutes, and deaerated 6M hydrochloric acid is added from a thistle tube to achieve a final concentration in the vessel of 0.5M. The system is run at room temperature for one hour which has been found to be sufficient to complete the extraction. Fig. 11 present the results of an experiment in which the time course of AVS extraction from a sediment (Long Island Sediment) is followed. It is clear that one hour is sufficiently long for the extraction to be completed.

F. Figure Captions

Figure. 1 Cadmium electrode calibration curve in 0.7M NaNO₃, pH = 8 corresponding to the ionic strength of seawater. Slope = 28.2. Sensitivity of the electrode is 0.1 mg/L

Figure. 2 Cadmium electrode calibration curve in 0.7M NaNO₃, 0.002M NaHCO₃, pH = 8, corresponding to the ionic strength and bicarbonate concentration of seawater. Slope = 28.2. Sensitivity of the electrode is 0.1 mg/L.

Figure 3. Hydroxide titration to determine pK₁ for the reaction: Cd²⁺ + OH⁻ <-> CdOH⁺.

Figure 4. Chloride titrations to determine K₁ for the reaction: Cd²⁺ + Cl⁻ <-> CdCl⁺.

Figure 5. Ampelisca Toxicity Test: Water Only Exposure. LC50 = 0.32 mg Cd/L (top) and 0.55 mg Cd/L

Figure 6. Cadmium concentrations in the toxicity test samples (mg Cd/L) determined using a polarographic method (top) and using an AA graphite furnace method (bottom) versus electrode concentration (mg Cd²⁺/L).

Figure 7. Cadmium concentrations in the toxicity test samples (mg Cd/L) determined using both a polarographic and AA graphite furnace method versus electrode concentration (mg Cd²⁺/L). Regression line is Cd²⁺ = 0.051 Cd, which implies a K₁ of the chloride reaction of K₁ = 26.4 M₋₁.

Figure 8. Design of diffusion sampler "peeper"

Figure 9. Time to equilibrium for membrane transfer.

Figure 10. Initial interstitial water sampling results. Static exposure.

Figure 11. Time course experiment for AVS extraction of Long Island Sound sediment.

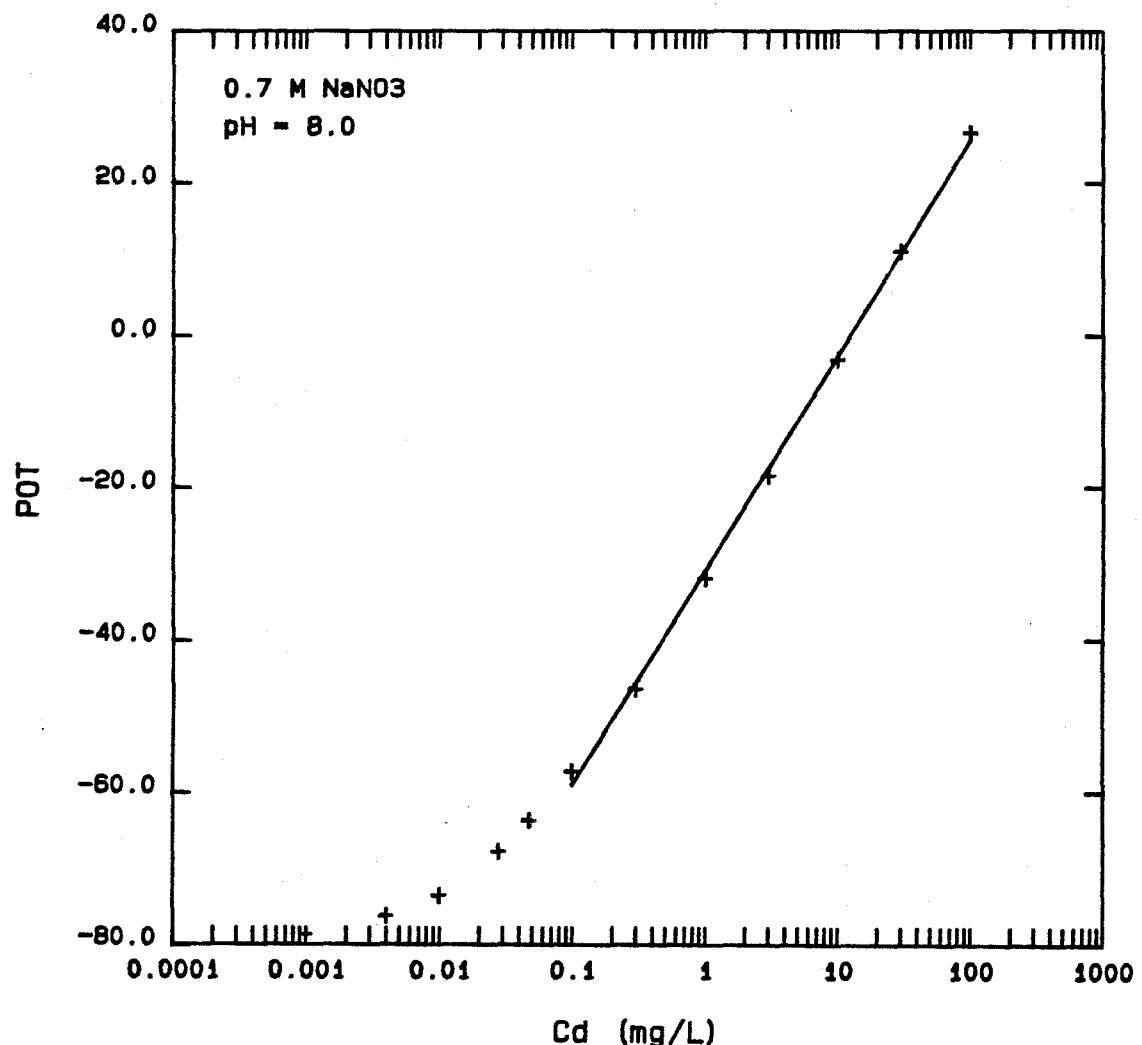


Figure. 1 Cadmium electrode calibration curve in 0.7M NaNO₃, pH = 8 corresponding to the ionic strength of seawater. Slope = 28.2. Sensitivity of the electrode is 0.1 mg/L

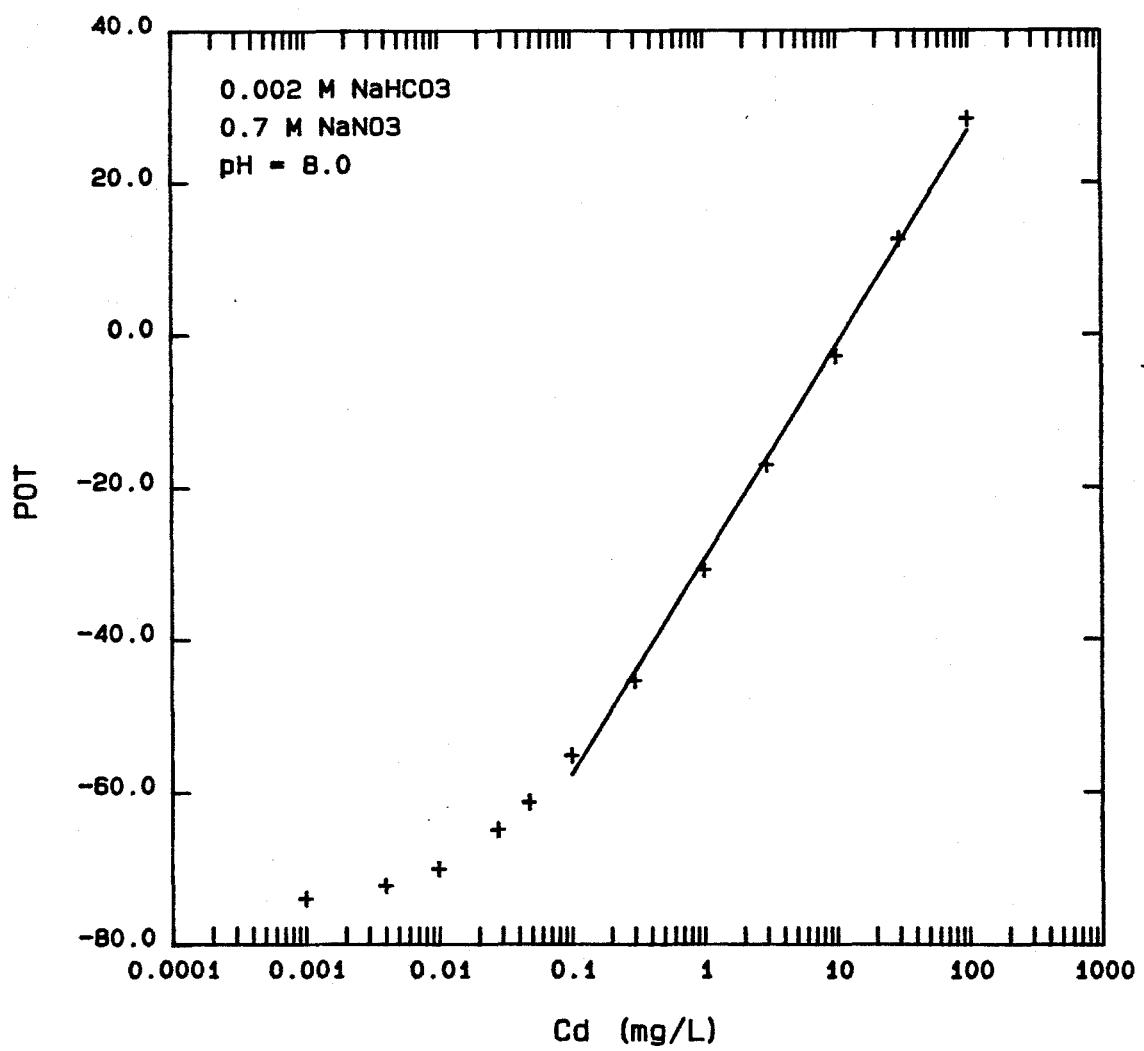


Figure. 2 Cadmium electrode calibration curve in 0.7M NaNO₃, 0.002M NaHCO₃, pH = 8, corresponding to the ionic strength and bicarbonate concentration of seawater. Slope = 28.2. Sensitivity of the electrode is 0.1 mg/L.

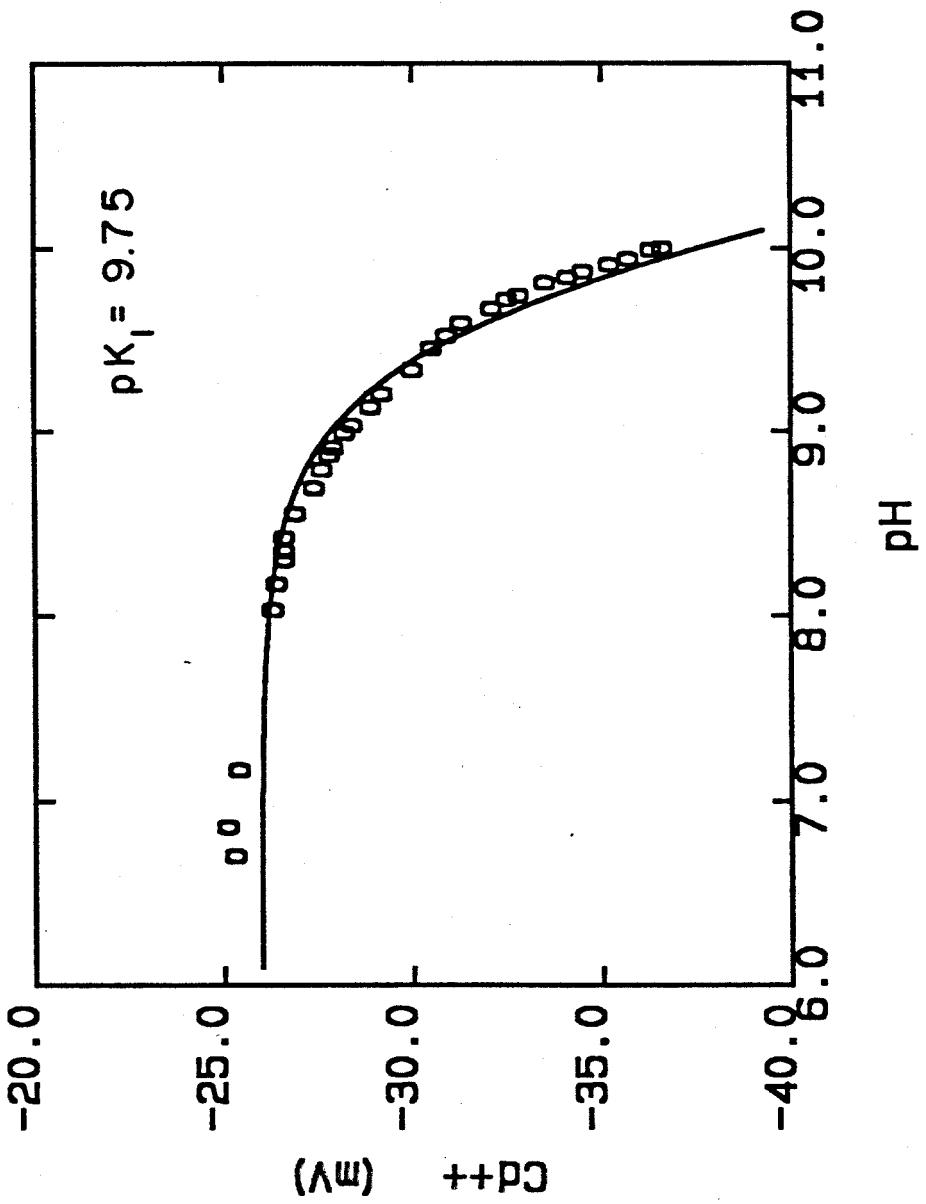


Figure 3. Hydroxide titration to determine $\text{p}K_1$ for the reaction: $\text{Cd}^{2+} + \text{OH}^- \rightleftharpoons \text{CdOH}^+$.

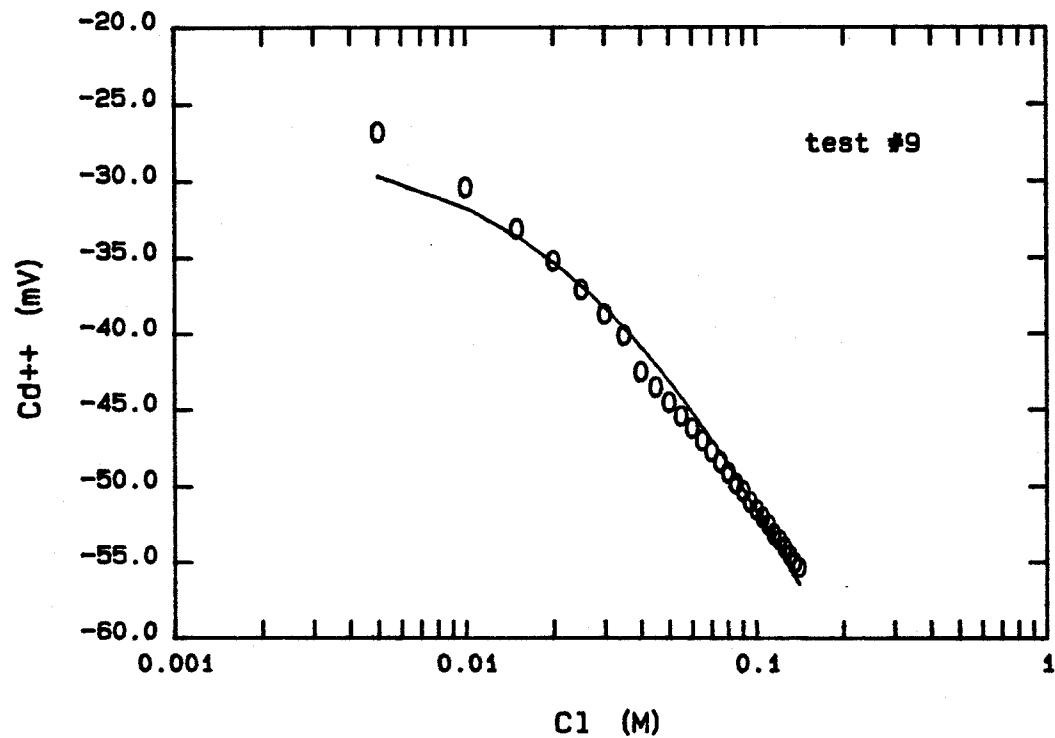
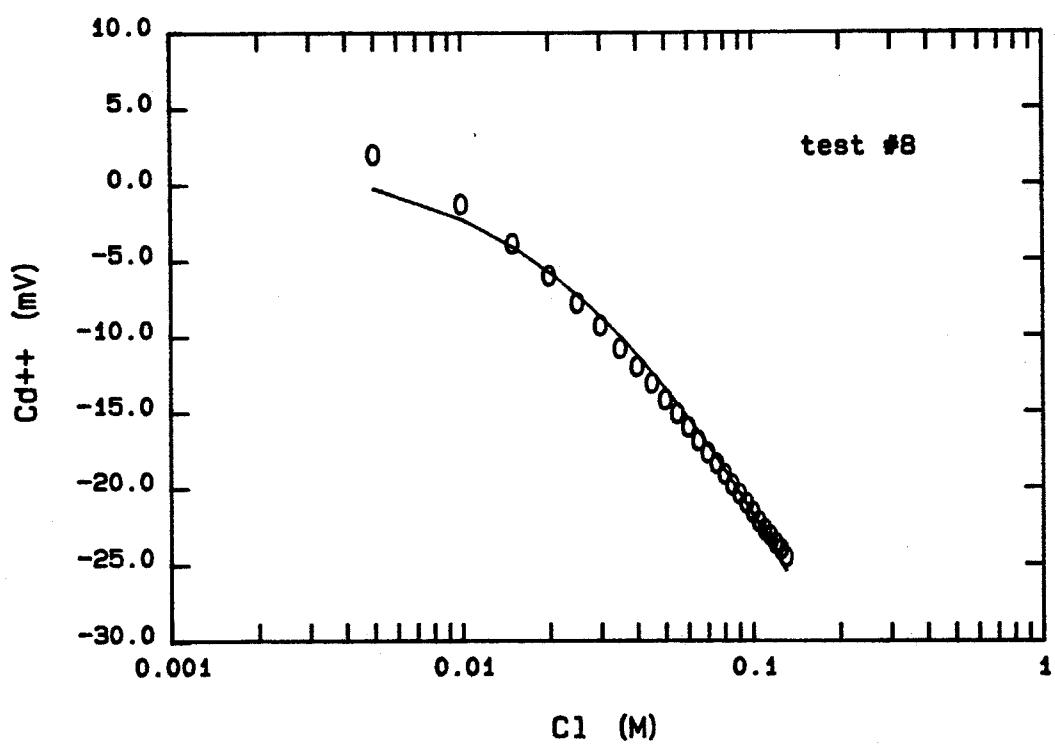


Figure 4. Chloride titrations to determine K_1 for the reaction: $\text{Cd}^{2+} + \text{Cl}^- \leftrightarrow \text{CdCl}^+$.

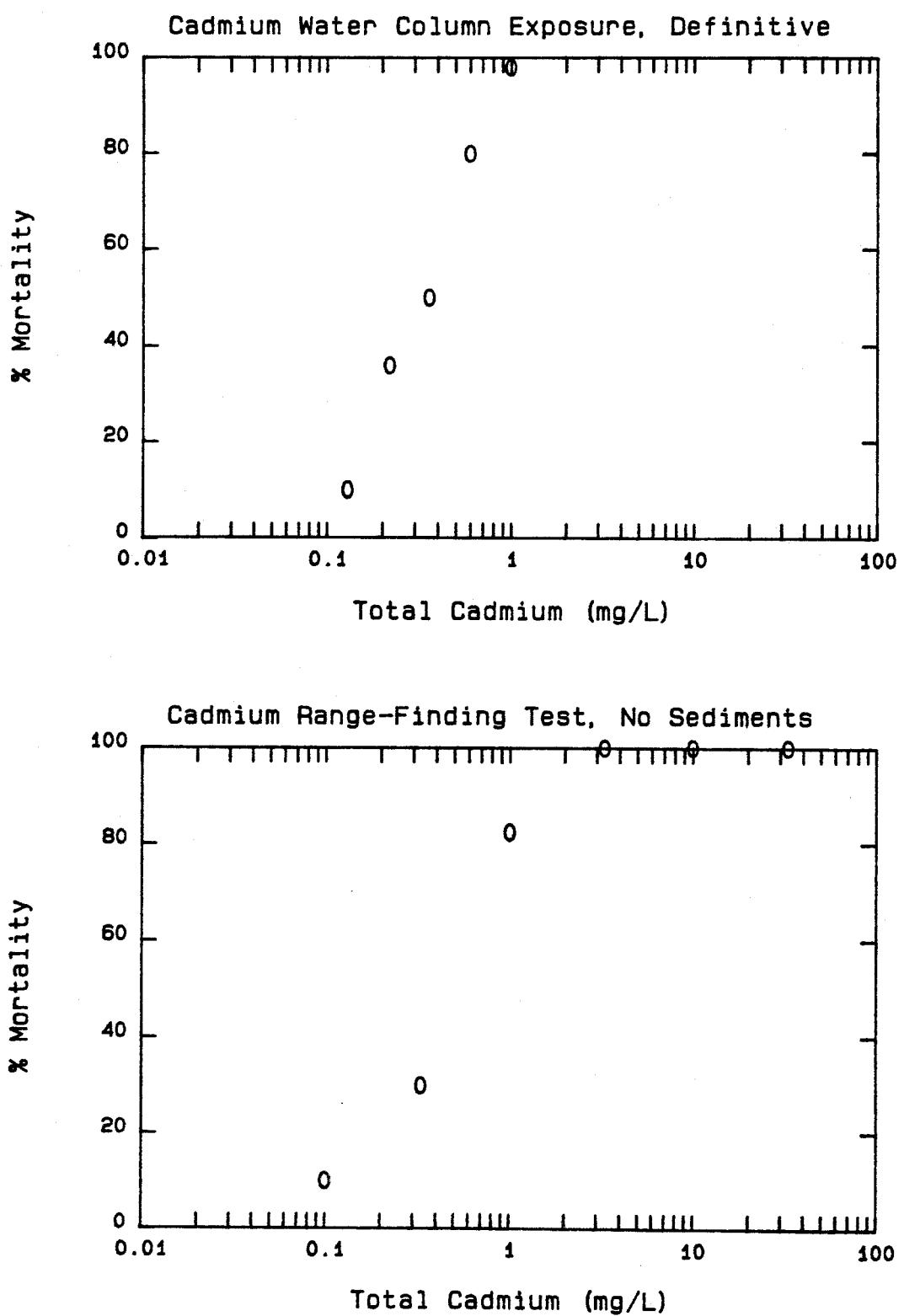


Figure 5. Ampelisca Toxicity Test: Water Only Exposure. LC₅₀ = 0.32 mg Cd/L (top) and 0.55 mg Cd/L

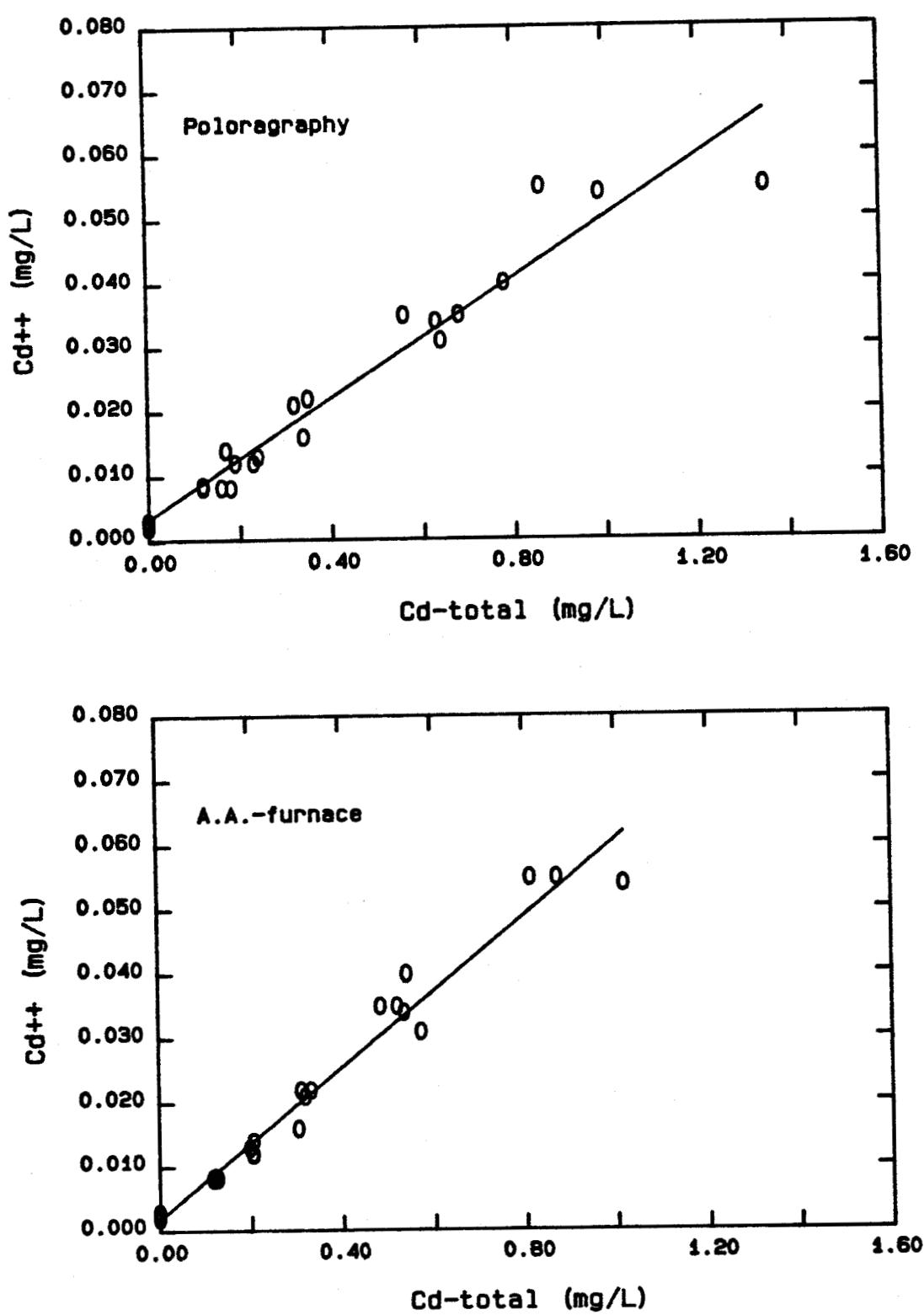


Figure 6. Cadmium concentrations in the toxicity test samples (mg Cd/L) determined using a polarographic method (top) and using an AA graphite furnace method (bottom) versus electrode concentration (mg Cd²⁺/L).

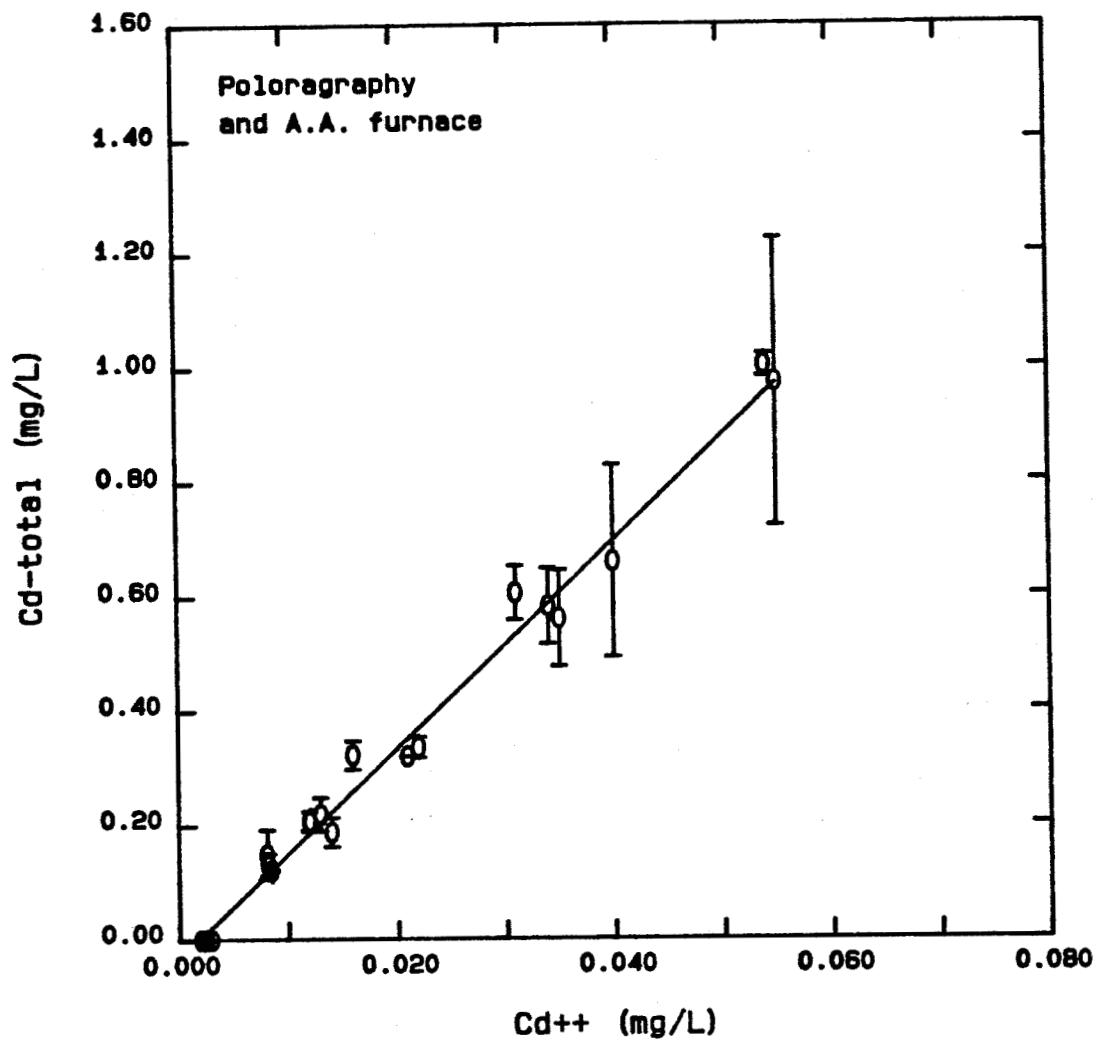


Figure 7. Cadmium concentrations in the toxicity test samples (mg Cd/L) determined using both a polarographic and AA graphite furnace method versus electrode concentration (mg Cd²⁺/L). Regression line is Cd²⁺ = 0.051 Cd, which implies a K₁ of the chloride reaction of K₁ = 26.4 M⁻¹.

PEEPER DESIGN DIMENSIONS (CM)

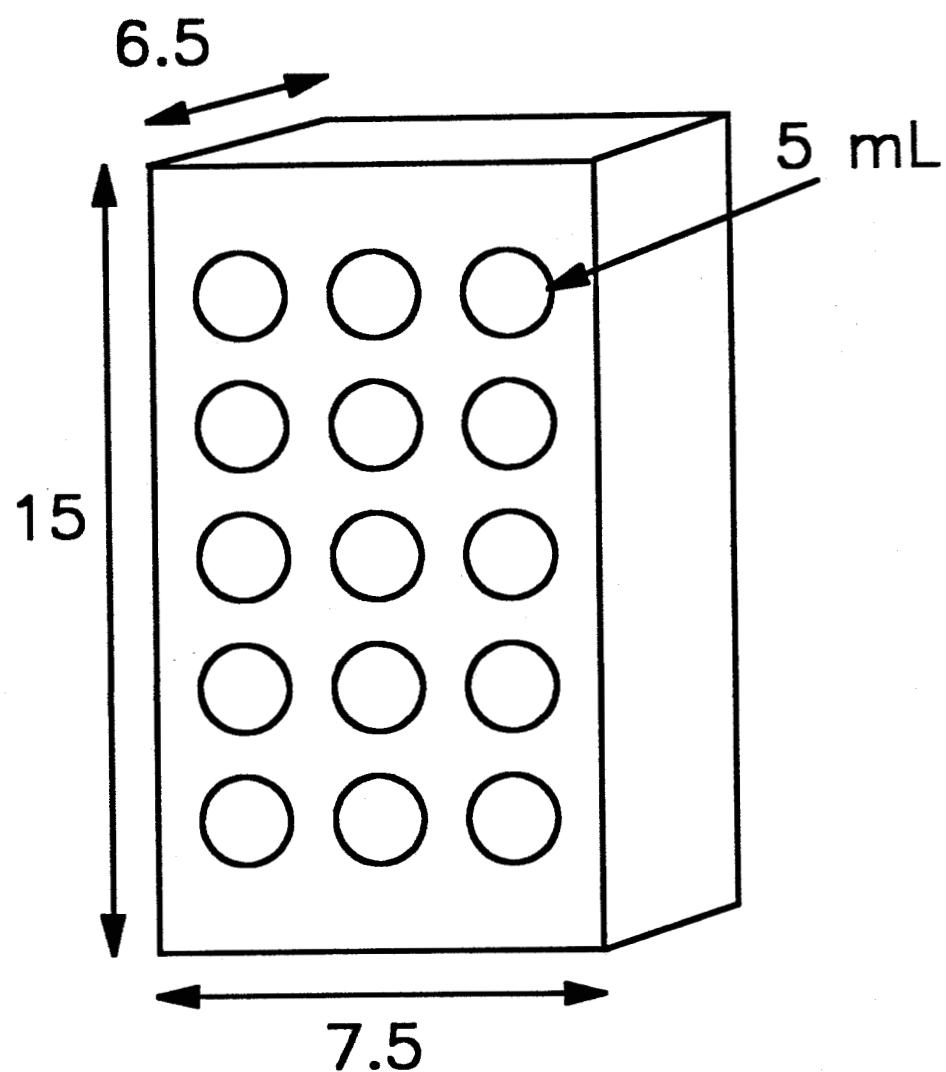


Figure 8. Design of diffusion sampler "peeper"

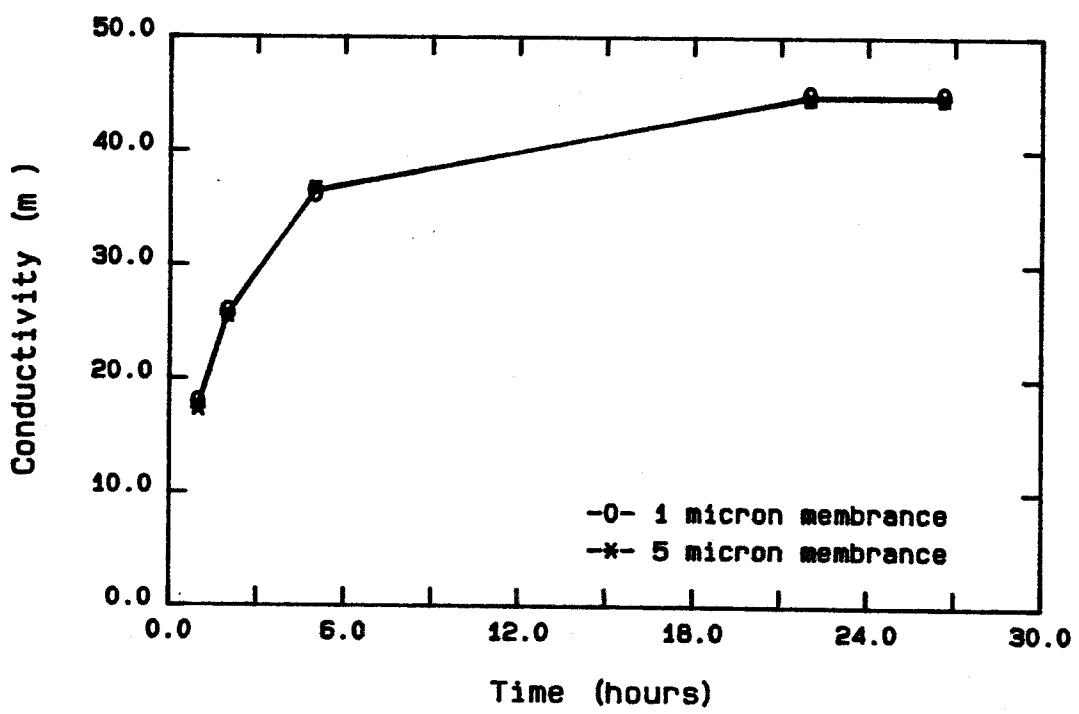
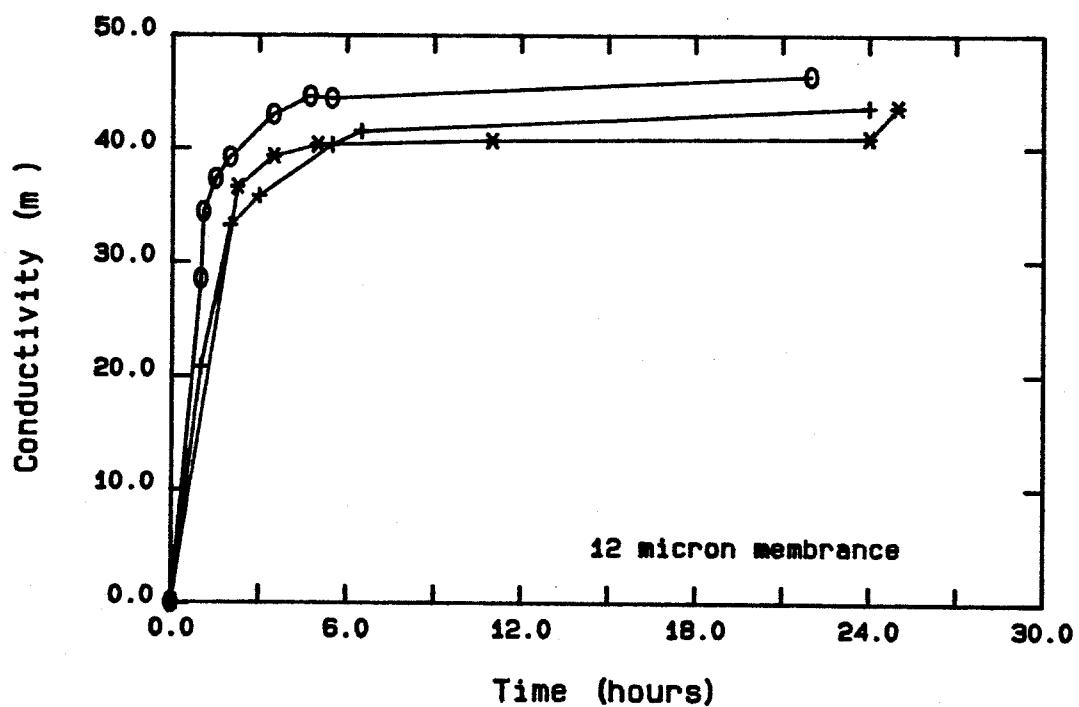


Figure 9. Time to equilibrium for membrane transfer.

SEDIMENT PORE WATER CADMIUM ACTIVITY

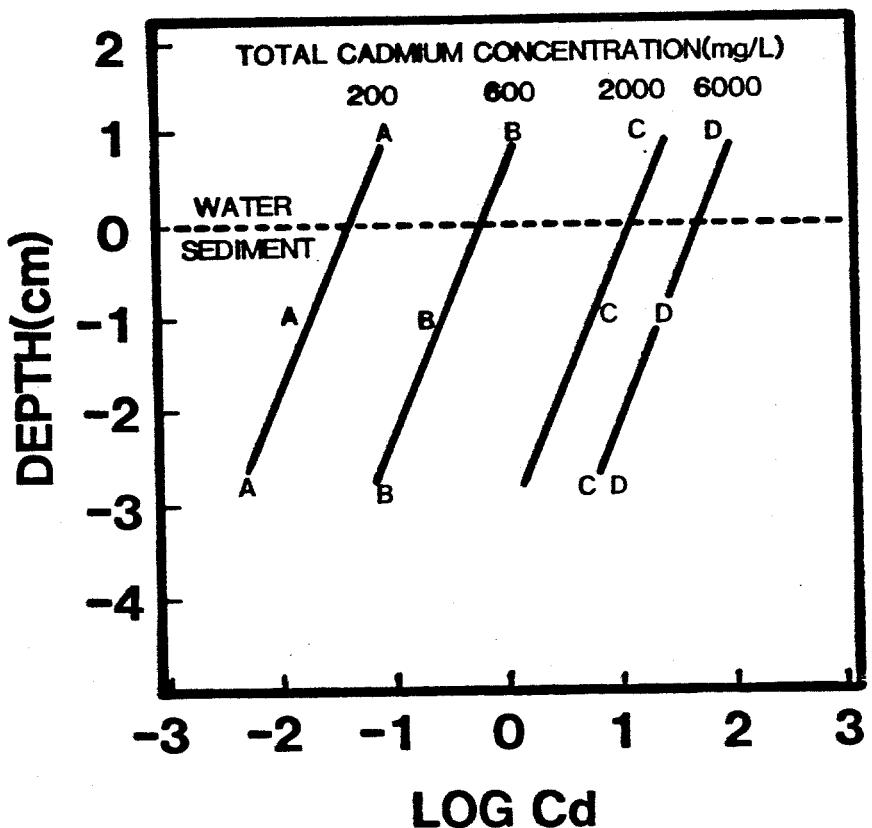


Figure 10. Initial interstitial water sampling results. Static exposure.

ACID VOLATILE SULFIDE
YIELD vs EXTRACTION TIME

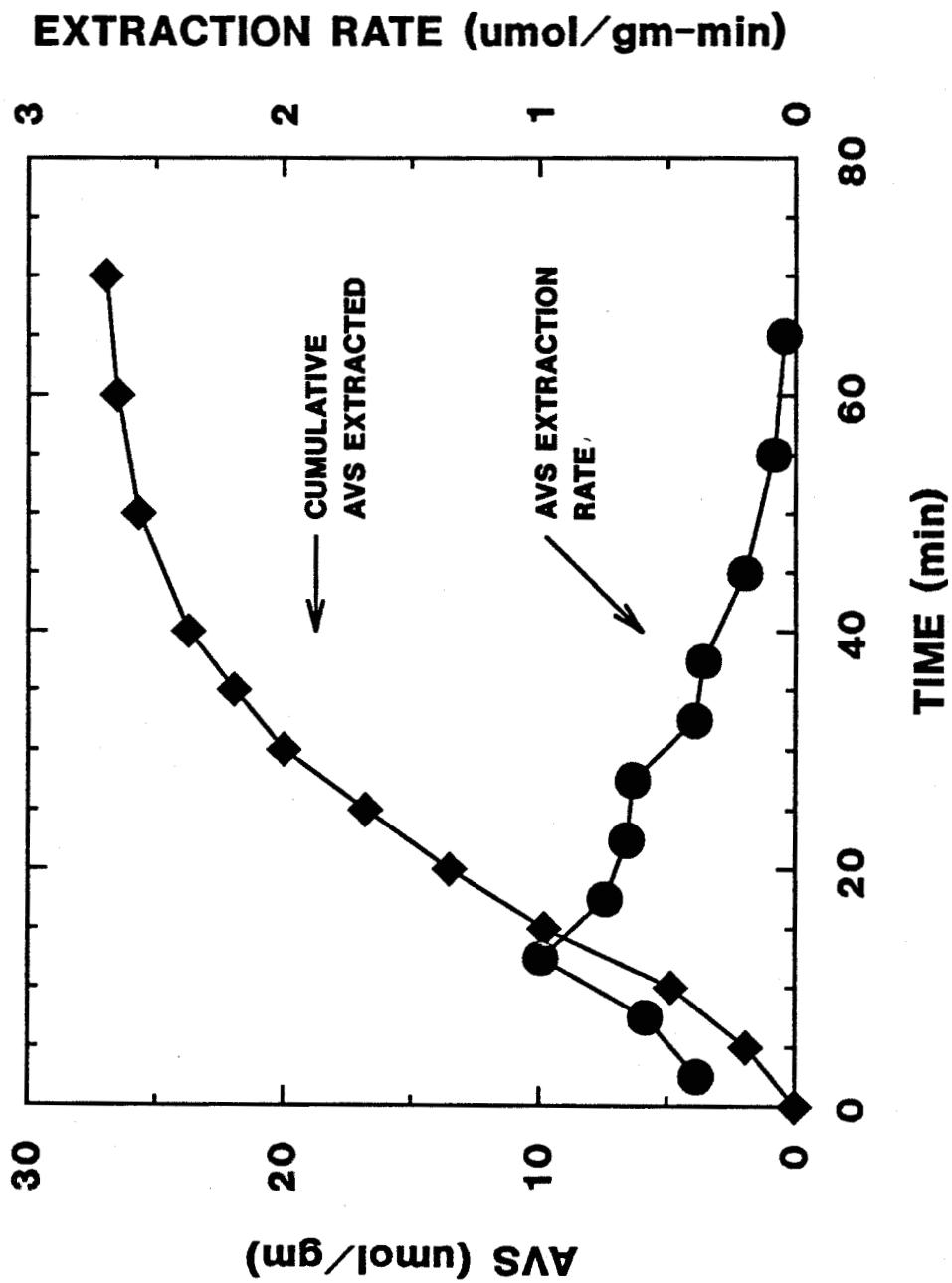


Figure 11. Time course experiment for AVS extraction of Long Island Sound sediment.

**DEVELOPMENT AND VERIFICATION OF A KINETIC
MODEL OF POLLUTANT DESORPTION
FROM DREDGE SEDIMENT AND SEWAGE SLUDGE**

I. Introduction

The purpose of this project is to develop a kinetic model of heavy metal desorption from sludges and sediments. This reaction significantly influences the fate of these chemicals during the ocean disposal of sludge and dredged sediments since the rapidity with which metals desorb determines, to a large extent, their ultimate fate. If desorption is rapid then the chemical is primarily in the aqueous phase and water column transport determines its fate. However if the metal remains in particulate form then settling and sedimentation will occur. Thus the rate and degree of desorption greatly influence the fate of these chemicals.

Significant progress has been made in the development of the models for this reaction. An equilibrium desorption model for reversibly bound heavy metals and organic chemicals was developed in a previous project (Di Toro and Mahony, 1986). The model describes the initial desorption of heavy metals from sewage sludge and Black Rock Harbor sediment upon mixing with seawater at various particle concentrations. Only a small fraction of the total particulate metal initially desorbs. Consecutive desorptions confirmed that only a small fraction was available during short term (1 hour) desorptions. However, it was noted that although desorption of the reversibly bound (or labile) metal fraction was rapid there was a distinct and slower reaction which also released particle bound metal into the aqueous phase. The time scales of this reaction is days rather than hours. This slow release phenomena, which had been observed experimentally by Rohatgi and Chen (1975) for digested sewage sludge, was confirmed by us for Black Rock Harbor sediment. Rohatgi and Chen speculated that the release might be due to an oxidation reaction which liberated heavy metals that either were bound to organic carbon or which were present as metal sulfides.

These observations, and our own experimental experience, lead us to propose a three component model of heavy metal desorption. Particulate metal was assumed to exist as either reversibly sorbed metal, for which the reversible partition coefficient follows the particle interaction model which we had developed in the previous project, and two other com-

ponents. A resistant component which initially is not desorbed but is gradually released over a period of five to ten days; and a refractory fraction which resists release for the duration of the experiment. This model was fit to Rohatgi and Chen's data and it provides a reasonable fit to the observations. An example is shown in fig. 1.

However, Rohatgi and Chen's experiments could not be used to distinguish between a number of other possible models that could easily fit the data as well. For example it is possible that the increase in dissolved concentration of metals during the experiment is due to slow, reversible, desorption kinetics. That is, it is possible that the desorption reaction for these particles takes a number of days before it reaches equilibrium so that the slow release is just due to the slowness of the desorption reaction. Desorption kinetics which are initially rapid but are followed by a slow phase have been observed for hydrophobic organic chemicals (Karickhoff, 1980).

Alternately it may be that the partition coefficient is decreasing during the experiment due to a change in particle properties. If the particle surface properties are being altered by exposure to oxygenated seawater during the experiment then the desorption reaction might still be rapid but it is adjusting to the slowly changing surface properties of the particles. The oxidation of particulate reactive organic carbon would influence the partitioning in this way.

The purpose of this research project is to experimentally distinguish between these possibilities and to determine, to the extent possible, the mechanism responsible for the slow release of metals and to incorporate any necessary changes into the desorption model. The experiments conducted to date, using Black Rock Harbor sediment and two digested sewage sludges from the Bergen County and Ridgewood sewage treatment plants, appear to confirm the three component model as originally formulated. It appears that the slow release of metals are from the fraction of the metal which is initially present as particulate metal sulfides. These are released as the particulate sulfides are oxidized to sulfate in the reactor. An initial experiment in sulfate free seawater exhibited an increase in sulfate over time confirming the oxidation of sulfide to sulfate. Additionally, an anaerobic reactor, for

which nitrogen gas bubbling was substituted for oxygen containing air bubbling, exhibited no increase in dissolved metal over time. Parallel experiments with radiotagged metal appear to confirm this interpretation although some short term (less than one day) effects are still to be explained.

Hence the data appears to confirm the three component desorption kinetic model. It remains to actually fit these data to the model equations in order to obtain the model parameters. The final version would then be applicable to the evaluation of the fate of heavy metals during the ocean disposal of sludges and dredged sediments.

II. Experimental Design and Results to Date

The experiments are all conducted in small (350 mL) reaction vessels. Metal free seawater and sludge or sediment particles are initially added. The reactors are kept aerobic by bubbling filtered air and are mixed using magnetic stirring. Duplicate reactors at two particle concentrations, 300 and 3000 mg/L, are used. A particle free control reactor which receives the same air, stirring, and sampling, is monitored to detect possible contamination. At various times during the experiment samples are taken and analyzed for total and dissolved metal (the separation is by centrifugation). The duration of the experiment is twenty to thirty days.

Fig. 2 presents the results obtained for copper from Black Rock Harbor sediment and Bergen County sewage sludge for two particle concentrations (300 and 3000 mg/l). The log scales for the concentration are increased by a factor of ten for the 3000 mg/L reactor so that the results for both reactors can be directly compared. The increase in dissolved copper concentration is approximately one order of magnitude for both particle types with the larger dissolved concentrations associated with the smaller particle concentration. This is a result of the decreased fraction of the metal that reversibly partitions to the particles at lower particle concentrations. Fig. 3 presents the results for zinc and cadmium for Black Rock Harbor sediment. Again the release increases by approximately an order of magnitude. However, the chromium

results in fig. 4 show no significant increase for either Black Rock Harbor sediment or Bergen County sludge. The significance of this fact is discussed below.

Changes in pH and total organic carbon (TOC) are shown in fig. 5. For Black Rock Harbor sediment the pH and TOC are essentially constant. For Bergen County sludge the pH increases slightly during the first day and stayed approximately constant thereafter. Although this may have affected the reversible partition coefficient during this time (it would increase with increasing pH) the release continues during the period of constant pH. The TOC decreased from 400 (40) mgC/L to 200 (20) mgC/L in the 3000 (300) mg/L reactor during the experiment. However the release patterns of metals is similar for Black Rock Harbor sediment and Bergen County sludge. Since no change in TOC or pH was observed for Black Rock Harbor sediment, it is unlikely that the pH and TOC changes were responsible for the metal release.

The results of an anaerobic reactor experiment are shown in fig. 6. The arrangement is the same as the aerobic reactor, the only difference is that nitrogen gas is substituted for the air that is bubbled through the reactors. As can be seen the dissolved copper concentration stayed constant for Black Rock Harbor sediment and the two sludges. The TOC also remained constant as expected since the experiment did not last long enough to initiate significant anaerobic breakdown of the sludges. Thus the presence of oxygen is clearly necessary for the slow metal release to occur.

The fact that copper, cadmium, and zinc are slowly released while chromium concentrations are constant is consistent with the hypothesis that the slow release of metals is due to the oxidation of particulate metal sulfides. The solubility of these metal-sulfides is very low as indicated below (Lindsay, 1979):

Solubility of MeS	Log Ksp
CuS (covellite)	-36.10
CdS (greenokite)	-27.07
ZnS (wurtzite)	-22.5

However no Cr(III) sulfide solid phase is reported in tabulations of sulfide mineral solubilities (Naumov et al., 1974).

A possible approach to a direct verification that particulate sulfide is being oxidized is to monitor the oxidation end product, sulfate, and observe its concentration in time. An increase in sulfate would indicate that reduced sulfur, presumably sulfide, was being oxidized in the reactor. The experiment is conducted in sulfate-free artificial seawater so that the large naturally occurring sulfate background does not obscure the increase. The results are shown in fig. 7. The upper panel presents the actual observations. The initial sulfate concentration is due to the sulfate in the supernatant of the Black Rock Harbor sediment stock which was added at the start of the experiment. Future experiments will separate the particles from the supernatant. The bottom panel presents the excess sulfate, defined as the difference between that initially present and that observed at the indicated time. It is interesting to note that the pattern of sulfate release is similar to that observed for the metal release.

In addition to measurements of the total and dissolved metal concentrations over time additional parallel experiments were conducted with radiotagged metals. The purpose of these experiments was to examine the question of the time scale of adsorption and the possible variation of the partition coefficient over time. The kinetic experiment used the same reactor setup. After the particles were added the reactor was tagged with radioactive metal. The total and dissolved radiotagged metal was monitored in time. Changes in the distribution between total and dissolved metal could be due to either slow adsorption kinetics or changes in the adsorption partition coefficient. But the absence of change would eliminate both mechanisms as significantly contributing to the release phenomena.

The results from these kinetic experiments are shown in fig. 8 for zinc. The Bergen County sludge results indicate that very little change occurred in the dissolved concentration suggesting that the variation of TOC did not significantly change the partitioning. However the Black Rock Harbor sediment results indicated a substantial increase in dissolved concentration and a decrease in total concentration that is

attributed to the accumulation of sediment particles on the walls of the reactor. However the changing total concentration casts some doubt on the validity of the experiment so we plan to repeat it.

The results for cadmium are shown in fig. 9. With the exception of the data for less than one day, the dissolved cadmium concentration remained constant for the duration of the experiment for both Black Rock Harbor sediment and Bergen County sludge.

These experiments do not yield a completely consistent interpretation. However they do eliminate one possible model - that the adsorption and desorption kinetics are slow and reversible. If sorption is slow but ultimately reversible then for an adsorption experiment the dissolved concentration should initially be large, since adsorption has not yet had an opportunity to occur, and it should decrease with time. For the experiments where changes were observed the reverse pattern was observed so that slow reversible sorption kinetics appear to be ruled out.

In order to discriminate between adsorption kinetics and partition coefficient changes, an additional set of parallel experiments were performed in order to examine if the partition coefficient is varying in time. A parallel reactor was used which was not initially tagged with radioactive metal. Rather, at various times during the experiment a small sample was taken. This sample was then spiked with a small amount of radioactive metal. The sample was agitated for one hour after which the total and dissolved radioactive metal concentration was determined. Following the adsorption step, a desorption step was performed in order to examine the behavior of the reversibly sorbed metal. If the distribution of total and dissolved metal was changing with time, that would indicate that the partition coefficients were changing with time.

The results from these spiking experiments are shown in fig. 10 for zinc. With the exception of the changing adsorption partition coefficient for Black Rock Harbor sediment, the other partition coefficients are constant. It is interesting that the desorption partition coefficient is constant for both Black Rock Harbor sediment and Bergen County sludge.

The results of the spiking experiments for cadmium are shown in fig. 11. Once again the partition coefficients appear to be quite constant for times greater than one day. But both the adsorption and desorption partition coefficients are changing initially.

The results of the kinetic and spiking experiments are not consistent with a single explanation. On the one hand, the Bergen County sludge zinc kinetic and the spike adsorption-desorption data, the Black Rock Harbor sediment spike desorption data, and both the kinetic and spike adsorption-desorption Black Rock Harbor sediment and Bergen County sludge cadmium data for time greater than one day suggest that no partition coefficient changes are occurring since the dissolved concentrations are essentially constant. However, the Black Rock Harbor sediment zinc kinetic and spike adsorption data, and all the less than one day cadmium data, suggest that something was changing during these time periods. Additional experiments are planned to investigate the cause of these unexpected and as yet unexplained results.

III. Future Directions

The major focus of this portion of the research will be to complete the development of the kinetic model for metal desorption from sediments and sludges. The reactor data will be fit to the model equations to estimate the parameters. Additional radiotag experiments will be performed in an attempt to understand the somewhat anomalous results described above. However the focus of the work will be to complete the development with a minimum of experimental effort so that the Sediment Criteria work can begin. A project report will be written that includes all the experimental data as well as the modeling results. A journal article will also be prepared.

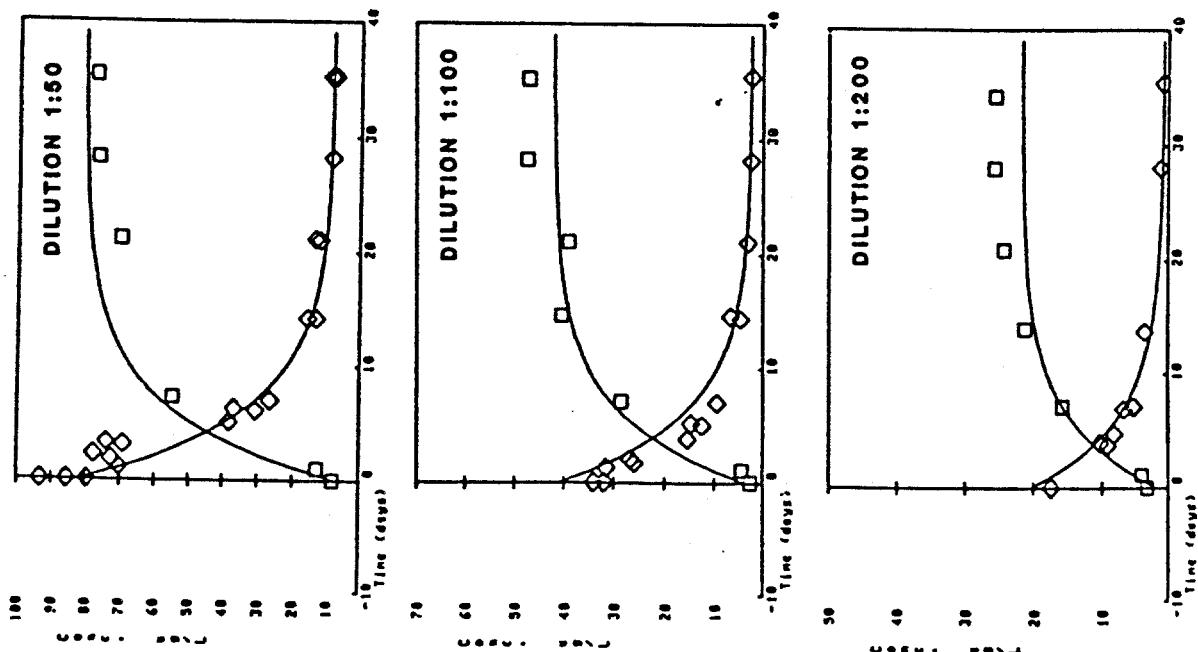
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- [5] Rohatgi, N. and Chen, K.Y. (1975): Transport of trace metals by suspended particulates on mixing with seawater. J. Water Pollut. Control Fed. 47: pp. 2298-2316.

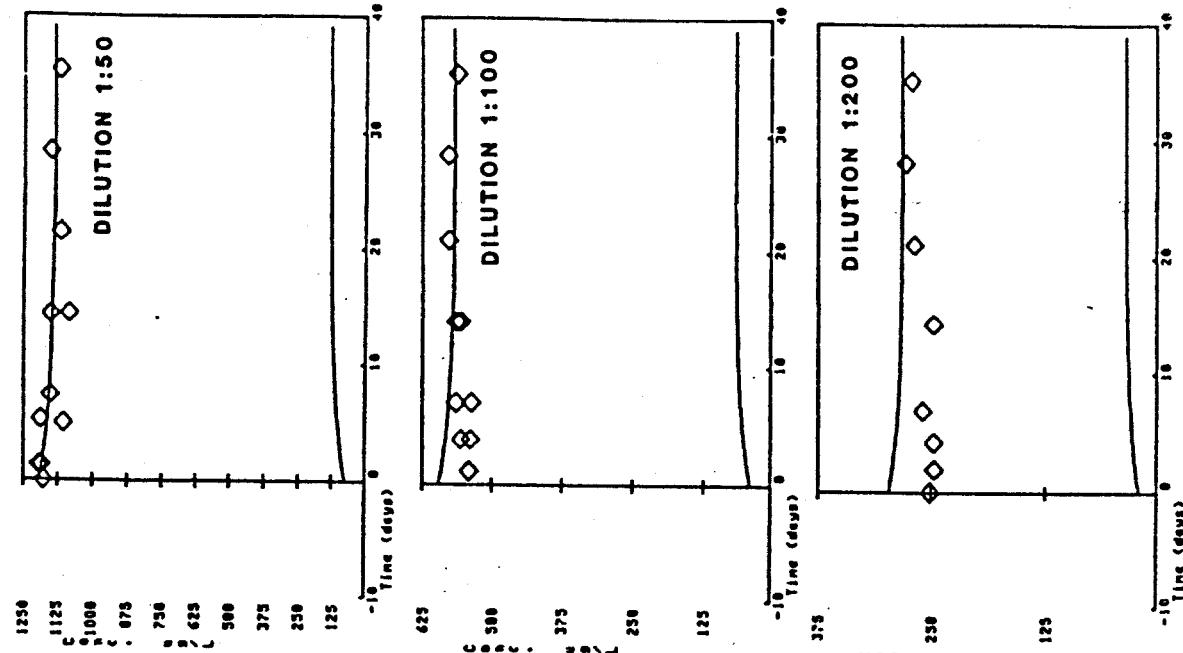
FIGURE CAPTIONS

1. Experimental data (Rohatgi and Chen, 1975) and the three component model fit for cadmium and chromium.
2. Total and dissolved copper concentrations versus time for Black Rock Harbor sediment and Bergen County sludge at 3000 mg/L (left side) and 300 mg/L (right side). Control reactor concentrations (C) are also shown.
3. Total and dissolved zinc (top) and cadmium (bottom) concentrations versus time for Black Rock Harbor sediment and Bergen County sludge at 3000 mg/L (left side) and 300 mg/L (right side).
4. Total and dissolved chromium concentrations versus time for Black Rock Harbor sediment and Bergen County sludge at 3000 mg/L (left side) and 300 mg/L (right side).
5. pH (top) and Total Organic Carbon (bottom) versus time for Black Rock Harbor sediment (left) and Bergen County sludge (right) reactors.
6. Anaerobic Reactor dissolved copper concentrations versus time for Black Rock Harbor (top left), Ridgewood (bottom left) and Bergen County (top right). Total organic carbon concentrations versus time for the three reactors are also shown (bottom right).
7. Sulfate (top) and excess sulfate (bottom) concentration versus time for Black Rock Harbor sediment.
8. Concentration of total and dissolved radioactive zinc versus time. Radioactive zinc is added at the start ($t=0$) of the experiment.
9. Concentration of total and dissolved radioactive cadmium versus time. Radioactive cadmium is added at the start ($t=0$) of the experiment.
10. Concentration of total and dissolved radioactive zinc at various times. Radioactive zinc is added to a subsample taken at the indicated time and a one hour - one hour adsorption-desorption measurement is made. The total and dissolved zinc at adsorption (top) and desorption (bottom) is shown for Black Rock Harbor (left) and Bergen County (right).
11. Concentration of total and dissolved radioactive cadmium at various times. Radioactive cadmium is added to a subsample taken at the indicated time and a one hour - one hour adsorption-desorption measurement is made. The total and dissolved cadmium at adsorption (top) and desorption (bottom) is shown for Black Rock Harbor (left) and Bergen County (right).

CADMIUM

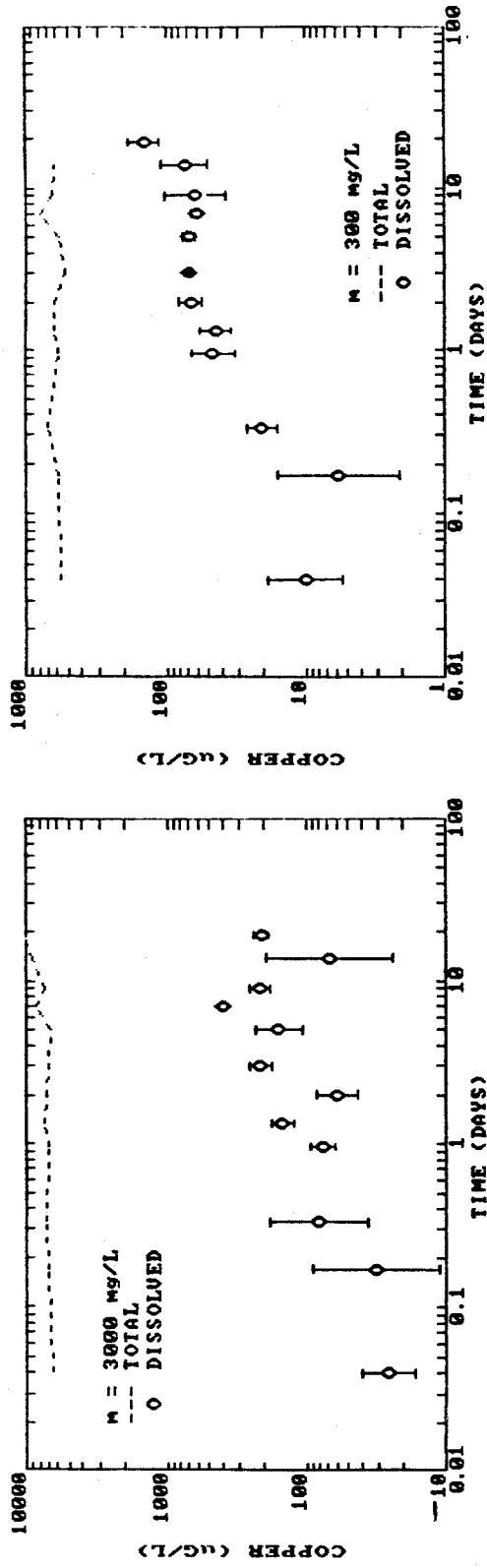


CHROMIUM

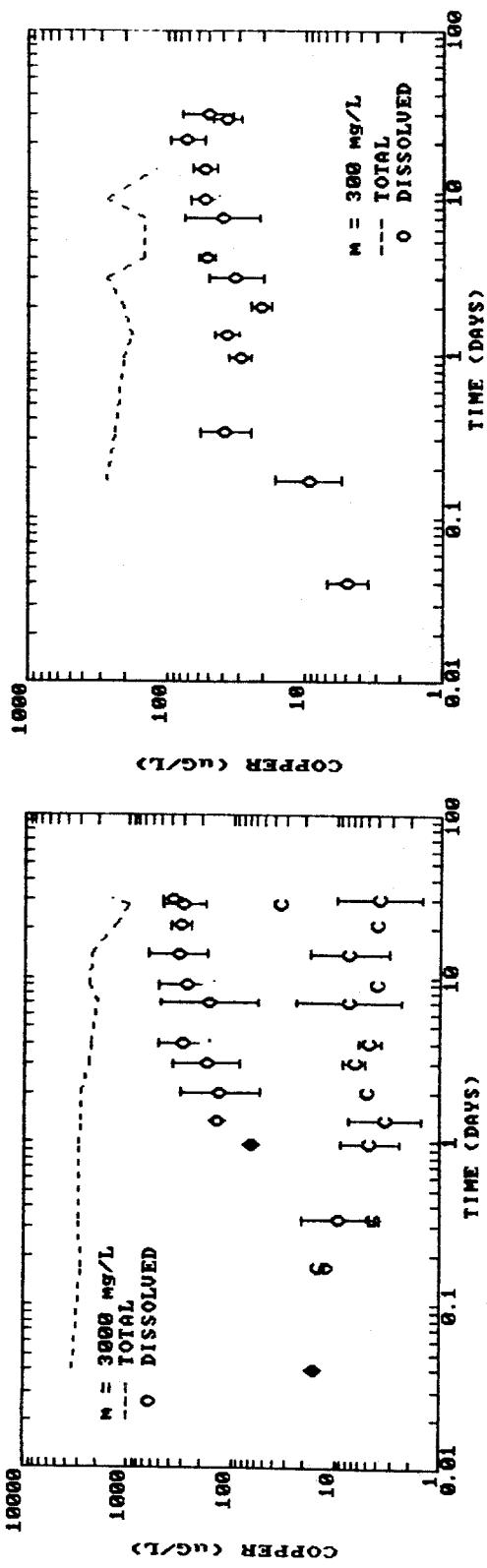


1. Experimental data (Rohatgi and Chen, 1975) and the three component model fit for cadmium and chromium.

BLACK ROCK HARBOR SEDIMENT

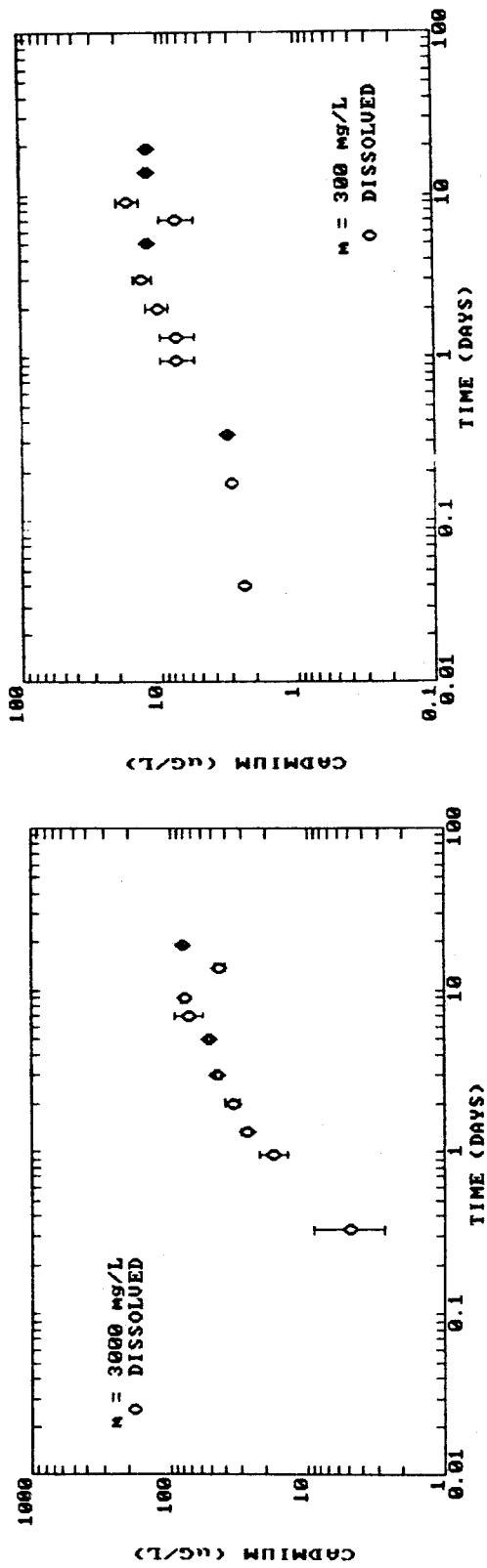
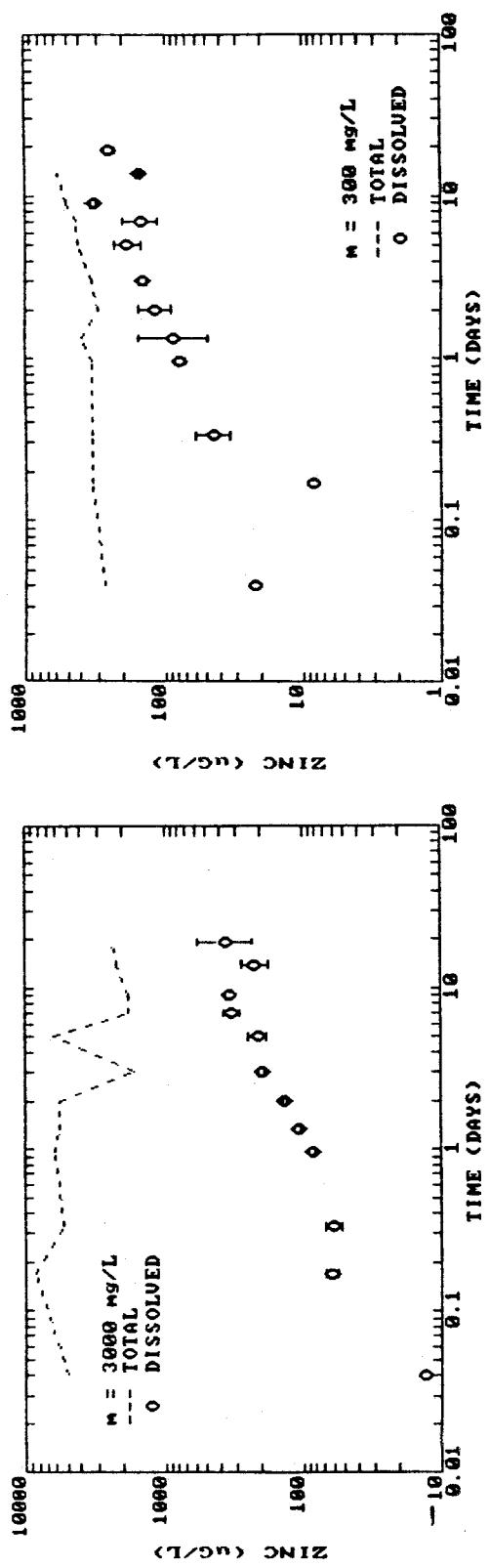


BERGEN COUNTY SLUDGE



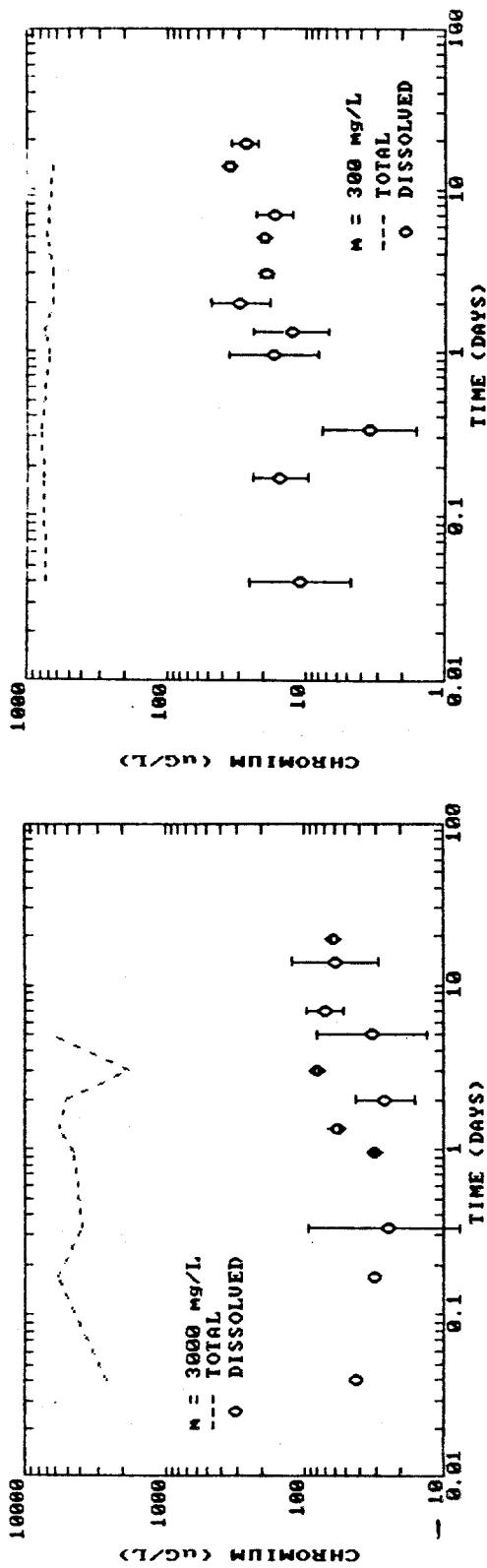
2. Total and dissolved copper concentrations versus time for Black Rock Harbor sediment and Bergen County sludge at 3000 mg/L (left side) and 300 mg/L (right side). Control reactor concentrations (C) are also shown.

BLACK ROCK HARBOR SEDIMENT

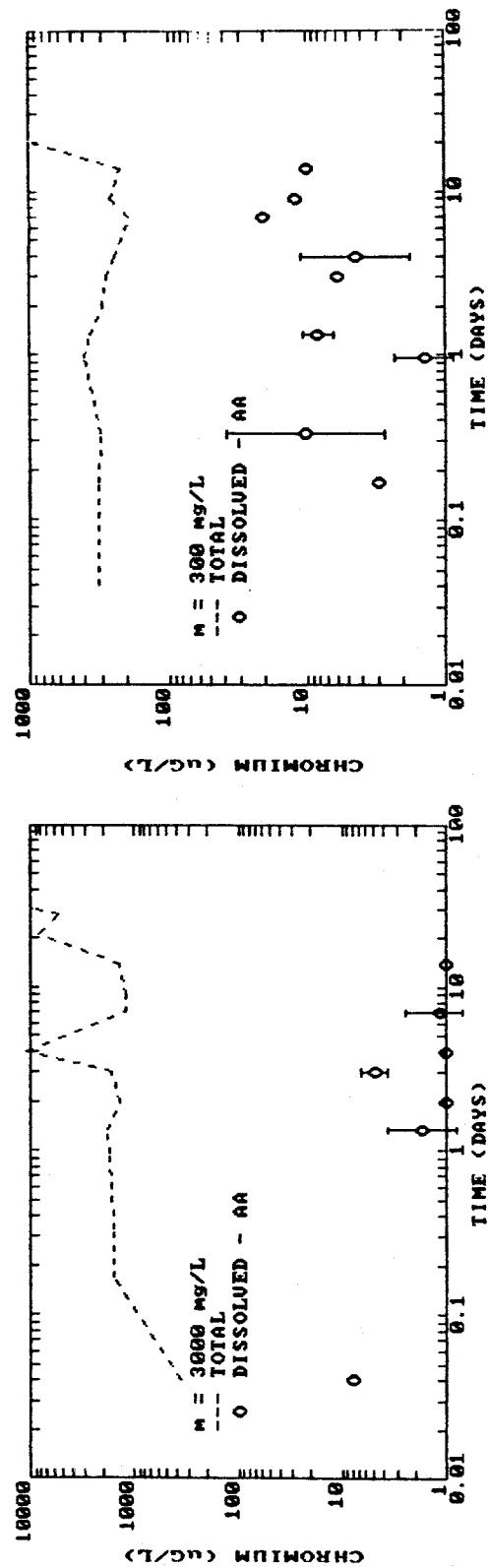


3. Total and dissolved zinc (top) and cadmium (bottom) concentrations versus time for Black Rock Harbor sediment and Bergen County sludge at 3000 mg/L (left side) and 300 mg/L (right side).

BLACK ROCK HARBOR SEDIMENT



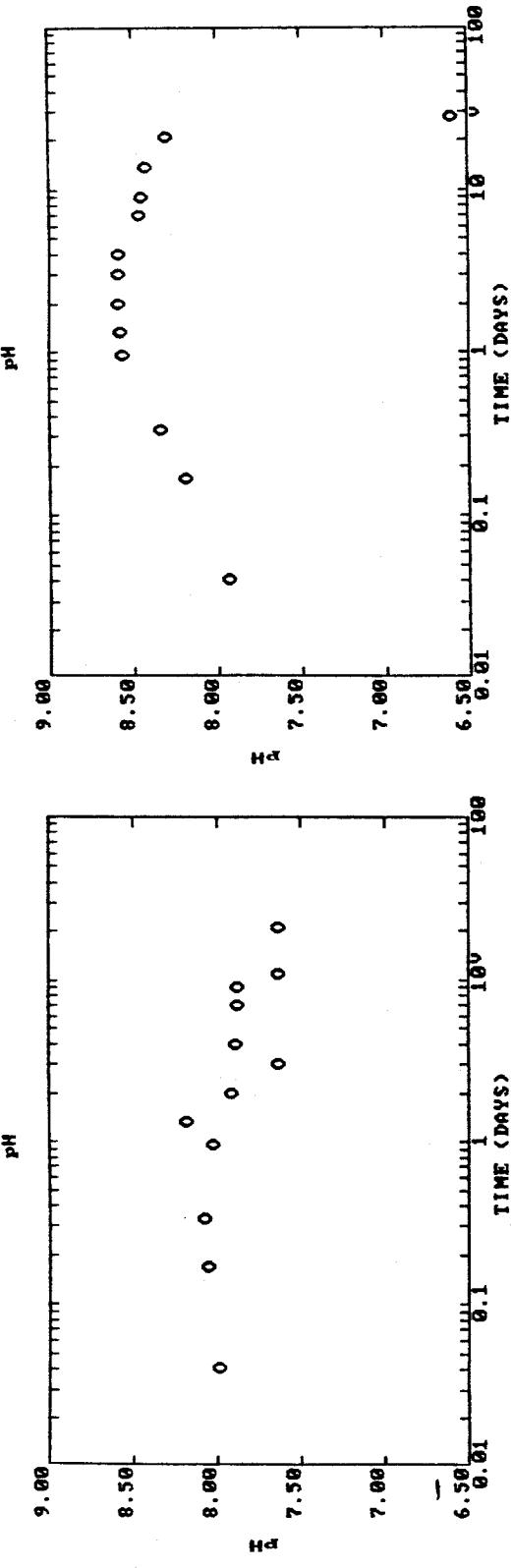
BERGEN COUNTY SLUDGE - #2



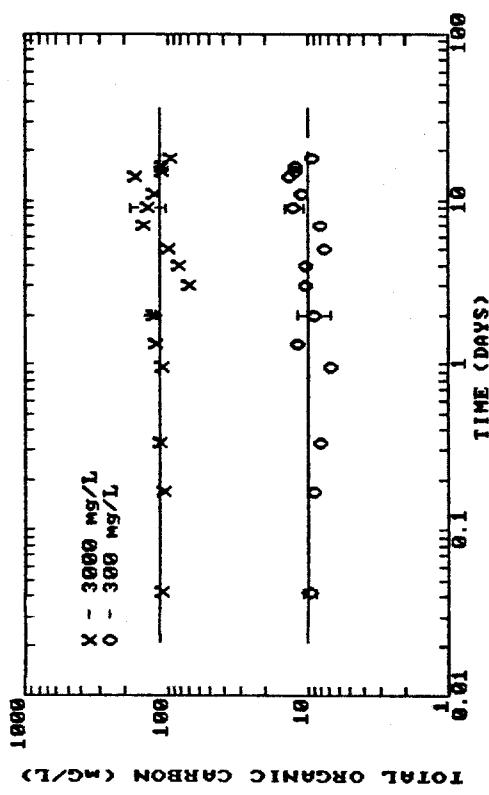
4. Total and dissolved chromium concentrations versus time for Black Rock Harbor sediment and Bergen County sludge at 3000 mg/L (left side) and 300 mg/L (right side).

BLACK ROCK HARBOR SEDIMENT - #4

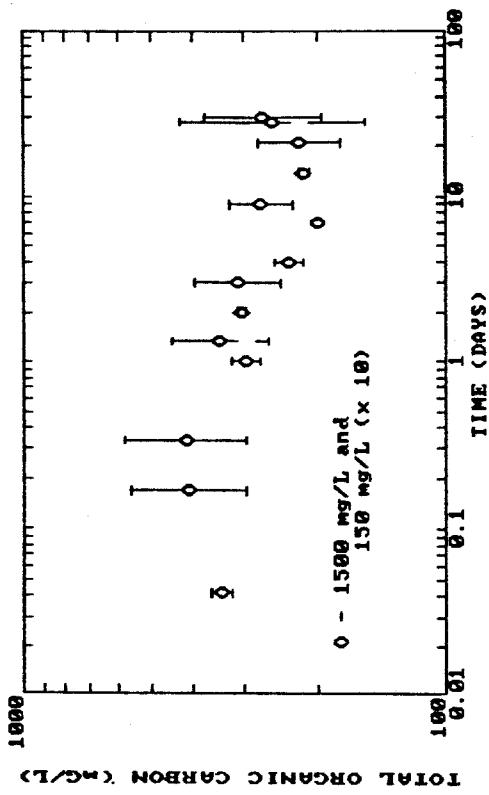
BERGEN COUNTY SLUDGE - #5



BLACK ROCK HARBOR SEDIMENT -- 6/85

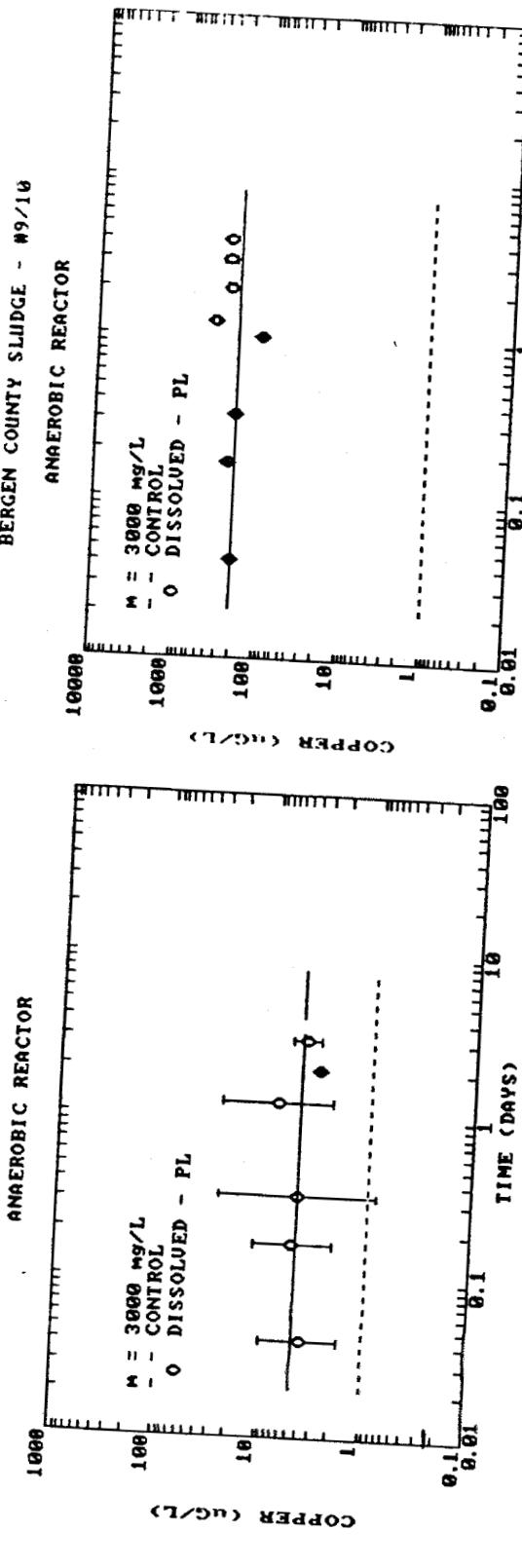


BERGEN COUNTY SLUDGE -- 7/85

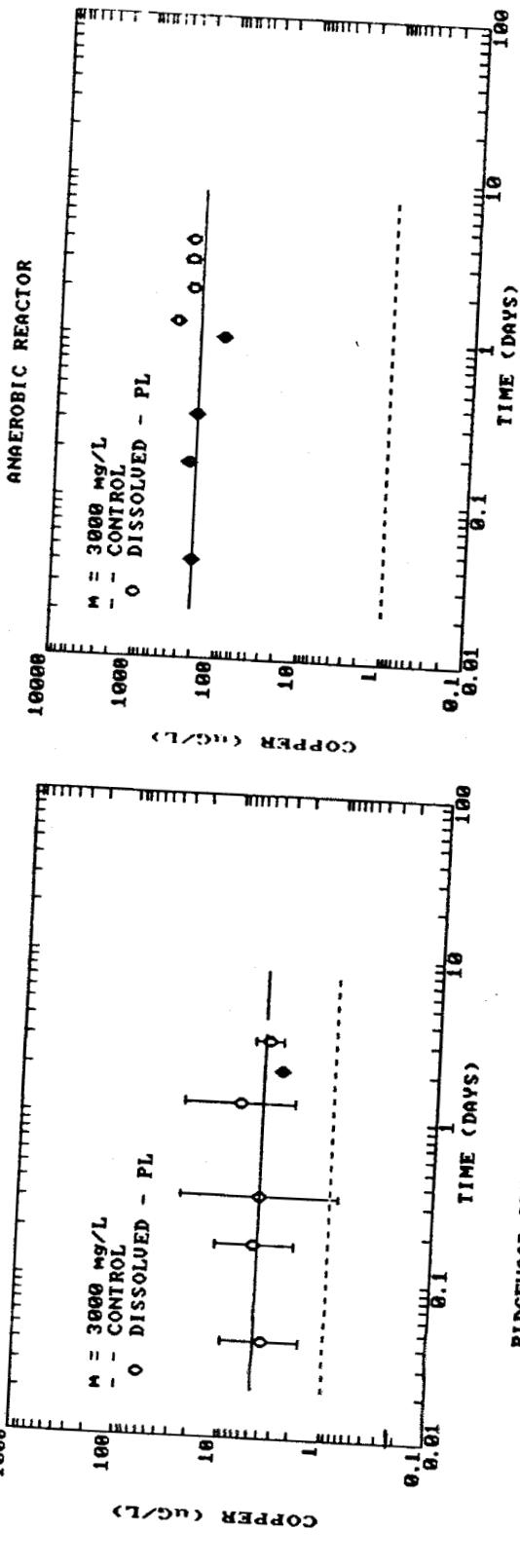


5. pH (top) and Total Organic Carbon (bottom) versus time for Black Rock Harbor sediment (left) and Bergen County sludge (right) reactors.

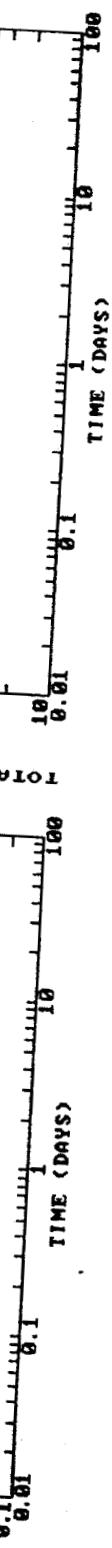
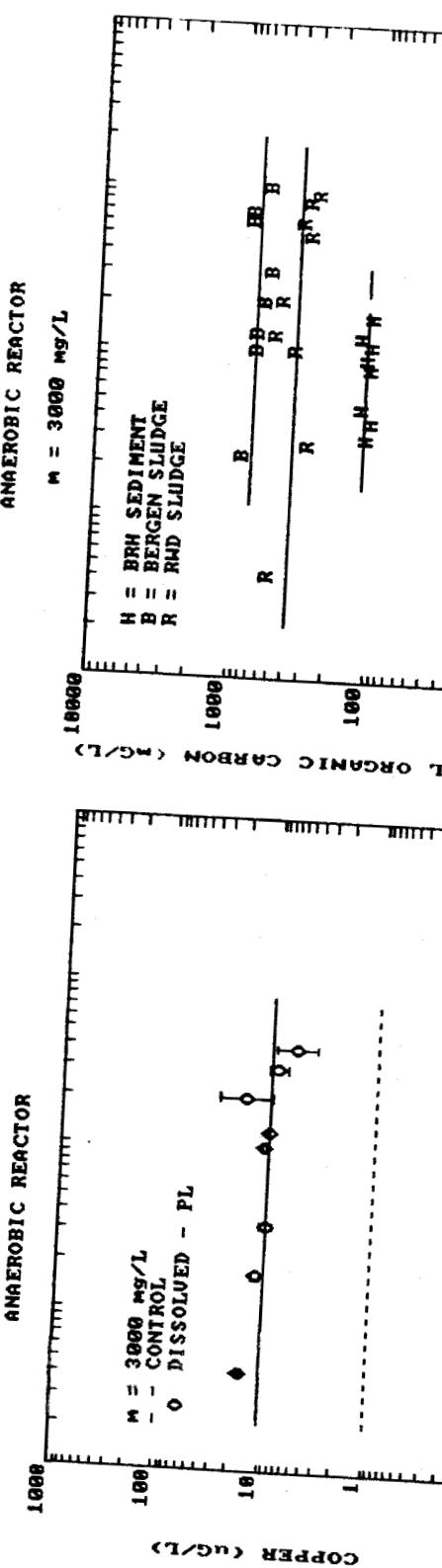
BLACK ROCK HARBOR SEDIMENT - #9/10



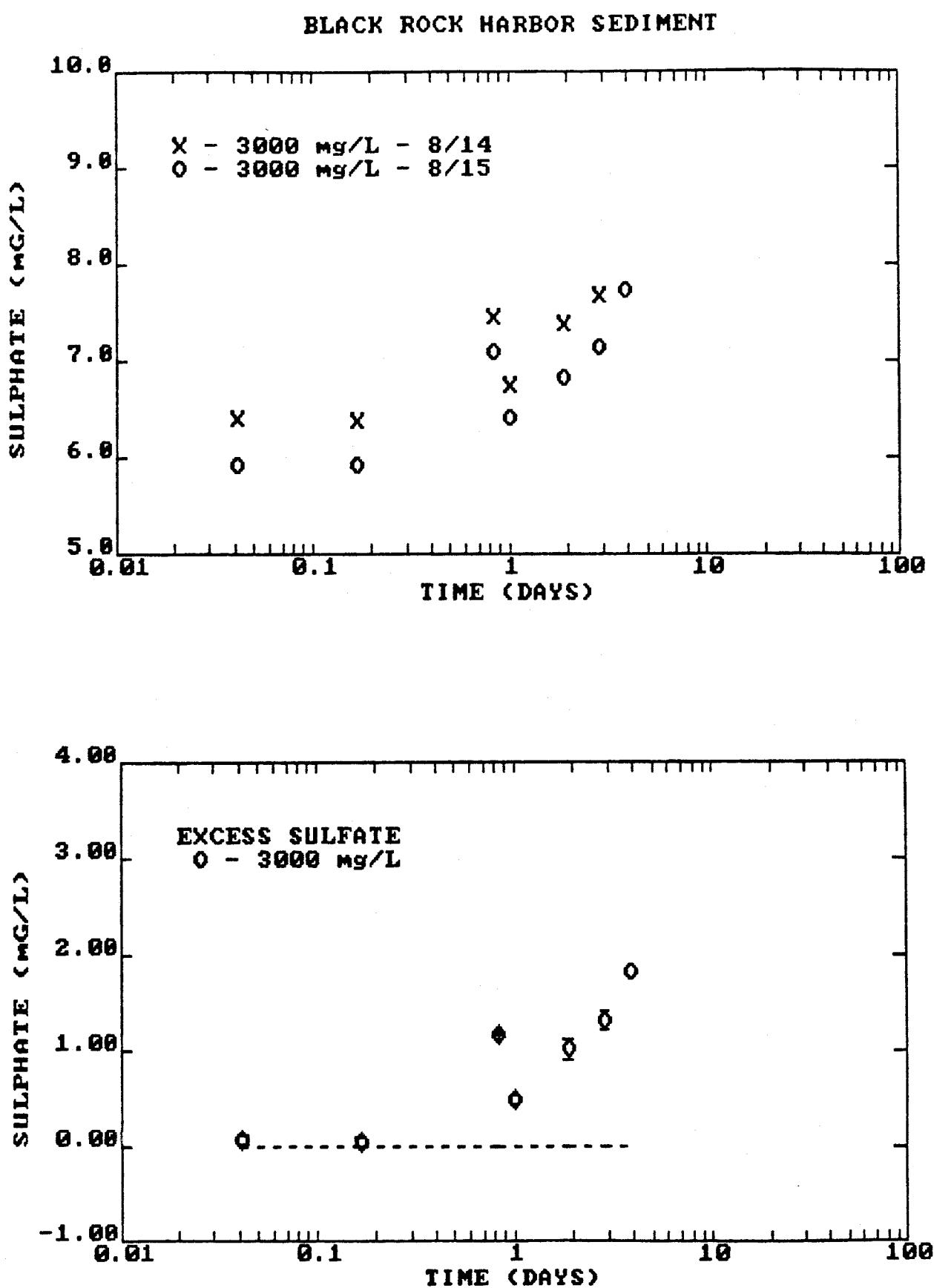
BERGEN COUNTY SLUDGE - #9/10



RIDGEWOOD SLUDGE - #9/10



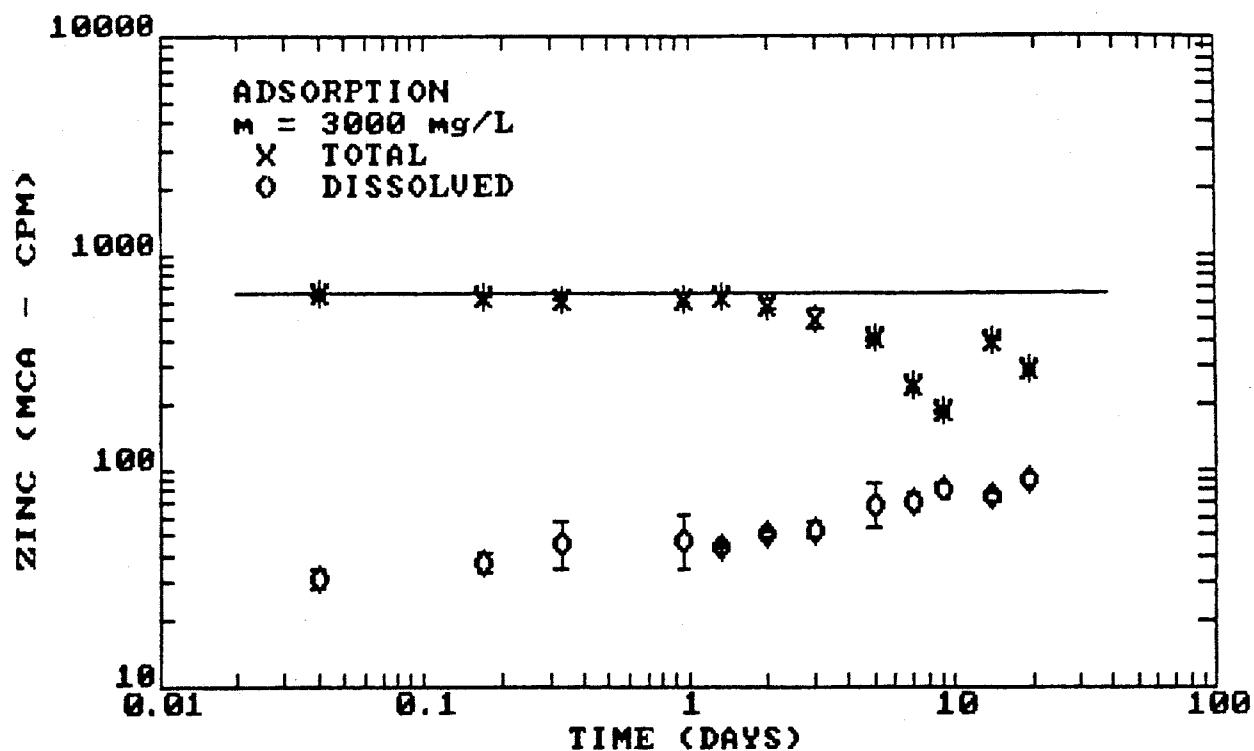
6. Anaerobic Reactor dissolved copper concentrations versus time for Black Rock Harbor (top left), Ridgewood (bottom left) and Bergen County (top right). Total organic carbon concentrations versus time for the three reactors are also shown (bottom right).



7. Sulfate (top) and excess sulfate (bottom) concentration versus time for Black Rock Harbor sediment.

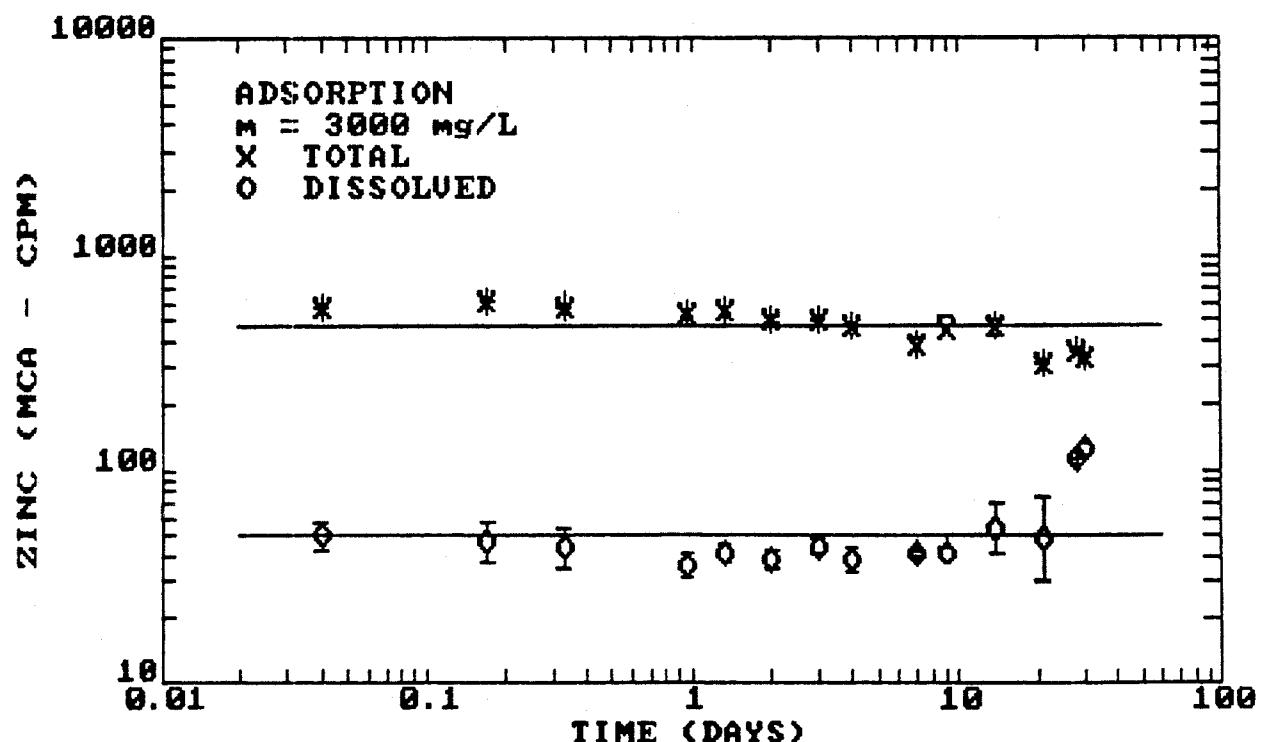
BLACK ROCK HARBOR SEDIMENT - #2

PARTITION COEFFICIENT KINETICS



BERGEN COUNTY SLUDGE - #2

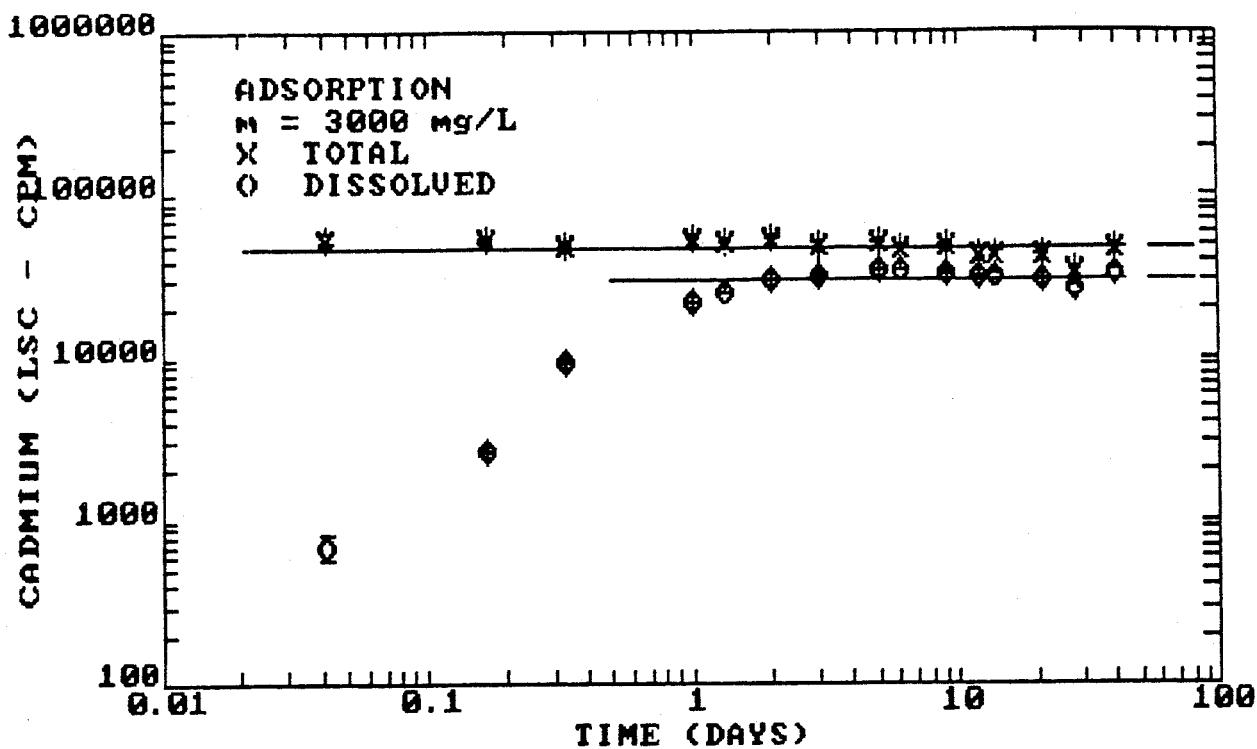
PARTITION COEFFICIENT KINETICS



8. Concentration of total and dissolved radioactive zinc versus time. Radioactive zinc is added at the start ($t=0$) of the experiment.

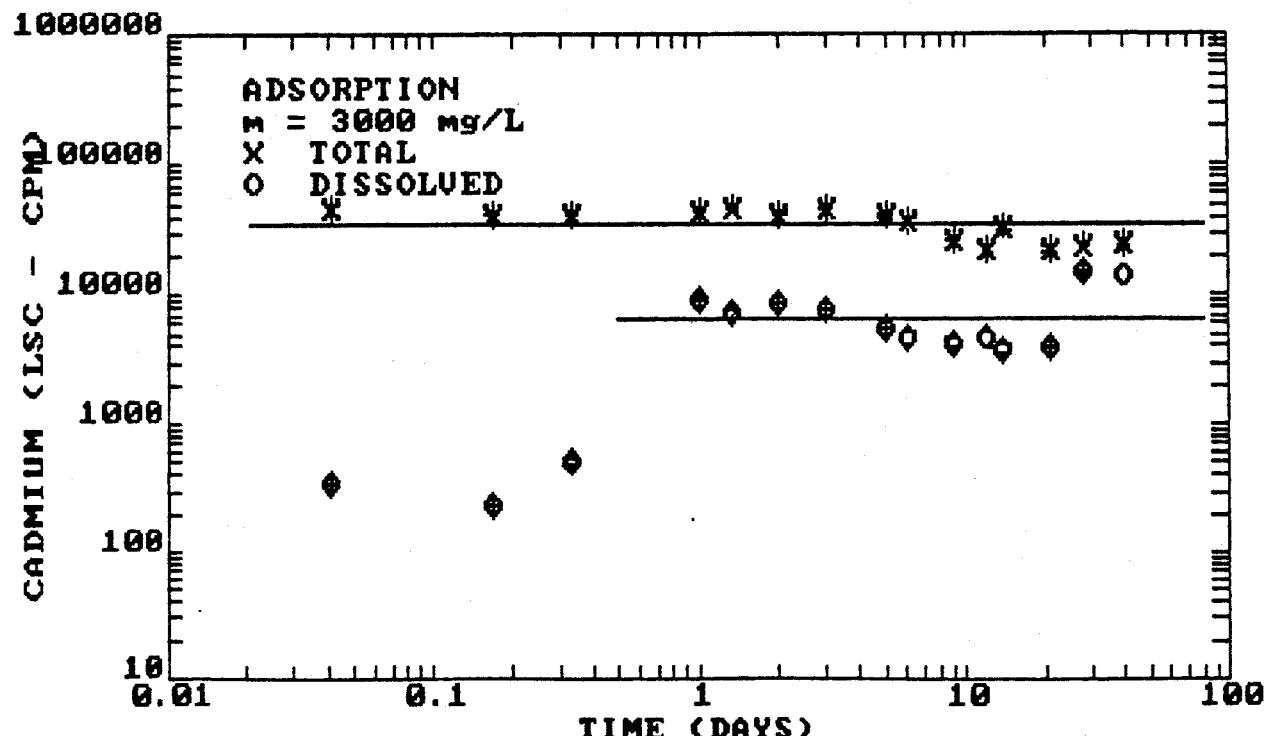
BLACK ROCK HARBOR SEDIMENT - #6

PARTITION COEFFICIENT KINETICS



BERGEN COUNTY SLUDGE - #7

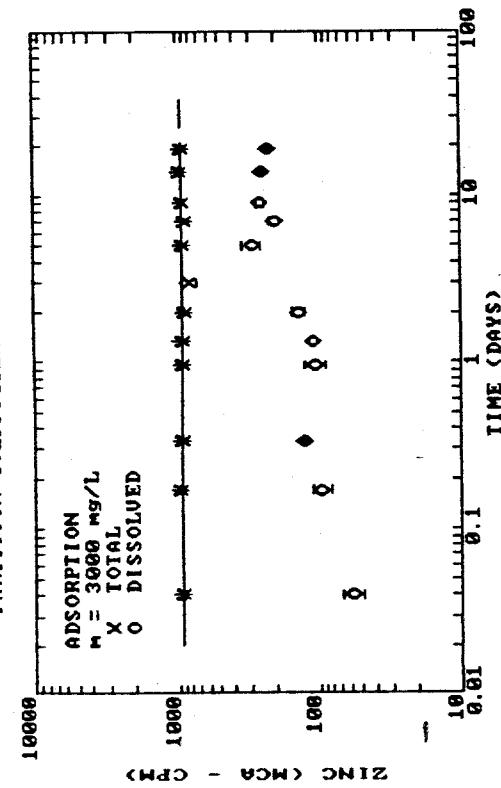
PARTITION COEFFICIENT KINETICS



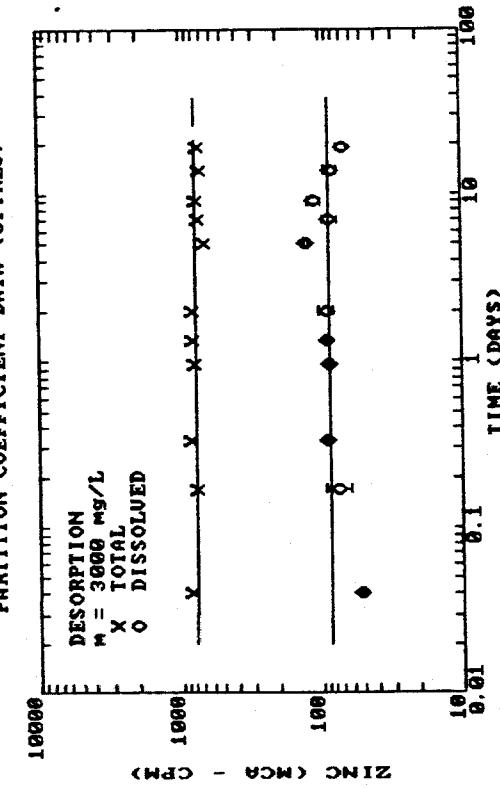
9. Concentration of total and dissolved radioactive cadmium versus time. Radioactive cadmium is added at the start ($t=0$) of the experiment.

BLACK ROCK HARBOR SEDIMENT - #2

PARTITION COEFFICIENT DATA (SPIKES)

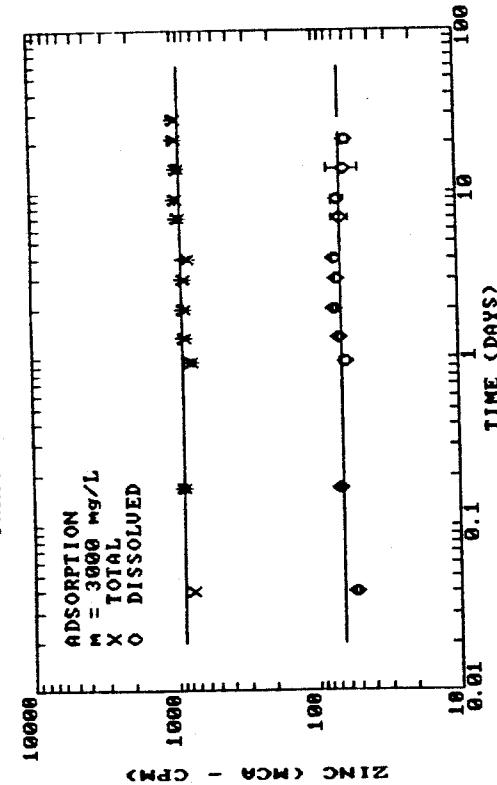


BLACK ROCK HARBOR SEDIMENT - #2
PARTITION COEFFICIENT DATA (SPIKES)

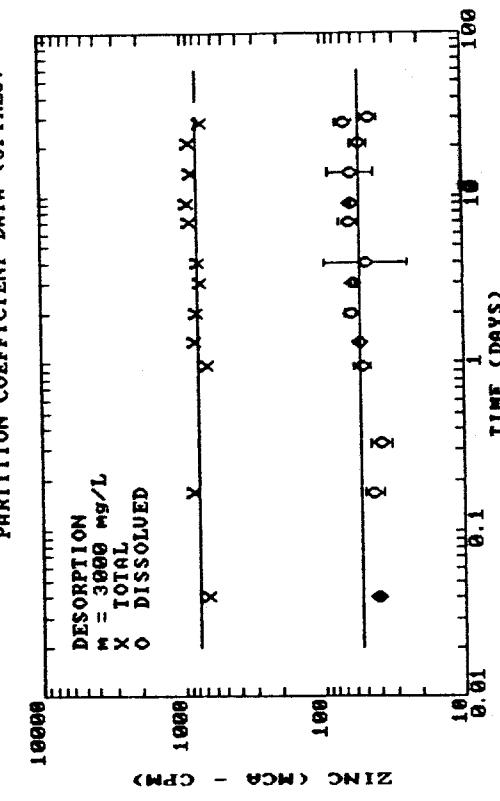


BERGEN COUNTY SLUDGE - #2

PARTITION COEFFICIENT DATA (SPIKES)



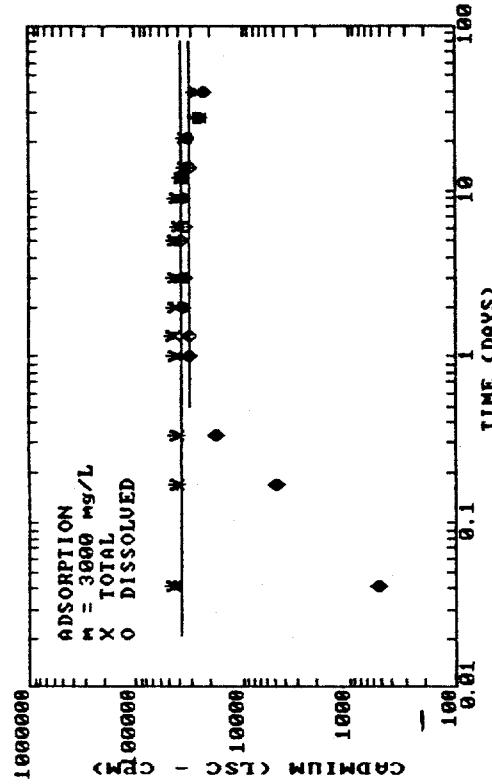
BERGEN COUNTY SLUDGE - #2
PARTITION COEFFICIENT DATA (SPIKES)



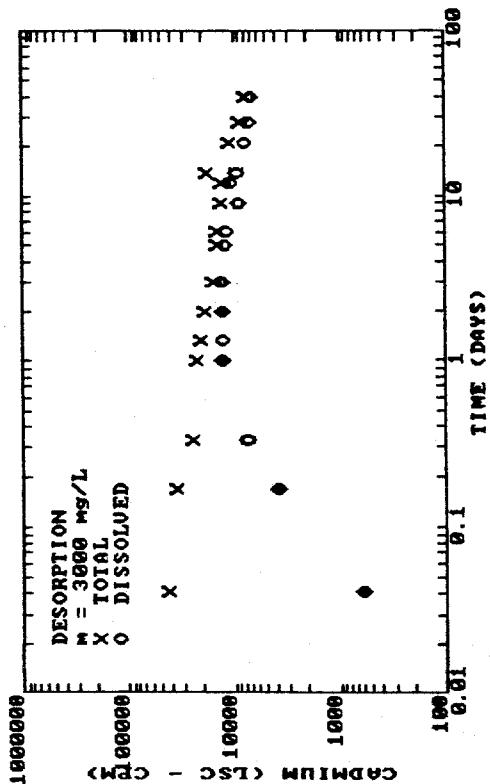
10. Concentration of total and dissolved radioactive zinc at various times. Radioactive zinc is added to a subsample taken at the indicated time and a one hour - one hour adsorption-desorption measurement is made. The total and dissolved zinc at adsorption (top) and desorption (bottom) is shown for Black Rock Harbor (left) and Bergen County (right).

BLACK ROCK HARBOR SEDIMENT - #6

PARTITION COEFFICIENT DATA (SPIKES)

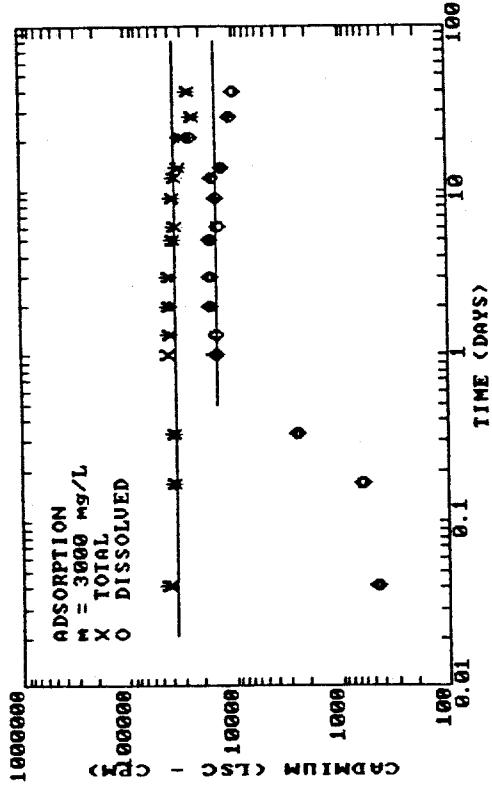


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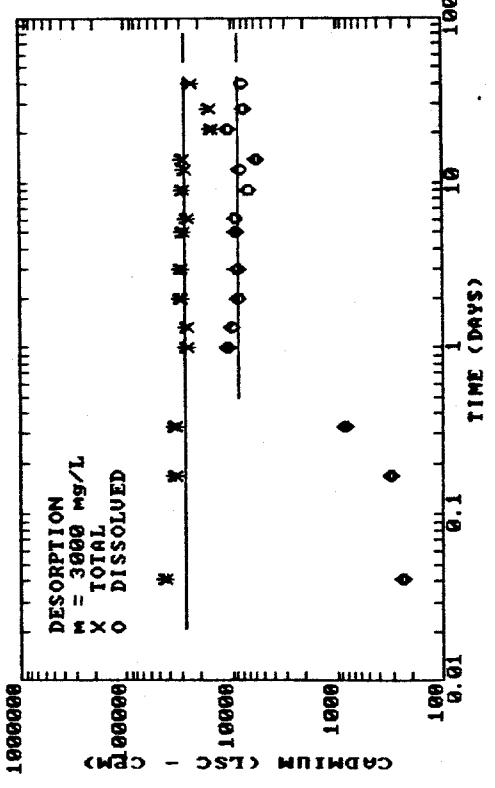


BERGEN COUNTY SLUDGE - #7

PARTITION COEFFICIENT DATA (SPIKES)



BERGEN COUNTY SLUDGE - #7
PARTITION COEFFICIENT DATA (SPIKES)



11. Concentration of total and dissolved radioactive cadmium at various times. Radioactive cadmium is added to a subsample taken at the indicated time and a one hour - one hour adsorption-desorption measurement is made. The total and dissolved cadmium at adsorption (top) and desorption (bottom) is shown for Black Rock Harbor (left) and Bergen County (right).